

Stimuli-induced Organ-specific Injury Enhancement of Organotropic Metastasis in a Spatiotemporal Regulation

Dongwei Gao · Sha Li

Received: 19 April 2013 / Accepted: 5 December 2013 / Published online: 20 December 2013
© Arányi Lajos Foundation 2013

Abstract The relationship between inflammation and tumorigenesis has been established. Recently, inflammation is also reported to be a drive force for cancer metastasis. Further evidences show that various stimuli directly induced-injury in a specific organ can also promote metastasis in this organ, which include epidemiological reports, clinical series and experimental studies. Each type of cancer has preferential sites for metastasis, which is also due to inflammatory factors that are released by primary cancer to act on these sites and indirectly induce injuries on them. Host factors such as stress, fever can also influence distant metastasis in a specific site through stimulation of immune and inflammatory effects. The five aspects support an idea that specific-organ injury directly induced by various stimuli or indirectly induced by primary tumor or host factors activation of proinflammatory modulators can promote metastasis in this organ through a spatiotemporal regulation, which has important implications for personalized prediction, prevention and management of cancer metastasis.

Keywords Stimuli · Organ-specific injury · Organotropic metastasis · Spatiotemporal regulation

Introduction

Metastasis is the primary cause of mortality from most solid cancers. Understanding the biology of metastasis will be vital

for discovering new targets to prevent/slow and treat metastasis. Although great advancements have been made in uncovering cancer metastasis, it still remains a great problem for cancer patients. The relationship between chronic inflammation and tumorigenesis has long been recognized [1–5]. Recently, inflammation is also found to play an important role in the process of metastasis [6–8]. According to this clue, we search for the literatures about the studies focusing on inflammation and metastasis and find that enhancement of cancer metastasis in a specific organ is not only due to chronic inflammation but also link with a variety of injuries on this organ induced by different stimuli. This can be date back to Jones's [9] study in 1914, who showed that secondary tumors grew at points of injury. To get more information on specific-organ injury and organotropism metastasis, we further enlarged the searching strategies by using keywords including inflammation, injury, infection, wound healing, trauma, asthma, fibrosis, bone fracture, skin scar, and metastasis from the databases of the Google scholar and PubMed. We also conducted a secondary searching from these literatures. These searched literatures supported an idea that there was a strong relationship between different stimulus-induced specific organ injury and tissue-specific metastasis. In the following statements, we reviewed the evidence for this idea and discussed its implications in the biology of metastasis and clinical practice.

Evidences from Epidemiological Studies, Clinical Series and Experimental Studies

Evidences from Epidemiological Studies

Epidemiological studies show that different stimuli such as smoking and asthma-induced lung injury can enhance lung metastases from different kinds of cancers. Smoking is a well

D. Gao
536 Hospital of PLA, 29# Xiadu street, Xining 810007,
Qinghai Province, People's Republic of China

D. Gao (✉) · S. Li
Department of Radiation Oncology, Lanzhou General Hospital of
PLA, 333# Southern Binhe Road, Lanzhou 730050, Gansu,
Province, People's Republic of China
e-mail: gdw3152007@hotmail.com

known risk factor for carcinogenesis, especially for lung cancer [10,11]. It can also predict poor prognosis in breast and other cancers [12,13], which may be due to short-term relapse in smoking patients. Shaw et al. [14] found that smoking could induce malignant melanoma metastasis, indicating that smoking not only contributed to carcinogenesis but also metastasis. Recently, it has been testified that smoking can increase lung metastasis from breast and esophageal cancer [15–17]. Smoking-induced lung injury is mainly through increasing the permeability of endothelial cells and assaulting epithelial cells by free oxygen radicals, local immune functions on pulmonary and system effect [10,18–21]. These effects are considered to contribute to enhancement of lung metastasis [15–17]. Padua et al.'s [22] showed that the mechanism of lung metastasis from breast cancer was attributed to the increasing permeability of lung capillaries caused by angiopoietin-like 4 (ANGPTL4). Smoking seems to have a similar effect to ANGPTL4 on increasing the permeability of lung capillaries by its compositions. Asthma is a common pulmonary disease with infiltration of perivascular/peribronchial leukocyte cells, subepithelial fibrosis, increasing mass of airway smooth muscle and secretion of cytokines [23,24]. Among these changes, fibroblasts, macrophage cells, transform growth factor-beta (TGF- β) have been also identified to play important roles in cancer metastasis [25–33]. So Taranova et al. [34] studied the relationship of asthma and lung metastasis from breast cancer and found that breast cancer patients with asthma underwent a high risk of lung metastasis compared with non-asthma patients, which might be due to the pulmonary changes at the cellular and molecular levels caused by asthma. Radiation is a local treatment for breast cancer. However, it can also induce side effects including skin and lung injuries [35]. Unfortunately, it was found that post-operative radiation could also enhance skin and pulmonary metastasis from breast cancer [36]. The lung damage from smoking and radiation is mainly a kind of injury not just inflammation while asthma-induced lung injury manifests itself as a process of airway remodeling and inflammation, which are regarded as different stimuli to induce lung injury through different mechanisms. All of them can enhance lung metastases from the same type of cancer. In this sense, smoking, radiation and asthma seem to have a common status in lung for recruitment of the same type of cancer cells. They have been reported in many literatures to induce structural and functional changes in the lungs, which include injured cells, actively inflammatory cells and their releasing cytokines, chemokines and growth factors. These changes are reported to enhance lung metastasis through the mechanisms of passively enriching cancer cells by structural changes of the lung and actively binding their receptors by cytokines, chemokines or growth factors in the above literatures, which may contribute to the common status. Identify this common status will be a direction to prevent lung metastasis. On the other hand, the

same stimulus-induced lung injury can increase lung metastases from different types of cancer, which mean that there is also a common pathway in different cancers. This common pathway can be recognized by the constitutions of microenvironment of the lung injury and is important to develop common target drugs for different cancers.

Other inflammatory sites have been also reported to be prone for cancer metastasis. Peritoneal metastasis is a frequent pattern of recurrence for many solid tumors. The relationship between peritoneal injury, repair and peritoneal metastasis has been recognized [37]. Pancreatic fistula (PF) is a common complication after biopsy or resection of pancreatic cancer. It was reported that PF after surgery of pancreatic cancer was a predictive factor for peritoneal recurrence [38]. PF can induce inflammation around the pancreatic anastomosis, which may be the reason for PF prediction of peritoneal recurrence. Oral metastasis is a rare event. A literature analysis showed that metastatic tumors were more often seen at the gingival than other oral mucosal sites [39]. In the oral cavity, the gingiva is prone to be affected by inflammation. This finding may indicate that the chronically inflamed gingiva may serve as a pre-metastatic niche for disseminated cancer cells [39].

Although these clinical studies mainly focused on pulmonary, peritoneal and gingiva metastasis, the following case reports showed that metastases occurred at the sites with injury, trauma, wound infection and chronic inflammation and supported the association between stimuli-induced organ-specific injury and organotropic metastasis.

Evidences from Clinical Series

The injured tissues have been recognized to be preferential sites of metastases since 1960s [36,40–42]. However, there are only case reports in clinical observations. These reports showed that metastases in the skin scars [42–56] such as the flap donor site, radiation dermatitis, unrelated biopsy site [57] and percutaneous gastrostomy sites [58,59], bone traumas and fractures [41,60–66], pulmonary trauma [67], lung fibrosis [68], focal infarction and infection of the brain [69,70], which suggested that injuries in the different sites might favor an environment for tumor locomotion and growth. Unusual metastases should be also noticed. Skeletal muscle metastasis is an infrequent event. Palazzo et al. [71] and Magee et al. [72] respectively reported skeletal muscle metastases mainly at the injury sites of skeletal muscles, indicating that injury and inflammatory oncotaxis contributed to metastasis at this tissue. Synovial metastasis is also very rare. Currall et al. [73] reported a case with synovial metastasis from colorectal adenocarcinoma after total knee arthroplasty, indicating the injury and repair of synovial tissue after operation provided a favorable environment for cancer cells. Dionigi et al. [74] reported a case that was diagnosed of intrathyroidal metastasis from renal clear cell

carcinoma. The patient had been evaluated toxic substernal goiter with chronic thyroiditis for 2 years before diagnosed of renal carcinoma, indicating that chronic thyroiditis could summon up cancer cells to lodge and favor them growth. Table 1 sum up some cases of the above reports. In these reports, some patients showed metastases at the injured sites before diagnosed of primary tumors; others exhibited metastases after resection of primary tumors. They have a common trait that inflammatory sites promote metastases even if corresponding primary tumors rarely metastasize to the organs, which support that different wounds can increase metastases in these inflammatory sites [75]. Walter [62] has proposed two possibilities to explain the mechanism. One is that locally damage tissues create a favorable environment that is permissive for seeding of metastatic cells from distant sites. The other is that micrometastatic foci are already present at the time of injured sites, and trauma initiates changes in the micro-environment that stimulate the proliferation of the metastatic cells. We support that both of them exist. Inflammatory sites can summon cancer cells and or stimulate the preexisting micrometastases at these sites. In clinical practice, doctors should be aware of these circumstances that internal and external inflammation sites may be possible focuses for tumor metastases. These literatures are only case reports and seem to be not enough for supporting this hypothesis. The probable reason may be that similar cases are ignored by clinical doctors or due to publication bias. However, the following experimental studies may further provide evidence for this idea.

Evidences from Experiment Studies

Researchers have investigated whether metastasis preference can be influenced by an injured organ. They have found that injuries on lung, liver, bone, kidney, and other organs can promote occurrences of metastasis in these wound organs no matter in artificial metastasis or spontaneous metastasis. This effect mainly occurs during the healing process. When these injured organs recover to normal status or develop to fibrosis, metastases will rarely occur. This supports that a particular organ undergoing a healing process after stimuli action may promote organ-specific metastasis through a spatiotemporal regulation.

The lung is an organ affected by both primary lung cancers and metastatic tumors from other solid tumors. Chronic inflammation has been testified to be strongly related with primary lung cancer [11]. The following experiments also show that different stimuli-induced lung injury can increase the formation of metastasis in the site. Smoking is a well known factor to induce lung injury. The retrospective reports showed that smoking increased lung metastasis from breast cancer [15,16], which was also testified in Murin et al.'s

experiment [76]. The potential mechanism may be related to smoking-associated inflammatory responses including local pulmonary changes and system effects. Protein kinase C (PKC) in activation status is a mediator to induce inflammatory response and cell death in human airway epithelial cells [77]. Gopalakrishna et al. [78] found that catechol and hydroquinone released by cigarette smoke could activate PKC to influence cancer cell invasion and metastasis, which might be the reason for smoking promotion of lung metastasis. Platelet hyperactivity is one of smoking induced system effects [79] and can lead to release aggregation-inducing factor aggrus/podoplanin that has been shown to promote pulmonary metastasis [80]. A lung infection is inflammation in the lungs caused by a virus, fungi or bacteria infection. It is present in about 47 % of cancer patients and often leads them to be death [81]. It has been found that acute lung infections can also promote pulmonary metastasis, which may be due to infections-induced lung inflammation that provides a favorable metastatic niche [81]. Yan and colleagues further find that the mechanistic basis for bacterial-induced lung injury promoting pulmonary metastasis was via the ubiquitin/CXCR4 chemokine axis [81]. The Chemokine CXCR4 has been found to play a critical role in lung metastasis from breast cancer [82], which can be modulated by multiple inflammatory components [83]. Cyclophosphamide is a cytotoxic drug to treat most of solid cancers. Nevertheless, it also has an opposite effect and is found to be able to enhance lung metastasis in experimental studies [84–89]. Multiple mechanisms are involved in the process, which include endothelial cell injuries, elevated vascular endothelial growth factor (VEGF), increased immunoreactivity, and macrophage infiltration [86,88,89]. Another cytotoxic drug, bleomycin is well known for inducing lung injury. Its damages on the lung include alveolar damage, collagen synthesis, macrophage infiltration, injuries of vascular endothelial cells and secretion of cytokines such as TGF- β , tumor necrosis factor alpha (TNF- α), connective tissue growth factor(CTGF) [90–92]. These responses are also reported to be vital for the formation of metastasis [25,26,32,33,93–99]. Thus, the investigations on the relationship between lung metastasis and bleomycin-induced pulmonary injury were also studied and showed that bleomycin-induced pulmonary damage increased lung metastasis [100–103]. Radiation-induced lung injury has been well known. It can also increase lung metastasis in preirradiation of lung of experiment animals [87,104–107]. This effect can be abolished by neutralizing antibody of TGF- β , indicating that the effect is partly through the regulation of TGF- β [108]. Other stimuli such as monocrotaline [109], carbon suspension [110], hyperoxia [100,111], ozone exposure [112] like smoking or radiation have different mechanisms to induce lung injury but have the same effect on enhancing lung metastasis, which is not only related to the status of lung damage but also the process of repair in these reports.

