REVIEW

Tissue-Specific Homing of Immune Cells in Malignant Skin Tumors

Hajnalka Jókai • Márta Marschalkó • Judit Csomor • József Szakonyi • Orsolya Kontár • Gábor Barna • Sarolta Kárpáti • Péter Holló

Received: 7 November 2011 / Accepted: 27 March 2012 / Published online: 24 April 2012 © Arányi Lajos Foundation 2012

Abstract Tissue-specific migration of immune cells involved both in physiological and pathological immune responses is a current research subject for medical science. Several homing molecules have been identified orchestrating extravasation of immune cells to certain peripheral nonlymphoid tissues such as gut, lung and skin. Regarding lymphocyte homing to skin, the first-line defense of human body cutaneous lymphocyte associated antigen (CLA) and a group of chemokine-chemokine receptor pairs are considered to be of crucial importance. The aim of the present review is to summarize existing knowledge about skin- and tumor-specific migration of immune cells playing a major pathogenetic role in host immune responses induced by nonlymphoid malignant skin tumors as well as in the development of primary cutaneous T-cell lymphomas (CTCL). Melanoma malignum, squamous and basal cell carcinoma evoke host immune responses and consequently a subset of reactive immune cells is recruited to site of the tumor. Regarding migratory process and exact functional role of these cells a

H. Jókai (⊠) • M. Marschalkó • J. Szakonyi • O. Kontár •
S. Kárpáti • P. Holló
Department of Dermatovenerology and Dermatooncology,
Semmelweis University,
Mária u. 41,
Budapest 1085, Hungary
e-mail: jokaihajnalka@gmail.com
J. Csomor • G. Barna
1st Institute of Pathology and Experimental Cancer Research,
Semmelweis University,
Üllői út 26,
Budapest 1085, Hungary
S. Kárpáti

Hungarian Academy of Sciences, Molecular Research Group, Mária utca 41, Budapest 1085, Hungary growing number of data is available in literature. On the other hand tissue-specific immune cell homing is regarded as a key process in the pathogenesis of CTCL where malignant T-lymphocytes can be found in circulation and symptomatic skin. Hereby homing mechanism of malignant T-cells in mycosis fungoides and Sézary-syndrome as separate clinical entities of CTCL is discussed. A precise insight into the molecular background of skin- and tumor-specific immune cell migration can contribute to developing efficient vaccine therapies in non-lymphoid malignant skin tumors and beneficial treatment modalities in CTCL.

Keywords Lymphocyte homing receptors · Cutaneous lymphocyte associated antigen · Malignant melanoma · Basal cell carcinoma · Squamous cell carcinoma · Cutaneous T-cell lymphoma

Abbreviations

Tem	effector memory T lymphocytes
Tcm	central memory T lymphocytes
CLA	cutaneous lymphocyte associated antigen
PSGL-1	P-selectin glycoprotein ligand 1
DC	dendritic cells
TILs	tumor infiltrating lymphocytes
Treg cells	regulatory T cells
LFA-1	leukocyte function associated antigen 1
VLA-4	very late antigen 4
ICAM	intercellular adhesion molecule
VCAM	vascular cell adhesion molecule
VEGF	vascular endothelial growth factor
TGF	fibroblast growth factor
SCC	squamous cell carcinoma
iNOS	inducible NO synthetase
BCC	basal cell carcinoma
CTCL	cutaneous T cell lymphoma