Table 1 Clinical reports of metastasis at the injured sites

Author(Year)	Primary cancer	Diagnostic methods for primary cancer	Metastatic sites and their traits	Interval between primary and metastasis
Cohen (1972) [42]	larynx cancer	Occasionally found PT in anesthesia	skin, bone, and lymph nodes in previously traumatized area	5 months
Marley (1982) [45]	Uterus adenocarcinoma	Routine diagnose of PT	Skin in the area of chronic radiation dermatitis	NA
Ito (1984) [46]	Breast cancer	Routine diagnose of PT	Skin in the areas previously treated by prophylactic irradiation	6 months
Carr (1986) [47]	the retromolar fossa	Routine diagnose of PT	the donor site of a temporalis muscle flap	NA
Betke (1993) [49]	gastric carcinoma	Routine diagnose of PT	a congenital melanocytic nevus	NA
Erol (2008) [53]	cutaneous melanoma	Routine diagnose of PT	skin graft donor site on the contralateral lower extremity	8 weeks after PT excision
Pradhan (2006) [52]	Cervix squamous cell carcinoma	Routine diagnose of PT	skin scar of a previous cesarean section before 26 years	12 months
Marenco (2009) [54]	Melanoma on the right supraclavicular	Routine diagnose of PT	Skin graft donor site at the left thigh	1 month
Shine (1981) [44]	Sigmoid colon adenocarcinoma	Routine diagnose of PT	Metastasis at the site of intradermal skin tests with a marked inflammatory response after operation of PT	3 months after PT and 2 months after skin test
Trefzer (1998) [50]	malignant melanoma on the left scapula (Clark level IV)	Routine diagnose of PT	At the graft donor site with a healing process	8 weeks after removal of PT and receiving grafts
Serrano-Ortega (2000) [51]	Acral lentiginous melanoma (ClarkLevel III, Breslow depth 1.3 mm)	Routine diagnose of PT	At the graft donor site with a healing process	1 month after removal of PT and receiving grafts
Ferguson (2004) [57]	an osteosarcoma of the proximal humerus	Routine diagnose of PT and concurrent with biopsy of the contralateral humerus	At the Remote Biopsy Site with a healing process	NA
Hameed (2009) [59]	squamous cell carcinoma of the oropharynx	Routine diagnose of PT and receiving percutaneous endoscopic gastrostomy	the gastrostomy site with granulation tissue	4 months
El Saghir N (2005) [60]	non small-cell lung cancer	Routine diagnose of PT	At the right lateral side of his head suffering from an accidental trauma	16 month after PT
Fukushima (2010) [61]	Liver cancer	Routine diagnose of PT	At a skull fracture after 2 years diagnose of PT	3 months after trauma and 27 months after diagnose of PT
Walter (2011) [62]	Lung cancer	Lung cancer was diagnosed after trauma	at sites of recent physical trauma	Metastatic tumors firstly showed symptoms
Bergqvist (1978) [63]	Gastric cancer	PT was diagnosed six months after metastasis was found	Calcaneal metastasis at the fractured bone 2 years ago	Metastasis was firstly found the onset of gastric symptoms
Nielsen (1983) [69]	Cervix adenosquamous carcinoma	Routine diagnose and treatment of PT	Brain metastasis at the area of a 4-week-old infarct	10 months
Kukita (1992) [68]	cholangiocellular carcinoma	Pulmonary metastasis was firstly found	At the lungs with marked fibrotic changes	NA
Palazzo (2000) [71]	Colonic carcinoma	Routine diagnose and treatment of PT	At the skeletal muscle and skin with inflammatory oncotaxis	NA
Magee (2002) [72]	8 cases	NA	skeletal metastases at the sites with a documented clinical history of previous trauma	average 28 months after the trauma
Currall (2008) [73]	Rectosigmoid adenocarcinoma	The patient was firstly diagnosed of a metastatic disease after a left total knee arthroplasty.	At the synovial with wound repair after surgery	Metastasis prior to primary tumor
Dionigi (2008) [74]	renal clear cell carcinoma	The patient was also diagnosed of a toxic substernal goiter when PT was diagnosed.	At the thyroid gland with chronic thyroiditis	2 years after PT

However, among these studies, researchers mainly discovered the effect of promoting metastasis by different stimuli-induced lung injury and seldom explored the precise mechanism. Several studies further made endeavor to explore which factors exerted effect in the process. Three reports showed that endothelial injury induced by hyperoxic through neutrophil action contributed to the formation of metastasis at the lung [111,113,114]. When the endothelium recovered to normal status, metastases also decreased to the level of control group. Kennel [115] used monoantibody to directly mediate pulmonary endothelial injury and found lung metastasis increasing, further testifying that pulmonary endothelial injury was an element in enhancing lung metastasis. Oxygen radical and neutrophil cells are the mediators to regulate the process [113,116]. Nature killer cells, macrophage and their releasing proinflammatory factors such as TNF- α , interleukin-6 (IL-6), Toll-like receptors, MMP9 and NF-kappa B are also reported to be responsible for increasing lung metastasis [94,117–125]. Although there were controversies on neutrophils or macrophages and some molecules [101,113,117,121,123], the injured lung did promote lung metastasis. From the external environment, the host lung can be injured by various stimuli. In the host's inner status, the lung produces responses and shows physiopathological changes including endothelial cells injuries, immune and inflammation cells of activation accumulation and aggregation at the injured sites, and subsequently secretion of cytokines such as TGF- β , TNF- α , NF-kappa B and VEGF. From the external stimuli to inner changes of the host, they constitute a hierarchical system and collectively play the role in enhancing lung metastasis. Meanwhile, the lung begins to repair when the organ receives stimuli. From the initial stage to the inflammatory stage after stimuli-induced lung injury, the incidence of lung metastasis begins to increase and achieves peak. When the injured lung recovers, lung metastasis starts to decrease and disappears. Therefore, promoting lung metastasis by different-stimuli induced-lung injury is a spatial and temporal effect.

Bone is a fertile reservoir for a complex array of cells, cytokines and growth factors that are involved in bone turnover, response to injury [126,127]. The above mentioned cases showed that metastatic tumors occurred at sites of bone trauma, fracture and bone graft site [41,60–66]. These bones affected by metastasis have a common trait that they undergo a process of injury and repair and the normal bone homeostasis is disrupted, which may constitute a hospitable milieu for cancer cells. Thus, the bone affected by injury or inflammatory disease may increase bone metastases at the bone sites of wound. Bone trauma has been shown to promote bone metastasis in an experimental study [128]. However, the mechanism was not identified in the report. In the process of bone metastasis from lung cancer, TGF- β plays an important role [129], whose role in metastasis has been considered to be through the inflammatory pathway [33]. Therefore, enhancing bone

metastasis by bone trauma may be mediated by inflammatory response. The inflammatory disease of bone is also reported to increase metastasis in the organ. For example, autoimmune arthritis (AA) is a disease with changes of inflammation and deformity of the joints and also has other systemic effects including cellular infiltration and inflammation in the lungs. It was found that bone and lung metastasis increased in an animal model of AA through inoculating breast cancer cells [130,131]. This mechanism was related to neutrophilic and granulocytic infiltration in lungs and bones in the pro-arthritis and arthritic mice and subsequent increasing of circulating proinflammatory cytokines such as IL-6, VEGF and TNF- α , all of which have also been identified to play important roles in cancer metastasis [132,133]. Trauma bone site can also arouse dormant cancer cells to grow overt metastasis [60]. Its possible mechanism may be that dormant cancer cells at the site of bone trauma were exposed to pro-inflammatory mediators, which thereby stimulated to overcome dormancy. Prostate cancers are prone to metastasize to bones, which have been identified to be via the stromal cell-derived factor-1/CXCR4 pathway [134]. It has been also found that the signaling pathway is critical for the recruitment of mesenchymal stem cells to the fracture site during skeletal repair after injury [135], which may provide the molecular basis for bone metastasis from prostate cancer. These studies suggest that an inflammatory microenvironment in the bone is vital for bone metastasis.

Liver is another common site for metastasis. It has been reported that liver regeneration and wound healing also increase liver metastasis. Liver regeneration can accelerate liver cancer occurrence in animal models receiving partial hepatectomy [136], indicating that liver regeneration offer a suitable microenvironment for cancer cells. Several studies also showed that partial hepatectomy enhanced liver metastasis from other types of cancer cells [137–139], suggesting that liver regeneration was also link with liver metastasis. In the process of liver regeneration after partial hepatectomy, there was no underlying liver inflammation, as was the case in humans with liver cancer. DNA damage-response machinery and genomic instability increasing accounted for the major mechanism of partial hepatectomy, which further resulted in hepatocyte apoptosis, cell-cycle arrest, and senescence that might be contributed to cancer cells colonization and growth at the liver [136,140]. Obese mouse can produce a heightened inflammatory response in the liver. Wu et al.'s [141] study showed that hepatic metastasis increased in wild type of obese mouse and rarely occurred in the rats with chronic insulin-like growth factor-I (IGF-I) deficiency. In the process, IGF-I was necessary to activate and sustain the inflammatory response in the liver, indicating that obesity, inflammation response and IGF-I collectively played the role in enhancing liver metastasis. Ischemia/reperfusion-induced liver injury [142–144] had the similar effect, which was mediated by MMP9 [144] that

had been well reported to be vital for metastasis [145]. These studies suggest that the local microenvironment during the processes of liver injury and regeneration after stimuli provides a favorable milieu for cancer metastasis. However, liver cirrhosis is also a kind of injury resulted from different stimuli. In clinical practice, metastases seldom occur in cancer patients with liver cirrhosis [146–148]. It seems that liver cirrhosis with rare metastasis is contradicted with the conclusion of the injured liver promoting liver metastasis. How to solve the controversy? Focused this question, Qi, et al. [149] established the animal model of liver cirrhosis by carbon tetrachloride and observed liver metastasis increasing from B16F1 melanoma cells. In Kuriyama's [150] report, hepatic metastasis from hepatocellular carcinoma in an orthotopic mouse was also observed to occur in cirrhotic but not in normal liver. In the two experimental studies, the liver is undergoing a process injury and repair and liver metastasis increases. In clinical practice, the cirrhosis is a result of a long process of injury and repair after stimuli and liver metastasis rarely occurs. These indicate that it is the healing process response to injury not the result of injury (cirrhosis or fibrosis) that contributes to enhance metastasis in the liver.

Brain metastases (BM) are common for many types of cancer and lead an increasingly important cause of death in cancer patients. Its mechanism is under exploration. Berghoff [151] investigated 17 human autopsic tissue specimens of BM from some types of cancer and found that inflammatory response existed in BM, which was characterized by profound microglia activation with marked peritumoral accumulation and independent from treatment of methods. Microglia is the main immune effector cell population in the central nervous system and control immune cell recruitment and is rapidly activated and recruited to the site of injury after any type of brain injury, which is also important for formation of BM [152]. However, it is unknown whether metastatic tumors induced inflammatory responses in the brain or inflammatory microenvironment in the brain enhanced brain metastasis. Massagué's group has identified genes to mediate breast cancer metastasis to the brain [153]. These genes are often involved in vascular permeability and leukocyte infiltration during brain inflammatory processes [153], suggest that brain metastasis from breast cancer may be related to brain inflammation mediated by specific genes. The following experiment studies supplied direct evidence that brain injury enhanced BM. The blood–brain-barrier (BBB) permeability is vital for cancer cells to metastasize to the brain [153–155] while disruption of BBB is a common effect in the pathophysiology of various inflammatory conditions of the central nervous system [156]. Acute stress can induce BBB disruption through activating brain mast cells and corticotropin-releasing hormone secreted [157,158] that can result in focal brain inflammation and neurotoxicity [159,160]. When acute stress acted on the mice bearing tumor burden, activating mast cells and

releasing corticotropin-releasing hormone at the brain statistically significant increased BM through disruption of BBB [161]. Polychlorinated biphenyls (PCBs) contained in fish and meat comprise a ubiquitous class of toxic substances that can lead neurotoxic properties in the brain [162]. When mice were exposed to PCBs and inoculated by melanoma cells and Lewis lung carcinoma cells, the rate of formation of BM was enhanced [155,163]. This effect of PCBs enhancement of BM is facilitated by disrupt blood–brain barrier integrity [155] and proinflammatory adhesion molecules of intercellular cell adhesion molecule-1 and vascular endothelial cell adhesion molecule-1 (VCAM1) [163] in the brain, which may further suggest that avoiding to exposure to PCBs may decrease the incidence of BM in some extent. Although the evidence is few, together with BM occurring at the sites of focal infarction and infection in the brain [69,70], they may suggest that particular stimulus induced brain injury can enhance BM.

Kidney metastasis and kidney injury was also reported. Radiation induced kidney injury was demonstrated to enhance metastasis in the organ [164]. Ammirati et al. [165] also showed that kidney metastasis was enhanced at the trauma site of kidney induced by milliwatt carbon dioxide laser. This type of kidney injury has no obvious inflammation process, implying that it is that the process of wound healing in injured kidney promotes kidney metastasis.

Peritoneal injury can be induced by PF, anastomotic leakage in colorectal surgery, trocar insertion in laparoscopic techniques, etc. It has been reviewed that these causes induced peritoneal injury are negative prognostic factors for some solid tumors in abdominal cavity [37,38]. It has been speculated that these wound sites in abdominal cavity release proinflammatory cytokines to form an inflammatory microenvironment, which promote growth of residual or implanted cancer cells and finally lead to peritoneal recurrence [38]. Although there are controversies on whether the port sites of wound are preferred or contaminated by cancer cells, the following experimental studies testified that inducing peritoneal tissue injury by different causes could also enhance peritoneal metastases [9,166–170]. In contrast, repair of injured peritoneum at trocar sites or minimizing peritoneal trauma could reduce the frequency of wound metastases [37,166,171]. Moreover, there is a temporal effect for peritoneal injury promoting metastasis. For example, Zeamari et al. [172] studied the relationship between a peritoneal wound and tumor cell seeding and stimulated growth in granulation tissues, and found that tumor seeding mainly occurred at the early granulation tissue. Thus, these experimental studies support that peritoneal injury may potentiate wound metastases and may be a possible reason for peritoneal recurrence after different causes induced-peritoneal injury in clinical reports.

These experimental studies are consistent with clinical observations and further support that different stimulus

directly induced-specific organ injury can enhance metastasis in the injured site. This is a system effect through a spatio-temporal regulation, which includes external stimuli, specific organ and the following inner changes at the cellular and molecular levels with time. From the above statement, different stimulus direct action on the same organ can enhance metastases at the organ from the same type of cancer or different types of cancer, which means that there is a common status to promote organ-specific metastasis. This status can be characterized by an inflammatory status. When a particular stimulus acts on a specific organ in an organism bearing tumor, a healing process of injury and repair occurring as an inflammatory status contributes to the formation of organ-specific metastasis. Moreover, the formation of metastasis is related to the stage of wound healing, which may occur at an acute phase (acute lung infection promoting pulmonary metastasis) or a chronic process (smoking promoting lung metastasis) according to a stimulus intensity and frequency and the degree of specific organ making response to stimuli. When the inflammatory status is mild and quickly recovers to normal status, metastasis also rarely occurs. When the process leads severe endings for the particular organ such as liver cirrhosis or for the organism such as death, metastasis rarely or never occurs. Although the status is hard to be accurately defined, it can be evaluated and detected for the factors from external stimuli to inner changes at the organic, cellular and molecular levels that resulting in the status. Evaluating these factors and identifying the status may be useful to prevent or inhibit the formation of metastasis in distant organs.

Primary Tumor and Its Treatment Influencing on Distant Metastasis Through Activation of Wound and Inflammation Effects

It has been recognized that the status of primary tumor can determine organ-specific metastasis, which is also shown to be related with inflammation. At the cellular level, disseminated cancer cells from primary cancer are the seeds of distant metastasis. Cancer stem cells (CSCs) have been identified in primary solid tumors and are considered to contribute to distant metastasis [173]. Chemokines are a group of small pro-inflammatory chemoattractant cytokines, which serve as the major regulators of cell trafficking [174]. The SDF-1–CXCR4 Axis can regulate the trafficking of both normal and cancer stem cells [175], which can be activated by the inflamed tissues [135]. Thus, it maybe regulate mesenchymal stem cells and CSCs to recruit at the site of injury. Self-renewal is an important trait of CSCs [176]. The inflammatory cytokines and pathways generate positive feedback loops to drive CSC self-renewal [177]. These cytokine loops and the pathways will be activated during wound healing and chronic inflammation. Thus, CSCs will be droved to proliferate and

grow under inflammatory microenvironments. However, the theory of CSCs has been challenged by a study [178]. The study analyzed two subpopulations of CD24+ and CD44+ cells from breast cancer stem cells and revealed genetic difference between the two subpopulations, which questioned the validity of the cancer-stem-cell hypothesis in breast cancer and supported the classical model of clonal evolution [178]. In fact, Massagué's group has separated cancer cells subpopulations for organ-specific metastasis (bone, lung, brain) from the parental MDA-MB-231 cell line through in vivo selection [153,179,180]. They express a set of genes to respectively mediate breast cancer metastasis to the bone, lung and brain. These genes are involved in inflammatory pathway. Although these molecules are not reported whether they can induce organ-specific injury, they result in the changes of targeted organ similar to inflammatory response in corresponding organs (e.g. BBB disruption in the brain) that may be related to organ-specific metastasis. Stromal cells of primary cancer can affect premetastatic organs through indirect injury by their secretion of inflammatory modulators. They include many types of cells, fibroblasts, macrophages, lymphatic cells, all of which have been identified to be involved cancer metastasis [98,123,181–185]. Thus, under the conditions of distant organ with no obvious injury, they can release proinflammatory modulators such as s100A8 and s100A9 [186] to induce injuries on distant organs. At the molecular level, the gene signatures of primary tumor as prognostic factors have been used to predict distant metastasis [187]. Among of them, wound gene signature in primary cancer cells is one of these prognostic factors for predicting distant metastasis in breast cancer patients [188], indicating that the genes regulation of the wound healing process are also involved in cancer metastasis [189]. This is further supported in recent studies. They show that primary tumor cells can secrete proinflammatory molecules to induce distant site injury for formation of organ-specific metastasis. For example, endogenous human microRNAs of miR-335 and miR-126 are also identified to be metastasis suppressors in breast cancer through targeting of Tenascin-C [190], a modulator of tissue injury and tumorigenesis [191]. When they are lost in primary tumors, tenascin-C will upregulate to promote lung metastasis [190] while its upregulation can be also induced by lung infections [191]. Primary tumor-secreted miR-21 and miR-29a can bind to receptors of the Toll-like receptor (TLR) family to trigger a TLR-mediated prometastatic inflammatory response, which ultimately lead to tumor growth and metastasis [192]. Nuclear factor-kappaB, a critical proinflammatory molecule, can induce lung injury to potentiate lung metastasis when it is secreted by Lewis lung cancer cells [121]. At the physical and chemical levels, primary tumor microenvironment such as hypoxia and acid pH value can also influence distant metastasis [193–195] through up-regulation of inflammatory modulators including proteolytic enzymes, proangiogenic factors,

and antiapoptotic proteins [196–200]. At the organic level, status of primary organ can also determine the behavior of metastasis. It is well known that primary liver cancer mainly developed from the basis of cirrhosis liver induced by different stimuli and often metastasize to the liver itself. Budhu et al. [201] identified a unique inflammation/immune response-related signature in noncancerous hepatic tissues from metastatic hepatocellular carcinoma was principally different from that of the tumor, indicating that inflammatory status of primary organ can influence the site-specific metastasis. On the other hand, local treatment regimens such as surgery and radiation on primary tumors can also induce distant metastasis through activation of inflammatory cells and signals [202–209]. For example, removal of primary breast cancer can led to a progressive phenotype of lung metastases through upregulated the expression of ITGB3, EGFR, HGF, IGF1, PDGR-B, TNF- α , VEGFA, VEGFC and MMP9 genes and down regulation of Cdkn2a, Cdh1, and Syk genes [210] that are also involved lung inflammation. These suggest that primary tumors and local treatment of them may indirectly induce injuries on distant sites to sustain the process of distant metastases.

Host Factors Increasing Distant Metastasis Through System Inflammation and Immune System

Host factors can increase distant metastasis through activation of immune system and system inflammation, which includes postoperative fever, chronic stress, obesity, particular status such as pregnancy and lactation. Postoperative fever, a frequent event in clinical practice, has been identified to be a poor prognostic factor among patients of colorectal cancer, breast cancer, and soft-tissue sarcoma [6,211,212]. Its mechanism is speculated that postoperative fever is a response to tissue injury or infection of unknown trauma sites in the body, which induce to distant metastasis in these wound sites and result in poor prognosis for cancer patients [6]. Stress, social isolation and other factors can also influence cancer progression and metastasis under the modulation of neuro-endocrine-immune network [213–215]. For example, Wu and cooperators found that social isolation stress promotion of liver metastasis was via suppressed splenic NK cells activity and decreasing macrophage-mediated cytotoxicity at the cellular level [216], TNF-alpha increasing and the proteolytic enzymes such as MMPs and u-PA upregulating in tumor and liver tissues at the molecular level [217]. Sloan et al. [218] found that chronic stress could increase distant metastasis through activation of sympathetic nervous system (SNS) and increasing the expression of proinflammatory and prometastatic genes including COX2, MMP9, VEGF, and VCAM1 that has been identified to be involved site-specific metastasis [120,132,219]. These indicated that stress activated

the inflammatory pathway under the modulation of neuro-endocrine-immune network. Stress is also a common complication of operation for cancer patients. Combination of beta-blockers and non-steroidal anti-inflammatory drugs was shown to be correlated with reduced recurrence, metastasis and mortality [220], which further testify that fever and chronic stress can enhance distant metastasis. Obese cancer patients were reported to have high rate of reoccurrence [221,222], which were considered to be due to inflammatory changes [223]. Therefore, host factors such as chronic stress, social isolation, postoperative fever, and obesity activate the inflammatory pathway to indirectly promote distant metastasis.