SS	Sézary-syndrome	
MF	mycosis fungoides	
SDF-1	stromal cell-derived factor 1	

Introduction

Tissue-specific homing of immune cells is involved in both physiological and pathological immune mediated processes [1–4]. Homing properties of effector memory T-lymphocytes (Tem) are mostly under investigation as in case of antigenrechallenge these cells are uniquely able to migrate in great numbers to a certain peripheral non-lymphoid tissue where they have previously encountered the antigen. Cytokines secreted by antigen presenting cells influence the expression of cell-surface homing markers on newly activated lymphocytes [5–8]. Regarding skin, the outermost barrier of human body the most specific homing molecule of Tem is known as cutaneous lymphocyte associated antigen (CLA) [6, 9-11]. CLA is a carbohydrate epitope of the cell surface protein Pselectin glycoprotein ligand 1 (PSGL-1) and binds E-selectin and P-selectin, both adhesion receptors expressed on endothelium of skin microvasculature [10, 12]. The expression of CLA is supposed to be strictly controlled. IL-12, a cytokine secreted by dendritic cells (DCs) and mediating Th1 differentiation of naiv T-cells is a major stimulatory factor of CLA expression in vivo. In contrast IL-4 promoting the Th2 pathway shows negative correlation with the rate of CLA expression [9]. However, CLA is not exclusively expressed on Th1 effector memory cells but also on Th2, Th17 lymphocytes and in a small percentage even on B-lymphocytes as well as peripheral monocytes and granulocytes [11]. In the local microenvironment of antigen presentation soluble molecules (eg. vitamin D, A) originating from skin or lymph node contribute to generation of the homing pattern [9, 13-15]. The coexpression of CLA and certain chemokine receptors is needed to a perfect skin migratory process. Chemokine receptors are expressed mainly on lymphocytes, monocytes, neutrophils and eosinophils. They selectively bind their matching chemokines, a group of small (~8-14 kDa), structurally related molecules that regulate tissue-specific migration of leukocytes [16]. Chemokine receptors expressed on skin-homing lymphocytes are attracted by chemokines produced in skin cells. Based on existing data, following chemokine receptorchemokine pairs are involved in skin-specific migration of T cells: CCR4-CCL17, CCL22; CCR6-CCL20; CCR10-CCL27 and CXCR3-CXCL9, CXCL10, CXCL11 [5, 17, 18]. The migration happens through a stepwise process initiated by a highly specific but loose interaction between CLA and E-selectin. Their binding enables the chemokine mediated activation of several non-specific adhesion molecules on immune and endothelial cells resulting in a firm, irreversible adhesion, a subsequent extravasation and migration to the site of inflammation or malignant cell proliferation [9, 19, 20] (Fig. 1). Hereby we give a summary on details of this migratory process that might greatly determine the pathogenesis and the clinical outcome of malignant skin tumors including nonlymphoid neoplasias and certain forms of cutaneous T-cell lymphomas.

Tumor-Specific Homing of Immune Cells in Melanoma

It was postulated years ago that there is a selective accumulation of immune cells with a tumor- and tissue-specific phenotype in malignant skin tumors [21]. The presence of tumor infiltrating lymphocytes (TILs) in primary cutaneous melanoma was first described by Clark et al. in 1969 [22] (Fig. 2). Later several studies demonstrated a prognostic significance of the number of these cells. As increased number of TILs in the tumor mass and microenvironment could be connected to a better clinical outcome, it was supposed that all lymphocytes migrated to the tumor burden exert antitumor functions [23-26]. Subsequently it became obvious that TILs don't represent a homogenous group. Beside CD4+ and CD8+ conventional $\alpha\beta$ T-lymphocytes, NK-cells and $\gamma\delta$ T-cells, all with antitumor activity, also CD4+CD25+Foxp3+ regulatory T-cells (Treg-cells) are present in melanoma lesions. Treg-cells own immunoinhibitory properties and promote tumor progression by hampering tumor-associated reactive lymphocytes [27-30]. Weishaupt et al. examined molecular and cellular changes in metastasizing melanoma and suggested that increased number of Treg-cells found both within and near the tumorbed is a hallmark of tumor progression [31]. Based on these findings the prognostic role of TILs is not determined by their number but rather the complexity of their different functions [27].

Antitumor or tumor promoting function of TILs is initiated by their tissue- and tumorspecific recruitment to the tumor mass. This complex homing and transmigratory process is enabled by a certain group of adhesion molecules as well as skin- and tumorspecific homing receptors. Additionally, differences in TIL-homing to primary tumors and metastatic lesions are of special interest as advantage of T-cell adoptive cancer therapy, the most promising targeted immunotherapy, is mostly expected in advanced, metastatic melanoma [32–34].