Host organ is the end point of disseminated tumor cells (DTC) and responsible for organ-selective metastasis. DTC can reach any organ, even if most of them are captured by the first capillary bed in their journey of migration. Whether DTC develop a new tumor in ectopic sites is depended on the results of mutual effects between them [194,224]. On one hand, an inflammatory microenvironment of a host organ can alter the behavior of DTC. Okada [225] found that weakly tumorigenic and nonmetastatic QR-32 cells derived from a fibrosarcoma in C57BL6 mouse could be converted to malignant once they grew in an inflammation microenvironment. In brain metastasis, activation of astrocytes was vital for cancer cells growth [226], which was often related and inflammatory microenvironment of the brain [227]. On the other hand, DTC with strong ability of metastasis can act on host organ through activation of local inflammatory cells and system inflammation. Wang et al. [228] has shown that highly metastatic carcinoma cells can trigger a rapid host proinflammatory response by inducing TNF- α production in resident Kupffer cells once they entry into the hepatic microcirculation while weakly metastatic cells can't. Therefore, the mutual effect mediated by inflammatory pathways between host organ and DTC may contribute to organ-selective metastasis.

Bone marrow is now found to play an important role in distant metastasis [229,230]. Several studies have shown that bone marrow is a reservoir for DTC, which is not only the predictor factor of bone metastasis but also other site's metastasis. Moreover, there are also reports to show that bone marrow-derived progenitor cells can home to pre-metastatic sites such as liver and lung to form a premetastatic niche before the arrival of tumour cells [186,231,232]. Nowadays, it is well known that bone marrow-derived mesenchymal stem cells (BMSCs) can repair injured tissues [233,234]. In the host, liver and pulmonary is easily inflamed by different stimuli and often undergo injury and repair. In the process, BMSCs play an important role [235–237]. This may partly explain why DTC can predict liver and lung metastasis and why liver and lung are the most sites for metastasis from different cancers.

Different Stimuli-induced Organ Injury Enhancement of Organotropism Metastases in a System Effect

From the above analyses, we can see that not only inflammation but also a wide meaning of injuries on a target organ can enhance metastases in this organ. Moreover, it is the process of repair or wound healing response to different kinds of injuries not the result of injury that exerts this effect. For example, the process of development of cirrhosis can enhance liver metastasis while metastasis rarely occurs in the liver when cirrhosis forms. Therefore, organ-preference metastasis enhanced by organ-specific injury is in a spatial and temporal regulation (Fig. 1). At the spatial level, stimuli, host factors, status of primary cancer and its organ, premetastatic organs' microenvironments, state of bone marrow, system and local immune and inflammatory effects including activated immune and inflammatory cells, released chemokines, growth factors and cytokines are all involved in the determining the site of distant metastasis [238]. From the temporal level, at the initial stage of wound healing in an organ after receiving a stimulus, metastasis begins to form. With the inflammatory effects development and repair increasing, metastasis will increase at the injured site. When the injured organ completely recovers or results in a bad state such as hepatic cirrhosis, metastasis rarely occurs. This is a system effect. Only when stimuli-induced organ injury reaches this status at the spatio-temporal effect, organotropism metastasis can be promoted, which may explain why a negative result exists between trauma and an increased rate of breast cancer recurrence in Allawi et al. [239]'s report. Deep understanding the above accurately mechanism has a significance for biological and clinical implications on cancer metastasis.

Firstly, when a patient is diagnosed of cancer, it should be recognized that potentially harmful stimuli are taking effects on his or her whole organism or particular organ. So these stimuli (e.g. smoking, PCBs in a food chain, etc.) should be avoided and the status of the injured organ should be converted into normal state. Secondly, some inflammatory diseases such as acute lung infections, asthma, inflamed gingiva, autoimmune arthritis lead to inflammatory status in target organs that are preferred sites of metastasis. These diseases should be positively managed in cancer patients. The relationship between other inflammatory diseases such as COPD and cancer metastasis should be also investigated in the future studies. The trauma sites or wound tissues undergoing a process of injury and repair may be preferred by cancer metastasis, which should be aware of and often checked after cancer diagnosis. Thirdly, host factors such as fever related to system inflammation and stress in cancer patients should be adjusted. Inhibition of system inflammation by using aspirin can decrease distant metastasis [240]. Detection of factors of system inflammation such as acute phase protein and IL-6 may be helpful to predict of metastasis [241–243]. It has been

shown that beta-blocker can increase the clinical outcome of breast cancer patients through inhibition the activities of SNS, which mainly reduces distant recurrence [244,245]. Thirdly, when a tumor is diagnosed, its primary organ status, and premetastatic organ or metastatic organ status should be evaluated so as to decrease metastasis occurrence (Fig. 1). Finally, because different stimuli induced organ-specific injury can enhance organotropism metastasis from the same type of cancer, it is vital for inhibition organ-specific metastasis to find similar effects on the injured organ produced by these stimuli. A same stimulus can also enhance organ-specific metastasis from different tumors, indicating that there is a common status for lodging in a same organ among these tumors. Discovering the common status will be also to treat different metastasis. These evaluations and measurements for cancer patients will make personalized in prediction, prevention and management of cancer metastasis.

Conclusions

In conclusion, we propose a notion that chronic injury not merely chronic inflammation in an organ can enhance metastasis in this organ through a spatiotemporal regulation. This is a system effect including multiple scales. From the spatial scales, external stimuli (chemical, physical and biological factors) directly act on a specific organ in an organism that makes

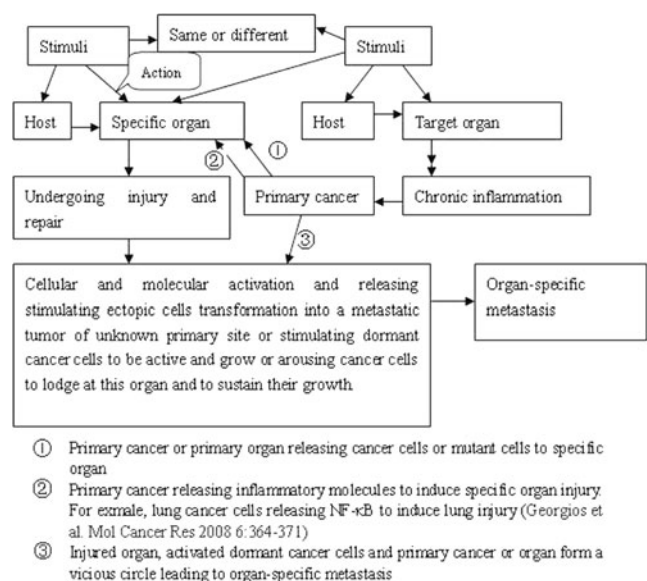


Fig. 1 Stimuli directly or indirectly induced specific organ injury and organotropic metastasis. When human is exposed to internal and external environments, the body will receive stimuli and make response to stimuli. In the reaction, stimuli always act on specific organ and induce organ-specific injury. When a particular organ undergoes persistent and chronic injury, it will occur to cancer. If there is another organ undergoing persistent injury, it will provide a favorable environment for cancer cells, this organ will be a preferred site for metastasis

response at cellular and molecular levels, which finally induce organ specific injury to promote organotropic metastasis. Primary cancer is not only the source of ‘seeds’ of distant metastases but also can release inflammatory molecules to induce injury on distant sites. Thus, cancer cells interaction with the injured organ contribute to organotropic metastasis. The host factors such as chronic stress, fever, obesity indirectly influence on distant metastases through activation of the inflammatory pathway under the modulation systems of neuro-endocrine-immune and bone marrow. Many factors can be included in each scale. They interact at the time scale and generate an inflammatory status at a particular organ or specific some organs in cancer patients to be suitable for cancer cells growth and proliferation. Finally, cancer metastases occur at one or more specific organs. Therefore, occurrence of organotropic metastasis is a stem effect and management of cancer metastasis should be comprehensively considered at multiple scales in a systems biology view. Evaluation of these factors at each scale using system biology methods and identifying organ-specific injury is vital for understanding the biology of organotropic metastasis and personalized management of metastasis.

Conflict of Interest Dongwei Gao has been recruited by Lanzhou General hospital of PLA but not permitted by the 536th hospital. He now works in the 536th hospital and conducts experimental studies in Lanzhou General Hospital of PLA. We declare no other personal, commercial or financial conflicts.

References

- Balkwill F, Mantovani A (2001) Inflammation and cancer: Back to Virchow? *Lancet* 357:539–545
- Hans C (2004) At the crossroads of inflammation and cancer. *Cell* 118:671–674
- Lu H, Ouyang W, Huang C (2006) Inflammation, a key event in cancer development. *Mol Cancer Res* 4:221–233
- Schottenfeld D, Beebe-Dimmer J (2006) Chronic inflammation: A common and important factor in the pathogenesis of neoplasia. *CA Cancer J Clin* 56:69–83
- Grivennikov SI, Karin M (2010) Inflammation and oncogenesis: A vicious connection. *Curr Opin Genet Dev* 20:65–71
- Nowacki MP, Janik P, Nowacki PM (1996) Inflammation and metastases. *Med Hypotheses* 47:193–196
- Mantovani A (2009) Cancer: Inflaming metastasis. *Nature* 457:36–37
- Solinas G, Marchesi F, Garlanda C, Mantovani A, Allavena P (2010) Inflammation-mediated promotion of invasion and metastasis. *Cancer Metastasis Rev* 29:243–248
- Jones FS, Rous P (1914) On the cause of the localization of secondary tumors at points of injury. *J Exp Med* 20:404–412
- Brody JS, Spira A (2006) State of the art. Chronic obstructive pulmonary disease, inflammation, and lung cancer. *Proc Am Thorac Soc* 3:535–537
- Walser T, Cui X, Yanagawa J, Lee JM, Heinrich E, Lee G, Sharma S, Dubinett SM (2008) Smoking and lung cancer: The role of inflammation. *Proc Am Thorac Soc* 5:811–815
- Calle EE, Miracle-McMahill HL, Thun MJ, Heath CJ (1994) Cigarette smoking and risk of fatal breast cancer. *Am J Epidemiol* 139:1001–1007
- Yu GP, Ostroff JS, Zhang ZF, Tang J, Schantz SP (1997) Smoking history and cancer patient survival: A hospital cancer registry study. *Cancer Detect Prev* 21:497–509
- Shaw HM, Milton GW (1981) Smoking and the development of metastases from malignant melanoma. *Int J Cancer* 28:153–156
- Scanlon EF, Suh O, Murthy SM, Mettlin C, Reid SE, Cummings KM (1995) Influence of smoking on the development of lung metastases from breast cancer. *Cancer* 75:2693–2699
- Murin S, Inciardi J (2001) Cigarette smoking and the risk of pulmonary metastasis from breast cancer. *Chest* 119:1635–1640
- Abrams JA, Lee PC, Port JL, Altorki NK, Neugut AI (2008) Cigarette smoking and risk of lung metastasis from esophageal cancer. *Cancer Epidemiol Biomarkers Prev* 17:2707–2713
- Meliska CJ, Stunkard ME, Gilbert DG, Jensen RA, Martinko JM (1995) Immune function in cigarette smokers who quit smoking for 31 days. *J Allergy Clin Immunol* 95:901–910
- Mayhan WG, Sharpe GM (1998) Nicotine impairs histamine-induced increases in macromolecular efflux: Role of oxygen radicals. *J Appl Physiol* 84:1589–1595
- Conklin BS, Zhao W, Zhong DS, Chen C (2002) Nicotine and cotinine up-regulate vascular endothelial growth factor expression in endothelial cells. *Am J Pathol* 160:413–418
- Lu Q, Newton J, Gabino-Miranda G, Ortiz M, Harrington EO, Rounds SIS (2010) Cigarette smoke extract increases lung endothelial permeability through oxidative stress-mediated alterations of small GTPases. *Am J Resp Crit Care* 181:A3427
- Padua D, Zhang XH, Wang Q, Nadal C, Gerald WL, Gomis RR, Massague J (2008) TGFbeta primes breast tumors for lung metastasis seeding through angiopoietin-like 4. *Cell* 133:66–77
- O’Byrne PM, Postma DS (1999) The many faces of airway inflammation. *Asthma and chronic obstructive pulmonary disease. Asthma Research Group. Am J Respir Crit Care Med* 159:S41–S63
- Bosse Y, Rola-Pleszczynski M (2007) Controversy surrounding the increased expression of TGF beta 1 in asthma. *Respir Res* 8:66
- Fidler IJ (1990) Critical factors in the biology of human cancer metastasis: Twenty-eighth G.H.A. clowes memorial award lecture. *Cancer Res* 50:6130–6138
- Mannel DN, Orosz P, Hafner M, Falk W (1994) Mechanisms involved in metastasis enhanced by inflammatory mediators. *Circ Shock* 44:9–13
- Fidler IJ (1995) Critical factors in the biology of human cancer metastasis. *Am Surg* 61:1065–1066
- Bogenrieder T, Herlyn M (2003) Axis of evil: Molecular mechanisms of cancer metastasis. *Oncogene* 22:6524–6536
- Lin WW, Karin M (2007) A cytokine-mediated link between innate immunity, inflammation, and cancer. *J Clin Invest* 117:1175–1183
- Liao D, Luo Y, Markowitz D, Xiang R, Reisfeld RA (2009) Cancer associated fibroblasts promote tumor growth and metastasis by modulating the tumor immune microenvironment in a 4 T1 murine breast cancer model. *PLoS One* 4:e7965
- Finger EC, Giaccia AJ (2010) Hypoxia, inflammation, and the tumor microenvironment in metastatic disease. *Cancer Metastasis Rev* 29:285–293
- Erez N, Coussens LM (2011) Leukocytes as paracrine regulators of metastasis and determinants of organ-specific colonization. *Int J Cancer* 128:2536–2544
- Yang L (2010) TGFbeta and cancer metastasis: An inflammation link. *Cancer Metastasis Rev* 29:263–271
- Taranova AG, Maldonado DR, Vachon CM, Jacobsen EA, Abdala-Valencia H, McGarry MP, Ochkur SI, Protheroe CA, Doyle A,

- Grant CS, Cook-Mills J, Birnbaumer L, Lee NA, Lee JJ (2008) Allergic pulmonary inflammation promotes the recruitment of circulating tumor cells to the lung. *Cancer Res* 68:8582–8589
35. Formenti SC, Demaria S (2009) Systemic effects of local radiotherapy. *Lancet Oncol* 10:718–726
36. Dao TL, Kovacic J (1962) Incidence of pulmonary and skin metastases in women with breast cancer who received postoperative irradiation. *Surgery* 52:203–212
37. Oosterling SJ, van der Bij GJ, van Egmond M, van der Sijp JR (2005) Surgical trauma and peritoneal recurrence of colorectal carcinoma. *Eur J Surg Oncol* 31:29–37
38. Nagai S, Fujii T, Koderia Y, Kanda M, Sahin TT, Kanzaki A, Hayashi M, Sugimoto H, Nomoto S, Takeda S, Morita S, Nakao A (2011) Recurrence pattern and prognosis of pancreatic cancer after pancreatic fistula. *Ann Surg Oncol* 18:2329–2337
39. Allon I, Pessing A, Kaplan I, Allon DM, Hirshberg A (2013) Metastatic tumors to the gingiva, the presence of teeth as a contributing factor- a literature analysis. *J Periodontol*
40. Black JW (1964) The localisation of metastatic brown-pearce carcinoma in granulation tissue. *Br J Cancer* 18:143–145
41. Benisch B, Toker C (1971) Metastasis to the site of recent trauma. *JAMA* 216:2142–2143
42. Cohen HJ, Laszlo J (1972) Influence of trauma on the unusual distribution of metastases from carcinoma of the larynx. *Cancer* 29:466–471
43. Cunha-Gomes D, Prasad R, Bhathena HM, Kavarana NM (1999) Tumor implantation at the flap donor site: A case report. *Acta Chir Plast* 41:75–76
44. Shine T, Wallack MK (1981) Inflammatory oncotaxis after testing the skin of the cancer patient. *Cancer* 47:1325–1328
45. Marley NF, Marley WM (1982) Skin metastases in an area of radiation dermatitis. *Arch Dermatol* 118:129–131
46. Ito H, Kubo A, Shigematsu N, Hashimoto S (1984) Skin metastases within the previous radiation field after prophylactic postoperative radiotherapy for breast cancer. *Clin Exp Metastasis* 2:235–239
47. Carr RJ, Gilbert PM (1986) Tumour implantation to a temporalis muscle flap donor site. *Br J Oral Maxillofac Surg* 24:102–106
48. Climo MS, Enos WF, King PG (1989) Squamous cell carcinoma of skin developing in a skin graft donor site. *Br J Plast Surg* 42:118
49. Betke M, Suss R, Hohenleutner U, Lubke S, Eckert F (1993) Gastric carcinoma metastatic to the site of a congenital melanocytic nevus. *J Am Acad Dermatol* 28:866–869
50. Trefzer U, Schwurzer-Voit M, Audring H, Jahn S, Thies E, Sterry W (1998) Multiple melanoma metastases in split-thickness skin graft donor sites. *J Am Acad Dermatol* 38:997–998
51. Serrano-Ortega S, Buendia-Eisman A, Ortega DOR, Linares SJ (2000) Melanoma metastasis in donor site of full-thickness skin graft. *Dermatology* 201:377–378
52. Pradhan S, Asthana AK, Sharan GK, Kumar M, Sharma OP (2006) Recurrence of carcinoma cervix in the scar of previous cesarean section: A case report. *Int J Gynecol Cancer* 16:900–904
53. Erol B, Ufuk U, Husamettin T, Yasin U, Aslihan C, Koray K (2008) True hematogenous metastases of melanoma on contralateral skin graft donor site: A case report. *Melanoma Res* 18:443–446
54. Marengo F, Fava P, Macripio G, Quagliano P, Savoia P, Bernengo MG (2009) Cutaneous melanoma metastases arising on a split-skin graft donor site. *Dermatol Surg* 35:1282–1285
55. Hussain A, Ekwobi C, Watson S (2011) Metastatic implantation squamous cell carcinoma in a split-thickness skin graft donor site. *J Plast Reconstr Aesthet Surg* 64:690–692
56. Dunn PT, Bigler CF (1997) Metastasis in an electrodesiccation and curettage scar. *J Am Acad Dermatol* 36:117–118
57. Ferguson PC, Somerville S, Grimer RJ (2004) Possible metastasis of osteosarcoma to a remote biopsy site: a case report. *Clin Orthop Relat Res* 424:216–220
58. Mincheff TV (2005) Metastatic spread to a percutaneous gastrostomy site from head and neck cancer: Case report and literature review. *JSL* 9:466–471
59. Hameed H, Khan YI (2009) Metastasis of carcinosarcoma of oesophagus to gastrostomy site. *Br J Oral Maxillofac Surg* 47:643–644
60. El Saghir N, Elhaji I, Geara F, Hourani M (2005) Trauma-associated growth of suspected dormant micrometastasis. *Bmc Cancer* 5:94
61. Fukushima M, Katagiri A, Mori T, Watanabe T, Katayama Y (2010) Case of skull metastasis from hepatocellular carcinoma at the site of skull fracture. *No Shinkei Geka* 38:371–377
62. Walter ND, Rice PL, Redente EF, Kauvar EF, Lemond L, Aly T, Wanebo K, Chan ED (2011) Wound healing after trauma may predispose to lung cancer metastasis: Review of potential mechanisms. *Am J Respir Cell Mol Biol* 44:591–596
63. Bergqvist D, Mattsson J (1978) Solitary calcaneal metastasis as the first sign of gastric cancer. A case report. *Ups J Med Sci* 83:115–118
64. De Simone P, Carrai P, Morelli L, Coletti L, Petrucci S, Filippini F, Doria R, Menichetti F, Vannozzi R (2005) Posttransplant hepatocellular carcinoma metastasis at a skull trauma site. *Transplantation* 80:1358–1359
65. Yip KM, Lin J, Kumta SM (1996) A pelvic osteosarcoma with metastasis to the donor site of the bone graft. A case report. *Int Orthop* 20:389–391
66. Wondergem JH, Holscher HC, Arndt JW, Bakker W, Pauwels EK (1987) A long term scintigraphic bone abnormality after trauma: A pitfall in the survey for metastatic disease. *Eur J Nucl Med* 13:321–323
67. Ivarsson L (1976) Pulmonary metastasis formation after trauma. *Acta Chir Scand Suppl* 463:1–46
68. Kukita K, Shirakawa T, Kojima A, Yoshida H, Yoshida K, Tokumomi H, Kurano R (1992) An autopsy case of pulmonary metastasis of cholangiocellular carcinoma associated with marked fibrotic change of the lungs. *Nihon Kyobu Shikkan Gakkai Zasshi* 30:1738–1742
69. Nielsen SL, Posner JB (1983) Brain metastasis localized to an area of infarction. *J Neurooncol* 1:191–195
70. Rodriguez RE, Valero V, Watanakunakorn C (1986) Salmonella focal intracranial infections: Review of the world literature (1884–1984) and report of an unusual case. *Rev Infect Dis* 8:31–41
71. Palazzo FF, New NE, Cullen PT (2000) An unusual cause of thigh pain in colonic cancer. Inflammatory oncotaxis? *Acta Chir Belg* 100:28–30
72. Magee T, Rosenthal H (2002) Skeletal muscle metastases at sites of documented trauma. *AJR Am J Roentgenol* 178:985–988
73. Currall VA, Dixon JH (2008) Synovial metastasis: An unusual cause of pain after total knee arthroplasty. *J Arthroplasty* 23:631–636
74. Dionigi G, Uccella S, Gandolfo M, Lai A, Bertocchi V, Rovera F, Tanda ML (2008) Solitary intrathyroidal metastasis of renal clear cell carcinoma in a toxic substernal multinodular goiter. *Thyroid Res* 1:6
75. DerHagopian RP, Sugarbaker EV, Ketcham A (1978) Inflammatory oncotaxis. *JAMA* 240:374–375
76. Murin S, Pinkerton KE, Hubbard NE, Erickson K (2004) The effect of cigarette smoke exposure on pulmonary metastatic disease in a murine model of metastatic breast cancer. *Chest* 125:1467–1471
77. Kim H, Zamel R, Bai XH, Liu M (2013) PKC activation induces inflammatory response and cell death in human bronchial epithelial cells. *PLoS One* 8:e64182
78. Gopalakrishna R, Chen ZH, Gundimeda U (1994) Tobacco smoke tumor promoters, catechol and hydroquinone, induce oxidative regulation of protein kinase C and influence invasion and metastasis of lung carcinoma cells. *Proc Natl Acad Sci U S A* 91:12233–12237
79. Blache D (1995) Involvement of hydrogen and lipid peroxides in acute tobacco smoking-induced platelet hyperactivity. *Am J Physiol* 268:H679–H685