Adams et al. analyzed the cell-surface phenotype of TILs cultured from human melanoma in order to understand their tumor-homing properties. They found a significant upregulation of the skin-homing receptor cutaneous lymphocyte associated antigen (CLA) which was proven to be independent from culture conditions and rather a characteristic feature of in vivo generated TILs [35]. Based on the finding

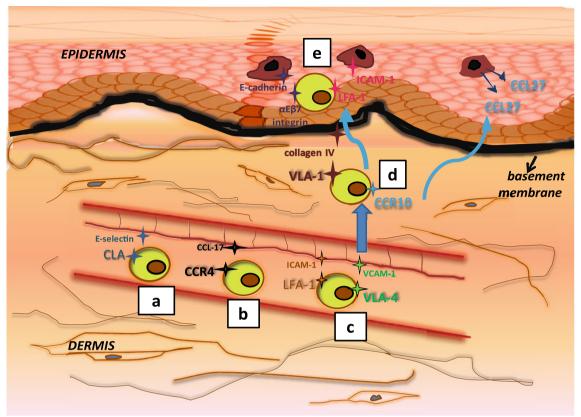


Fig. 1 Lymphocyte extravasation to the skin a) specific but loose CLA–E-selectin binding \rightarrow ROLLING of T-cells along vessel wall b) binding of CCR4 (on T-cell) to CCL17 (on endothelial cell); coexpression of CLA and CCR4 on T-cells is responsible for skin-specific homing. c) LFA-1--ICAM-1, VLA-4--VCAM-1 binding (firm, but less specific interaction) \rightarrow extravasation d) VLA-1 on T-cell binds to

that on melanoma vasculature there is a moderate expression of CLA-binding E-selectin, they suggested a role of the interaction of these two molecules in tumor-specific homing of TILs [35, 36]. Importance of the interaction between

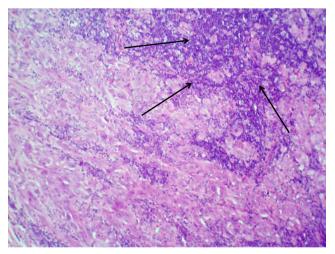


Fig. 2 Histopathological picture of melanoma malignum. Marking shows the presence of tumor infiltrating lymphocytes

collagen IV of the basement membrane \rightarrow T-cells migrate to the epidermis; CCR10+ T-lymphocytes are attracted to the epidermis by keratinocyte-secreted CCL27 e) cell-surface $\alpha E\beta 7$ integrin on epidermal T-cells binds to E-cadherin on keratinocytes; LFA-1 on epidermal T-cells binds to ICAM-1 integrin on keratinocytes. \rightarrow T-cells are captured in epidermis

CLA and E-, P-selectin is underlined by several studies focusing on molecular differences between primary melanoma and advanced melanoma with metastases. Several authors found that the overall moderate expression rate of tumor endothelial adhesion molecules E- and P-selectin decreases over progression which in turn inhibits the migration of CLA+TILs to the tumor [31, 37, 38]. Beside the highly specific binding of CLA and E-, P-selectin other, less-specific adhesion molecule pairs also influence the homing ability of TILs. Microvasculature of metastatic melanoma can not upregulate adhesion molecules ICAM-1 (intercellular adhesion molecule) and VCAM-1 (vascular cell adhesion molecule) and is therefore less permeable to tumor-reactive lymphocytes expressing counter-ligands LFA-1 (leukocyte function associated antigen) and VLA-4 (very late antigen), respectively [39]. The expression of endothelial adhesion receptors might be modulated by VEGF (vascular endothelial growth factor) and TGF (tumor fibroblast growth factor) that are potent stimulators of angiogenesis [31, 40-42]. The lack of adhesion molecules on the tumor microvasculature can be a plausible explanation why Weishaupt et al. found T cells with an effector, lytic

function present in a small proportion in melanoma metastases and accumulated in great numbers in the closely surrounding tissue where in contrast a strong expression of the above mentioned adhesion receptors were noted. Similarly, mature antigen presenting dendritic cells with antitumor activity were localized to the peritumoral areas [31].

Gelb et al. found that CLA+T-lymphocytes are present in primary melanoma malignum, but they are undetectable in tumor metastases, including even cutaneous metastatic lesions. They supposed that only primary tumors (not cutaneous and extracutaneous metastases) are associated with the involvement (or irritation) of healthy epidermis which seems to be necessary to the recruitment of CLA+immune cells [21]. In inflammatory skin lesions keratinocytes secrete the cytokines TNF- α (tumor necrosis factor α) and IL-1 (interleukin-1) that are able to effectively upregulate the expression of CLA binding E-selectin on vascular endothelial cells, thereby promoting the skin-homing of inflammatory CLA+T cells. Additional keratinocyte expressed cytokines serve as chemoattractants for these T-lymphocytes [7, 21, 43]. Malignant tumor cells must lack some of these required soluble factors and tumor-specific CLA+lymphocyte accumulation is therefore possible only with the involvement of healthy but irritated epidermis [21]. The irritation is caused by epidermal invasion of the tumor cells that slightly damages surrounding healthy cells. This event leads to an inflammatory condition in primary tumors with the recruitment of immune cells that target malignant tumor cells [44]. Similarly, Weishaupt et al. found that T-cells trafficking to melanoma metastases are not bearing skin- and lymph node-specific homing receptors [31].

Several studies investigating host antitumor responses suggest that beside common and tissue-specific adhesion molecules also cytokines and chemokine-chemokine receptor pairs play an additional role in guiding of tumor infiltrating immune cells. Cytokine and chemokine synthesis is attributed to melanoma cells and additional cell types in the tumor microenvironment, such as macrophages, endothelial cells and recruited T-cells [45-48]. Key chemokines and corresponding chemokine receptors involved in the homing of melanoma infiltrating immune cells are summarized in Table 1. Angiostatic chemokines CXCL9, 10, 11 recruit reactive Th1-lymphocytes expressing chemokine receptor CXCR3 [44, 49-51]. An associtation was demonstrated between strong expression of the chemokine CXCL9 and a great number of infiltrating T-lymphocytes in human malignant melanoma [52]. Graves et al. first described the production of chemokine CCL2 by melanoma cells [53]. Secretion of CCL5 by melanoma cells is suggested as a potential tumor escape mechanism as CCL5 was found to induce apoptosis of tumor infiltrating CD8+ T-lymphocytes [54]. Expression of CCL20 in malignant melanoma leads to accumulation of mature DCs and effector T-cells in the tumor mass, thereby promoting antitumor host response [55].

In spite of the generally low number or absence of TILs in advanced-stage melanoma individual metastatic lesions occasionally can be rich in successfully migrated lymphocytes. One plausible explanation of this fact is that several oncogenic pathways (eg. Ras, B-Raf, Akt, Notch, Stat3) result in a malignant phenotype from the melanocyte lineage [45, 56–59]. The different signaling pathways can differently influence the expression of chemokine genes as well as other factors contributing to the tumor-specific migration of immune cells [45].

Mechanisms of Homing in Epithelial Skin Cancers (Squamous Cell Carcinoma, Basal Cell Carcinoma)

The hypothesis that effector and effector memory Tlymphocytes are important in controlling skin tumor growth is certified even by the fact that incidence of squamous cell carcinoma (SCC) is notably increased among transplant recipients on immunosuppressive therapy. Even therefore it is of special interest why also tumor reactive T-cells in patients with manifested SCC and without immunosuppression are unable to inhibit tumor manifestation [60]. A possible explanation is the significant downregulation of vascular E-selectin in human SCC and consquently, the failure of antitumor CLA+T-cells to sufficiently migrate to the tumor tissue - a similar event as observed in advanced melanoma. NO (nitric oxide) secreted by iNOS (inducible nitric oxide synthase)-expressing dendritic cells within tumorbed might contribute to impaired ability of endothelial cells to express E-selectin [61]. However, Hald et al. found that head and neck SCC contains numerous T-lymphocytes with the predominance of CD8+ cytotoxic T-cells [62]. In the study of Clark et al.⁶² more than 50 % of the infiltrating T-cells were CD4+CD25+Foxp3+ Treg-cells suppressing antitumor effector lymphocytes. As a difference from normal Treg cells they lacked the expression of skin addressins CLA and CCR4 while expressed lymph node homing markers (L-selectin, CCR7) and resembled the central memory T-cell phenotype this way. However, SCC vessels expressed neither L-selectin ligands nor the chemokine pairs of CCR7 (CCL19, CCL21) [61]. Now tumor cells are considered to elaborate certain chemotactic factors that specifically attract Treg cells [27, 63]. Figure 3 shows histopathological picture of SCC with the presence of TILs.