80. Kunita A, Kashima TG, Morishita Y, Fukayama M, Kato Y, Tsuruo T, Fujita N (2007) The platelet aggregation-inducing factor aggrus/podoplanin promotes pulmonary metastasis. *Am J Pathol* 170: 1337–1347
81. Yan L, Cai Q, Xu Y (2013) The ubiquitin-CXCR4 axis plays an important role in acute lung infection-enhanced lung tumor metastasis. *Clin Cancer Res* 19:4706–4716
82. Muller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, McClanahan T, Murphy E, Yuan W, Wagner SN, Barrera JL, Mohar A, Verastegui E, Zlotnik A (2001) Involvement of chemokine receptors in breast cancer metastasis. *Nature* 410:50–56
83. Smith HA, Kang Y (2013) Acute infection induces a metastatic niche: A double menace for cancer patients. *Clin Cancer Res* 19: 4547–4549
84. Carmel RJ, Brown JM (1977) The effect of cyclophosphamide and other drugs on the incidence of pulmonary metastases in mice. *Cancer Res* 37:145–151
85. Milas L, Malenica B, Allegretti N (1979) Enhancement of artificial lung metastases in mice caused by cyclophosphamide. *Cancer Immunol Immunother* 6:191–196
86. Yamauchi K, Yang M, Hayashi K, Jiang P, Yamamoto N, Tsuchiya H, Tomita K, Moossa AR, Bouvet M, Hoffman RM (2008) Induction of cancer metastasis by cyclophosphamide pretreatment of host mice: An opposite effect of chemotherapy. *Cancer Res* 68: 516–520
87. Gasic GJ, Iwakawa A, Gasic TB, Viner ED, Milas L (1984) Leech salivary gland extract from *Haementeria officinalis*, a potent inhibitor of cyclophosphamide- and radiation-induced artificial metastasis enhancement. *Cancer Res* 44:5670–5676
88. Man S, Zhang Y, Gao W, Yan L, Ma C (2008) Cyclophosphamide promotes pulmonary metastasis on mouse lung adenocarcinoma. *Clin Exp Metastasis* 25:855–864
89. Wu YJ, Muldoon LL, Dickey DT, Lewin SJ, Varallyay CG, Neuwelt EA (2009) Cyclophosphamide enhances human tumor growth in nude rat xenografted tumor models. *Neoplasia* 11:187–195
90. Sleijfer S (2001) Bleomycin-induced pneumonitis. *Chest* 120:617–624
91. Chung MP, Monick MM, Hamzeh NY, Butler NS, Powers LS, Hunninghake GW (2003) Role of repeated lung injury and genetic background in bleomycin-induced fibrosis. *Am J Respir Cell Mol Biol* 29:375–380
92. Schmidt R, Ruppert C, Markart P, Lubke N, Ermer L, Weissmann N, Breithacker A, Ermer M, Seeger W, Gunther A (2004) Changes in pulmonary surfactant function and composition in bleomycin-induced pneumonitis and fibrosis. *Toxicol Appl Pharmacol* 195: 218–231
93. Opdenakker G, Van Damme J (1992) Chemotactic factors, passive invasion and metastasis of cancer cells. *Immunol Today* 13:463–464
94. Hiratsuka S, Nakamura K, Iwai S, Murakami M, Itoh T, Kijima H, Shipley JM, Senior RM, Shibuya M (2002) MMP9 induction by vascular endothelial growth factor receptor-1 is involved in lung-specific metastasis. *Cancer Cell* 2:289–300
95. Mueller MM, Fusenig NE (2004) Friends or foes - bipolar effects of the tumour stroma in cancer. *Nat Rev Cancer* 4:839–849
96. Shimo T, Kubota S, Yoshioka N, Ibaragi S, Isowa S, Eguchi T, Sasaki A, Takigawa M (2006) Pathogenic role of connective tissue growth factor (CTGF/CCN2) in osteolytic metastasis of breast cancer. *J Bone Miner Res* 21:1045–1059
97. Affara NI, Coussens LM (2007) IKKalpha at the crossroads of inflammation and metastasis. *Cell* 129:25–26
98. Solinas G, Germano G, Mantovani A, Allavena P (2009) Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation. *J Leukoc Biol* 86:1065–1073
99. Valastyan S, Weinberg RA (2011) Tumor metastasis: Molecular insights and evolving paradigms. *Cell* 147:275–292
100. Orr FW, Young L, King GM, Adamson IY (1988) Preferential growth of metastatic tumors at the pleural surface of mouse lung. *Clin Exp Metastasis* 6:221–232
101. Adamson IY, Orr FW, Young L (1986) Effects of injury and repair of the pulmonary endothelium on lung metastasis after bleomycin. *J Pathol* 150:279–287
102. Orr FW, Adamson IY, Young L (1986) Promotion of pulmonary metastasis in mice by bleomycin-induced endothelial injury. *Cancer Res* 46:891–897
103. Orr FW, Adamson IY, Young L (1986) Quantification of metastatic tumor growth in bleomycin-injured lungs. *Clin Exp Metastasis* 4: 105–116
104. Saiki I, Milas L, Hunter N, Fidler IJ (1986) Treatment of experimental lung metastasis with local thoracic irradiation followed by systemic macrophage activation with liposomes containing muramyl tripeptide. *Cancer Res* 46:4966–4970
105. van den Brenk HA, Kelly H (1974) Potentiating effect of prior local irradiation of the lungs on pulmonary metastases. *Br J Radiol* 47: 332–336
106. Brown JM (1973) The effect of lung irradiation on the incidence of pulmonary metastases in mice. *Br J Radiol* 46:613–618
107. Withers HR, Milas L (1973) Influence of preirradiation of lung on development of artificial pulmonary metastases of fibrosarcoma in mice. *Cancer Res* 33:1931–1936
108. Biswas S, Guix M, Rinehart C, Dugger TC, Chytil A, Moses HL, Freeman ML, Arteaga CL (2007) Inhibition of TGF-beta with neutralizing antibodies prevents radiation-induced acceleration of metastatic cancer progression. *J Clin Invest* 117:1305–1313
109. Vincic L, Orr FW, Warner DJ, Suyama KL, Kay JM (1989) Enhanced cancer metastasis after monocrotaline-induced lung injury. *Toxicol Appl Pharmacol* 100:259–270
110. Orr FW, Adamson IY, Young L (1985) Pulmonary inflammation generates chemotactic activity for tumor cells and promotes lung metastasis. *Am Rev Respir Dis* 131:607–611
111. Adamson IY (1987) Tumor metastasis after hyperoxic injury and repair of the pulmonary endothelium. *Lab Invest* 57:71–77
112. Kobayashi T, Todoroki T, Sato H (1987) Enhancement of pulmonary metastasis of murine fibrosarcoma NR-FS by ozone exposure. *J Toxicol Environ Health* 20:135–145
113. Orr FW, Warner DJ (1987) Effects of neutrophil-mediated pulmonary endothelial injury on the localization and metastasis of circulating Walker carcinosarcoma cells. *Invasion Metastasis* 7:183–196
114. Orr FW, Warner DJ (1990) Effects of systemic complement activation and neutrophil-mediated pulmonary injury on the retention and metastasis of circulating cancer cells in mouse lungs. *Lab Invest* 62: 331–338
115. Kennel SJ, Lankford TK, Ullrich RL, Jamasbi RJ (1988) Enhancement of lung tumor colony formation by treatment of mice with monoclonal antibodies to pulmonary capillary endothelial cells. *Cancer Res* 48:4964–4968
116. Orr FW, Adamson IY, Warner D, Leroyer V, Werner L, Shaughnessy S, Young L (1988) The effects of oxygen radical-mediated pulmonary endothelial damage on cancer metastasis. *Mol Cell Biochem* 84:189–198
117. Hafner M, Orosz P, Kruger A, Mannel DN (1996) TNF promotes metastasis by impairing natural killer cell activity. *Int J Cancer* 66: 388–392
118. Wu WJ, Pruett SB (1999) Ethanol decreases host resistance to pulmonary metastases in a mouse model: Role of natural killer cells and the ethanol-induced stress response. *Int J Cancer* 82:886–892
119. Harney JH, Bucana CD, Lu W, Byrne AM, McDonnell S, Lynch C, Bouchier-Hayes D, Dong Z (2002) Lipopolysaccharide-induced metastatic growth is associated with increased angiogenesis, vascular permeability and tumor cell invasion. *Int J Cancer* 101:415–422

120. Gupta GP, Nguyen DX, Chiang AC, Bos PD, Kim JY, Nadal C, Gomis RR, Manova-Todorova K, Massague J (2007) Mediators of vascular remodelling co-opted for sequential steps in lung metastasis. *Nature* 446:765–770
121. Stathopoulos GT, Sherrill TP, Han W, Sadikot RT, Yull FE, Blackwell TS, Fingleton B (2008) Host nuclear factor-kappaB activation potentiates lung cancer metastasis. *Mol Cancer Res* 6: 364–371
122. Wang W, Xu GL, Jia WD, Ma JL, Li JS, Ge YS, Ren WH, Yu JH, Liu WB (2009) Ligation of TLR2 by versican: A link between inflammation and metastasis. *Arch Med Res* 40:321–323
123. Kim S, Takahashi H, Lin W, Descargues P, Grivennikov S, Kim Y, Luo J, Karin M (2009) Carcinoma-produced factors activate myeloid cells through TLR2 to stimulate metastasis. *Nature* 457:102–106
124. Maru Y (2010) Premetastatic milieu explained by TLR4 agonist-mediated homeostatic inflammation. *Cell Mol Immunol* 7:94–99
125. Orosz P, Echtenacher B, Falk W, Ruschoff J, Weber D, Mannel DN (1993) Enhancement of experimental metastasis by tumor necrosis factor. *J Exp Med* 177:1391–1398
126. Ai-Aql ZS, Alagl AS, Graves DT, Gerstenfeld LC, Einhorn TA (2008) Molecular mechanisms controlling bone formation during fracture healing and distraction osteogenesis. *J Dent Res* 87:107–118
127. Robling AG, Castillo AB, Turner CH (2006) Biomechanical and molecular regulation of bone remodeling. *Annu Rev Biomed Eng* 8: 455–498
128. Lee JY, Murphy SM, Scanlon EF (1994) Effect of trauma on implantation of metastatic tumor in bone in mice. *J Surg Oncol* 56:178–184
129. Vicent S, Luis-Ravelo D, Anton I, Garcia-Tunon I, Borrás-Cuesta F, Dotor J, De Las RJ, Lecanda F (2008) A novel lung cancer signature mediates metastatic bone colonization by a dual mechanism. *Cancer Res* 68:2275–2285
130. Das RL, Pathangey LB, Tinder TL, Schettini JL, Gruber HE, Mukherjee P (2009) Breast-cancer-associated metastasis is significantly increased in a model of autoimmune arthritis. *Breast Cancer Res* 11:R56
131. Roy LD, Ghosh S, Pathangey LB, Tinder TL, Gruber HE, Mukherjee P (2011) Collagen induced arthritis increases secondary metastasis in MMTV-PyV MT mouse model of mammary cancer. *Bmc Cancer* 11:365
132. Steeg PS (2006) Tumor metastasis: Mechanistic insights and clinical challenges. *Nat Med* 12:895–904
133. Roodman GD (2004) Mechanisms of bone metastasis. *N Engl J Med* 350:1655–1664
134. Taichman RS, Cooper C, Keller ET, Pienta KJ, Taichman NS, McCauley LK (2002) Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res* 62:1832–1837
135. Kitaori T, Ito H, Schwarz EM, Tsutsumi R, Yoshitomi H, Oishi S, Nakano M, Fujii N, Nagasawa T, Nakamura T (2009) Stromal cell-derived factor 1/CXCR4 signaling is critical for the recruitment of mesenchymal stem cells to the fracture site during skeletal repair in a mouse model. *Arthritis Rheum* 60:813–823
136. Barash H, GE R, Edrei Y, Ella E, Israel A, Cohen I, Corchia N, Ben-Moshe T, Pappo O, Pikarsky E, Goldenberg D, Shiloh Y, Galun E, Abramovitch R (2010) Accelerated carcinogenesis following liver regeneration is associated with chronic inflammation-induced double-strand DNA breaks. *Proc Natl Acad Sci U S A* 107:2207–2212
137. Fisher B, Fisher ER (1959) Experimental studies of factors influencing hepatic metastases. II. Effect of partial hepatectomy. *Cancer* 12: 929–932
138. Murthy SM, Goldschmidt RA, Rao LN, Ammirati M, Buchmann T, Scanlon EF (1989) The influence of surgical trauma on experimental metastasis. *Cancer* 64:2035–2044
139. Uotani H, Yamashita I, Nagata T, Kishimoto H, Kashii Y, Tsukada K (2001) Induction of E-selectin after partial hepatectomy promotes metastases to liver in mice. *J Surg Res* 96:197–203
140. Harun N, Nikfarjam M, Muralidharan V, Christophi C (2007) Liver regeneration stimulates tumor metastases. *J Surg Res* 138:284–290
141. Wu Y, Brodt P, Sun H, Mejia W, Novosyadlyy R, Nunez N, Chen X, Mendoza A, Hong SH, Khanna C, Yakar S (2010) Insulin-like growth factor-I regulates the liver microenvironment in obese mice and promotes liver metastasis. *Cancer Res* 70:57–67
142. Nijkamp MW, Hoogwater FJ, Govaert KM, Steller EJ, Verheem A, Kranenburg O, Borel RI (2011) A role for CD95 signaling in ischemia/reperfusion-induced invasion and outgrowth of colorectal micrometastases in mouse liver. *J Surg Oncol* 104:198–204
143. Doi K, Horiuchi T, Uchinami M, Tabo T, Kimura N, Yokomachi J, Yoshida M, Tanaka K (2002) Hepatic ischemia-reperfusion promotes liver metastasis of colon cancer. *J Surg Res* 105:243–247
144. Nicoud IB, Jones CM, Pierce JM, Earl TM, Matrisian LM, Chari RS, Gorden DL (2007) Warm hepatic ischemia-reperfusion promotes growth of colorectal carcinoma micrometastases in mouse liver via matrix metalloproteinase-9 induction. *Cancer Res* 67: 2720–2728
145. Nguyen DX, Bos PD, Massague J (2009) Metastasis: From dissemination to organ-specific colonization. *Nat Rev Cancer* 9:274–284
146. Uetsuji S, Yamamura M, Yamamichi K, Okuda Y, Takada H, Hioki K (1992) Absence of colorectal cancer metastasis to the cirrhotic liver. *Am J Surg* 164:176–177
147. Melato M, Laurino L, Mucli E, Valente M, Okuda K (1989) Relationship between cirrhosis, liver cancer, and hepatic metastases. An autopsy study. *Cancer* 64:455–459
148. Seymour K, Chamley RM (1999) Evidence that metastasis is less common in cirrhotic than normal liver: A systematic review of post-mortem case-control studies. *Br J Surg* 86:1237–1242
149. Qi K, Qiu H, Sun D, Minuk GY, Lizardo M, Rutherford J, Orr FW (2004) Impact of cirrhosis on the development of experimental hepatic metastases by B16F1 melanoma cells in C57BL/6 mice. *Hepatology* 40:1144–1150
150. Kuriyama S, Yamazaki M, Mito A, Tsujimoto T, Kikukawa M, Tsujinoue H, Nakatani T, Toyokawa Y, Yoshiji H, Fukui H (1999) Hepatocellular carcinoma in an orthotopic mouse model metastasizes intrahepatically in cirrhotic but not in normal liver. *Int J Cancer* 80:471–476
151. Berghoff AS, Lassmann H, Preusser M, Hoftberger R (2013) Characterization of the inflammatory response to solid cancer metastases in the human brain. *Clin Exp Metastasis* 30:69–81
152. Noda M, Seike T, Fujita K, Yamakawa Y, Kido M, Iguchi H (2009) The role of immune cells in brain metastasis of lung cancer cells and neuron-tumor cell interaction. *Russ Fiziol Zh Im IM Sechenova* 95: 1386–1396
153. Bos PD, Zhang XH, Nadal C, Shu W, Gomis RR, Nguyen DX, Minn AJ, van de Vijver MJ, Gerald WL, Foekens JA, Massague J (2009) Genes that mediate breast cancer metastasis to the brain. *Nature* 459:1005–1009
154. Wilhelm I, Molnar J, Fazakas C, Hasko J, Krizbai IA (2013) Role of the blood-brain barrier in the formation of brain metastases. *Int J Mol Sci* 14:1383–1411
155. Seelbach M, Chen L, Powell A, Choi YJ, Zhang B, Hennig B, Toborek M (2010) Polychlorinated biphenyls disrupt blood-brain barrier integrity and promote brain metastasis formation. *Environ Health Perspect* 118:479–484
156. Coisne C, Engelhardt B (2011) Tight junctions in brain barriers during central nervous system inflammation. *Antioxid Redox Signal* 15:1285–1303
157. Esposito P, Gheorghe D, Kandere K, Pang X, Connolly R, Jacobson S, Theoharides TC (2001) Acute stress increases permeability of the blood-brain-barrier through activation of brain mast cells. *Brain Res* 888:117–127

158. Esposito P, Chandler N, Kandere K, Basu S, Jacobson S, Connolly R, Tutor D, Theoharides TC (2002) Corticotropin-releasing hormone and brain mast cells regulate blood–brain-barrier permeability induced by acute stress. *J Pharmacol Exp Ther* 303:1061–1066
159. Theoharides TC, Asadi S, Patel AB (2013) Focal brain inflammation and autism. *J Neuroinflammation* 10:46
160. Theoharides TC, Cochrane DE (2004) Critical role of mast cells in inflammatory diseases and the effect of acute stress. *J Neuroimmunol* 146:1–12
161. Rozniecki JJ, Sahagian GG, Kempuraj D, Tao K, Jacobson S, Zhang B, Theoharides TC (2010) Brain metastases of mouse mammary adenocarcinoma is increased by acute stress. *Brain Res* 1366:204–210
162. Li A, Rockne KJ, Sturchio N, Song W, Ford JC, Wei H (2009) PCBs in sediments of the great lakes—distribution and trends, homolog and chlorine patterns, and in situ degradation. *Environ Pollut* 157:141–147
163. Sipos E, Chen L, Andras IE, Wrobel J, Zhang B, Pu H, Park M, Eum SY, Toborek M (2012) Proinflammatory adhesion molecules facilitate polychlorinated biphenyl-mediated enhancement of brain metastasis formation. *Toxicol Sci* 126:362–371
164. Van Den Brenk HA, Kelly H (1973) Stimulation of growth of metastases by local x-irradiation in kidney and liver. *Br J Cancer* 28:349–353
165. Ammirati MRLMM (1989) Partial nephrectomy in mice with milliwatt carbon dioxide laser and its influence on experimental metastasis. *J Surg Oncol* 3:153–159
166. Aoki Y, Shimura H, Li H, Mizumoto K, Date K, Tanaka M (1999) A model of port-site metastases of gallbladder cancer: The influence of peritoneal injury and its repair on abdominal wall metastases. *Surgery* 125:553–559
167. Hopkins MP, Dulai RM, Occhino A, Holda S (1999) The effects of carbon dioxide pneumoperitoneum on seeding of tumor in port sites in a rat model. *Am J Obstet Gynecol* 181:1329–1334
168. Ost MC, Patel KP, Rastinehad AR, Chu PY, Anderson AE, Smith AD, Lee BR (2008) Pneumoperitoneum with carbon dioxide inhibits macrophage tumor necrosis factor- α secretion: Source of transitional-cell carcinoma port-site metastasis, with prophylactic irrigation strategies to decrease laparoscopic oncologic risks. *J Endourol* 22:105–112
169. Jones DB, Guo LW, Reinhard MK, Soper NJ, Philpott GW, Connett J, Fleshman JW (1995) Impact of pneumoperitoneum on trocar site implantation of colon cancer in hamster model. *Dis Colon Rectum* 38:1182–1188
170. Bouvy ND, Marquet RL, Jeekel H, Bonjer HJ (1996) Impact of gas(less) laparoscopy and laparotomy on peritoneal tumor growth and abdominal wall metastases. *Ann Surg* 224(694–700):700–701
171. van den Tol PM, van Rossen EE, van Eijck CH, Bonthuis F, Marquet RL, Jeekel H (1998) Reduction of peritoneal trauma by using nonsurgical gauze leads to less implantation metastasis of spilled tumor cells. *Ann Surg* 227:242–248
172. Zeamari S, Roos E, Stewart FA (2004) Tumour seeding in peritoneal wound sites in relation to growth-factor expression in early granulation tissue. *Eur J Cancer* 40:1431–1440
173. Li F, Tiede B, Massague J, Kang Y (2007) Beyond tumorigenesis: Cancer stem cells in metastasis. *Cell Res* 17:3–14
174. Schier AF (2003) Chemokine signaling: Rules of attraction. *Curr Biol* 13:R192–R194
175. Kucia M, Reca R, Miekus K, Wanzeck J, Wojakowski W, Janowska-Wieczorek A, Ratajczak J, Ratajczak MZ (2005) Trafficking of normal stem cells and metastasis of cancer stem cells involve similar mechanisms: Pivotal role of the SDF-1-CXCR4 axis. *Stem Cells* 23:879–894
176. Arwert EN, Hoste E, Watt FM (2012) Epithelial stem cells, wound healing and cancer. *Nat Rev Cancer* 12:170–180
177. Korkaya H, Liu S, Wicha MS (2011) Regulation of cancer stem cells by cytokine networks: Attacking cancer's inflammatory roots. *Clin Cancer Res* 17:6125–6129
178. Shipitsin M, Campbell LL, Argani P, Weremowicz S, Bloushtain-Qimron N, Yao J, Nikolskaya T, Serebryskaya T, Beroukhim R, Hu M, Halushka MK, Sukumar S, Parker LM, Anderson KS, Harris LN, Garber JE, Richardson AL, Schnitt SJ, Nikolsky Y, Gelman RS, Polyak K (2007) Molecular definition of breast tumor heterogeneity. *Cancer Cell* 11:259–273
179. Kang Y, Siegel PM, Shu W, Drobnjak M, Kakonen SM, Cordon-Cardo C, Guise TA, Massague J (2003) A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell* 3:537–549
180. Minn AJ, Gupta GP, Siegel PM, Bos PD, Shu W, Giri DD, Viale A, Olshen AB, Gerald WL, Massague J (2005) Genes that mediate breast cancer metastasis to lung. *Nature* 436:518–524
181. Mueller L, Goumas FA, Affeldt M, Sandtner S, Gehling UM, Briloff S, Walter J, Karnatz N, Lamszus K, Rogiers X, Broering DC (2007) Stromal fibroblasts in colorectal liver metastases originate from resident fibroblasts and generate an inflammatory microenvironment. *Am J Pathol* 171:1608–1618
182. Steeg PS (2005) Cancer biology: Emissaries set up new sites. *Nature* 438:750–751
183. Duda DG, Duyverman AM, Kohno M, Snuderl M, Steller EJ, Fukumura D, Jain RK (2010) Malignant cells facilitate lung metastasis by bringing their own soil. *Proc Natl Acad Sci U S A* 107:21677–21682
184. Tan W, Zhang W, Strasner A, Grivennikov S, Cheng JQ, Hoffman RM, Karin M (2011) Tumour-infiltrating regulatory T cells stimulate mammary cancer metastasis through RANKL-RANK signaling. *Nature* 470:548–553
185. Al-Shibli KIDTAS (2008) Prognostic effect of epithelial and stromal lymphocyte infiltration in non-small cell lung cancer. *Clin Cancer Res* 14:5220–5227
186. Hiratsuka S, Watanabe A, Aburatani H, Maru Y (2006) Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis. *Nat Cell Biol* 8:1369–1375
187. Ring BZ, Ross DT (2005) Predicting the sites of metastases. *Genome Biol* 6:241
188. Chang HY, Nuyten DS, Sneddon JB, Hastie T, Tibshirani R, Sorlie T, Dai H, He YD, Van'T VL, Bartelink H, van de Rijn M, Brown PO, van de Vijver MJ (2005) Robustness, scalability, and integration of a wound-response gene expression signature in predicting breast cancer survival. *Proc Natl Acad Sci U S A* 102:3738–3743
189. Novak K (2005) A healing process. *Nat Rev Cancer* 5(4):244–244
190. Tavazoie SF, Alarcon C, Oskarsson T, Padua D, Wang Q, Bos PD, Gerald WL, Massague J (2008) Endogenous human microRNAs that suppress breast cancer metastasis. *Nature* 451:147–152
191. Midwood KS, Orend G (2009) The role of tenascin-C in tissue injury and tumorigenesis. *J Cell Commun Signal* 3:287–310
192. Fabbri M, Paone A, Calore F, Galli R, Gaudio E, Santhanam R, Lovat F, Fadda P, Mao C, Nuovo GJ, Zanesi N, Crawford M, Ozer GH, Wernicke D, Alder H, Caligiuri MA, Nana-Sinkam P, Perrotti D, Croce CM (2012) MicroRNAs bind to toll-like receptors to induce prometastatic inflammatory response. *Proc Natl Acad Sci U S A* 109:E2110–E2116
193. Sung SY, Hsieh CL, Wu D, Chung LW, Johnstone PA (2007) Tumor microenvironment promotes cancer progression, metastasis, and therapeutic resistance. *Curr Probl Cancer* 31:36–100
194. Lorusso G, Ruegg C (2008) The tumor microenvironment and its contribution to tumor evolution toward metastasis. *Histochem Cell Biol* 130:1091–1103
195. Rofstad EK (2000) Microenvironment-induced cancer metastasis. *Int J Radiat Biol* 76:589–605
196. Margaretten NC, Witschi HR (1988) Effects of hyperoxia on growth of experimental lung metastasis. *Carcinogenesis* 9:433–439

197. Vaupel P, Kallinowski F, Okunieff P (1989) Blood flow, oxygen and nutrient supply, and metabolic microenvironment of human tumors: A review. *Cancer Res* 49:6449–6465
198. Rofstad EK, Halsor EF (2002) Hypoxia-associated spontaneous pulmonary metastasis in human melanoma xenografts: Involvement of microvascular hot spots induced in hypoxic foci by interleukin 8. *Br J Cancer* 86:301–308
199. Rofstad EK, Mathiesen B, Kindem K, Galappathi K (2006) Acidic extracellular pH promotes experimental metastasis of human melanoma cells in athymic nude mice. *Cancer Res* 66:6699–6707
200. Brizel DM, Scully SP, Harrelson JM, Layfield LJ, Bean JM, Prosnitz LR, Dewhirst MW (1996) Tumor oxygenation predicts for the likelihood of distant metastases in human soft tissue sarcoma. *Cancer Res* 56:941–943
201. Budhu A, Forgues M, Ye QH, Jia HL, He P, Zanetti KA, Kammula US, Chen Y, Qin LX, Tang ZY, Wang XW (2006) Prediction of venous metastases, recurrence, and prognosis in hepatocellular carcinoma based on a unique immune response signature of the liver microenvironment. *Cancer Cell* 10:99–111
202. Ben-Eliyahu MBAS (2010) Surgery as a double-edged sword: A clinically feasible approach to overcome the metastasis-promoting effects of surgery by blunting stress and prostaglandin responses. *Cancers* 2:1929–1951
203. Zhao T, Xia WH, Zheng MQ, Lu CQ, Han X, Sun YJ (2008) Surgical excision promotes tumor growth and metastasis by promoting expression of MMP-9 and VEGF in a breast cancer model. *Exp Oncol* 30:60–64
204. Camphausen K, Moses MA, Beecken WD, Khan MK, Folkman J, O'Reilly MS (2001) Radiation therapy to a primary tumor accelerates metastatic growth in mice. *Cancer Res* 61:2207–2211
205. von Essen CF (1991) Radiation enhancement of metastasis: A review. *Clin Exp Metastasis* 9:77–104
206. Nathanson SD, Nelson L, Anaya P, Havstad S, Hetzel FW (1991) Development of lymph node and pulmonary metastases after local irradiation and hyperthermia of footpad melanomas. *Clin Exp Metastasis* 9:377–392
207. Vikram B, Strong EW, Shah JP, Spiro R (1984) Failure at distant sites following multimodality treatment for advanced head and neck cancer. *Head Neck Surg* 6:730–733
208. Demicheli R, Retsky MW, Hrushesky WJ, Baum M, Gukas ID (2008) The effects of surgery on tumor growth: A century of investigations. *Ann Oncol* 19:1821–1828
209. Pidgeon GP, Harney JH, Kay E, Da CM, Redmond HP, Bouchier-Hayes DJ (1999) The role of endotoxin/lipopolysaccharide in surgically induced tumour growth in a murine model of metastatic disease. *Br J Cancer* 81:1311–1317
210. Al-Sahaf O, Wang JH, Browne TJ, Cotter TG, Redmond HP (2010) Surgical injury enhances the expression of genes that mediate breast cancer metastasis to the lung. *Ann Surg* 252:1037–1043
211. Yan T, Yin W, Zhou L, Jiang Y, Shen Z, Shao Z, Lu J (2010) Postoperative fever: The potential relationship with prognosis in node negative breast cancer patients. *PLoS One* 5:e15903
212. Nowacki MP, Szymendera JJ (1983) The strongest prognostic factors in colorectal carcinoma. Surgicopathologic stage of disease and postoperative fever. *Dis Colon Rectum* 26:263–268
213. Lutgendorf SK, Sood AK, Antoni MH (2010) Host factors and cancer progression: Biobehavioral signaling pathways and interventions. *J Clin Oncol* 28:4094–4099
214. Li S, Sun Y, Gao D (2013) Role of the nervous system in cancer metastasis. *Oncol Lett* 5:1101–1111
215. Moreno-Smith M, Lutgendorf SK, Sood AK (2010) Impact of stress on cancer metastasis. *Future Oncol* 6:1863–1881
216. Wu W, Yamaura T, Murakami K, Murata J, Matsumoto K, Watanabe H, Saiki I (2000) Social isolation stress enhanced liver metastasis of murine colon 26-L5 carcinoma cells by suppressing immune responses in mice. *Life Sci* 66:1827–1838
217. Wu W, Yamaura T, Murakami K, Ogasawara M, Hayashi K, Murata J, Saiki I (1999) Involvement of TNF-alpha in enhancement of invasion and metastasis of colon 26-L5 carcinoma cells in mice by social isolation stress. *Oncol Res* 11:461–469
218. Sloan EK, Priceman SJ, Cox BF, Yu S, Pimentel MA, Tanganangkul V, Arevalo JM, Morizono K, Karanikolas BD, Wu L, Sood AK, Cole SW (2010) The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res* 70:7042–7052
219. Chiang AC, Massague J (2008) Molecular basis of metastasis. *N Engl J Med* 359:2814–2823
220. Benish M, Bartal I, Goldfarb Y, Levi B, Avraham R, Raz A, Ben-Eliyahu S (2008) Perioperative use of beta-blockers and COX-2 inhibitors may improve immune competence and reduce the risk of tumor metastasis. *Ann Surg Oncol* 15:2042–2052
221. Ewertz M, Jensen MB, Gunnarsdottir KA, Hojris I, Jakobsen EH, Nielsen D, Stenbygaard LE, Tange UB, Cold S (2011) Effect of obesity on prognosis after early-stage breast cancer. *J Clin Oncol* 29:25–31
222. Sestak I, Distler W, Forbes JF, Dowsett M, Howell A, Cuzick J (2010) Effect of body mass index on recurrences in tamoxifen and anastrozole treated women: An exploratory analysis from the ATAC trial. *J Clin Oncol* 28:3411–3415
223. Sinicrope FA, Dannenberg AJ (2011) Obesity and breast cancer prognosis: Weight of the evidence. *J Clin Oncol* 29:4–7
224. Nicolson GL (1988) Organ specificity of tumor metastasis: Role of preferential adhesion, invasion and growth of malignant cells at specific secondary sites. *Cancer Metastasis Rev* 7:143–188
225. Okada F, Shionoya H, Kobayashi M, Kobayashi T, Tazawa H, Onuma K, Iuchi Y, Matsubara N, Ijichi T, Dugas B, Hosokawa M (2006) Prevention of inflammation-mediated acquisition of metastatic properties of benign mouse fibrosarcoma cells by administration of an orally available superoxide dismutase. *Br J Cancer* 94:854–862
226. Fidler IJ, Balasubramanian K, Lin Q, Kim SW, Kim SJ (2010) The brain microenvironment and cancer metastasis. *Mol Cells* 30:93–98
227. Seike T, Fujita K, Yamakawa Y, Kido MA, Takiguchi S, Teramoto N, Iguchi H, Noda M (2011) Interaction between lung cancer cells and astrocytes via specific inflammatory cytokines in the microenvironment of brain metastasis. *Clin Exp Metastasis* 28:13–25
228. Wang N, Thuraingam T, Fallavollita L, Ding A, Radzioch D, Brodt P (2006) The secretory leukocyte protease inhibitor is a type 1 insulin-like growth factor receptor-regulated protein that protects against liver metastasis by attenuating the host proinflammatory response. *Cancer Res* 66:3062–3070
229. Pantel K, Brakenhoff RH (2004) Dissecting the metastatic cascade. *Nat Rev Cancer* 4:448–456
230. Alix-Panabieres C, Riethdorf S, Pantel K (2008) Circulating tumor cells and bone marrow micrometastasis. *Clin Cancer Res* 14:5013–5021
231. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, MacDonald DD, Jin DK, Shido K, Kerns SA, Zhu Z, Hicklin D, Wu Y, Port JL, Altorki N, Port ER, Ruggero D, Shmelkov SV, Jensen KK, Rafii S, Lyden D (2005) VEGFR1-Positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature* 438:820–827
232. Kaplan RN, Psaila B, Lyden D (2006) Bone marrow cells in the 'pre-metastatic niche': Within bone and beyond. *Cancer Metastasis Rev* 25:521–529
233. Herzog EL, Chai L, Krause DS (2003) Plasticity of marrow-derived stem cells. *Blood* 102:3483–3493
234. Uccelli A, Moretta L, Pistoia V (2008) Mesenchymal stem cells in health and disease. *Nat Rev Immunol* 8:726–736
235. Rojas M, Xu J, Woods CR, Mora AL, Spears W, Roman J, Brigham KL (2005) Bone marrow-derived mesenchymal stem cells in repair of the injured lung. *Am J Respir Cell Mol Biol* 33:145–152

236. Yamada M, Kubo H, Kobayashi S, Ishizawa K, Numasaki M, Ueda S, Suzuki T, Sasaki H (2004) Bone marrow-derived progenitor cells are important for lung repair after lipopolysaccharide-induced lung injury. *J Immunol* 172:1266–1272
237. Kallis YN, Alison MR, Forbes SJ (2007) Bone marrow stem cells and liver disease. *Gut* 56:716–724
238. Mareel M, Oliveira MJ, Madani I (2009) Cancer invasion and metastasis: Interacting ecosystems. *Virchows Arch* 454:599–622
239. Allawi Z, Cuzick J, Baum M (2011) Does trauma or an intercurrent surgical intervention lead to a short-term increase in breast cancer recurrence rates? *Ann Oncol*
240. Holmes MD, Chen WY, Li L, Hertzmark E, Spiegelman D, Hankinson SE (2010) Aspirin intake and survival after breast cancer. *J Clin Oncol* 28:1467–1472
241. Motoyama S, Miura M, Hinai Y, Maruyama K, Usami S, Saito H, Minamiya Y, Satoh S, Murata K, Suzuki T, Ogawa J (2009) CRP genetic polymorphism is associated with lymph node metastasis in thoracic esophageal squamous cell cancer. *Ann Surg Oncol* 16: 2479–2485
242. Clinchy B, Fransson A, Druvefors B, Hellsten A, Hakansson A, Gustafsson B, Sjodahl R, Hakansson L (2007) Preoperative interleukin-6 production by mononuclear blood cells predicts survival after radical surgery for colorectal carcinoma. *Cancer* 109: 1742–1749
243. Alcover J, Filella X, Luque P, Molina R, Izquierdo L, Auge JM, Alcaraz A (2010) Prognostic value of IL-6 in localized prostatic cancer. *Anticancer Res* 30:4369–4372
244. Ganz PA, Cole SW (2011) Expanding our therapeutic options: Beta blockers for breast cancer? *J Clin Oncol* 29:2612–2616
245. Barron TI, Connolly RM, Sharp L, Bennett K, Visvanathan K (2011) Beta blockers and breast cancer mortality: A population-based study. *J Clin Oncol* 29:2635–2644