Regarding immune cell infiltration of basal cell carcinoma (BCC) only a limited amount of data is available in the literature. Successful administration of topically applied immune response modifier imiquimod induced to get more attention on the topic of potential host immune response mechanisms in this semimalignant skin tumor. Tumor cells

Table 1 Main chemokines and corresponding chemokine receptors involved in immune cell infiltration of melanoma malignum

Chemokines	Corresponding chemokine receptors	Recruited immune cells due to binding of chemokine-chemokine receptor (under physiological/inflammatory conditions)	Special importance of chemokine-chemokine receptor binding in melanoma malignum
CCL2	CCR2	Th1, Th2, CD8+ CTL, NK-cells, monocytes, DC, iDC("inflammatory"conditions, eg. atherosclerosis)	tumor promoting role: recruitment of myeloid suppressor cells, consequent inhibition of reactive immune cells; accumulation of monocytes/ macrophages: angiogenesis, matrix degradation tumor inhibiting role: recruitment of reactive, inflammatory immune cells with antitumor function
CCL3	CCR1	iDC, T-cells, NK-cells, monocytes/macrophages (adhesion to activated vascular endothelium, transendothelial migration; allograft rejection; granuloma formation)	migration of tumor infiltrating immune cells with antitumor properties to the melanoma tumor mass
CCL4	CCR5 CCR5	naive CD8+, CD8+, CTL, Th1, Treg, NK-cells, DC, iDC (recruitment of immune cells to infectious foci, viral infections, developing	tumor promoting role: angiogenesis, metastases tumor inhibiting role: recruitment of tumor infiltrating immune cells with antitumor properties
CCL5	CCR5	granulomas; mainly cellular immune response enhancement)	enhancement of tumor progression by inducing apoptosis of tumor infiltrating lymphocytes
	CCR1	iDC, T-cells, NK-cells, monocytes/macrophages (mainly promoting Th1 mediated inflammatory cellular immune responses; granuloma formation)	accumulation of immune cells with antitumor properties in the tumor mass
	CCR3	T-cells, monocytes, DC, neutrophils, eosinophils (inflammatory immune responses, eosinophil recruitment in atopic dermatitis)	migration of immune cells with antitumor properties to the tumor mass
CCL17	CCR4	skin-homing CD4+ T-cells (Th2, Th17, Treg) (skin-specific migration under homeostatic and inflammatory conditions)	migration of skin-homing T-cells to the tumor mass (both inflammatory and regulatory T-lymphocytes involved)
CCL20	CCR6	Th17, B-cells, iDC, Treg (lymphorganogenesis, inflammatory immune responses)	migration of iDC that contribute to efficient antitumor response after maturation; recruitment of regulatory T-cells with tumor promoting function
CCL22	CCR4	skin-homing CD4+ T-cells (Th2, Th17, Treg) (skin-specific migration under homeostatic and inflammatory conditions)	migration of skin-homing T-cells to the tumor mass (both inflammatory and regulatory T-lymphocytes involved)
CCL27	CCR10	skin-homing CD8+ and CD4+ T-cells (skin-specific migration under homeostatic and inflammatory conditions)	migration of skin-homing antitumor T-lymphocytes to the tumor mass
CXCL9	CXCR3	Th1, Th17, CD8+ T-cells, NK-cells	tumor promoting role: development
CXCL10	CXCR3	(infection provoked inflammation;	of metastases tumor inhibiting role:
CXCL11	CXCR3	inflammatory skin diseases; allograft rejection; atherosclerosis)	accumulation of inflammatory T-cells with antitumor function; angiostatic effect

DC dendritic cells, *iDC* immature dendritic cells, *NK-cells* natural killer cells, *Th1* T helper 1, *Th2* T helper 2, *Th17* T helper 17, *CTL* cytotoxic T-lymphocytes, *Treg* regulatory T-cells

of BCC often express HLA class I molecules and are immunogenic this way [64]. Similar to melanoma and SCC there is a significant downregulation of adhesion molecules in tumor microvasculature that results in decreased ability of immune cells to migrate to the tumor burden [65–68]. Multiple action of imiquimod contains enhanced expression of ICAM-1 on tumor endothelial cells mediated by upregulated proinflammatory cytokines (eg. IL-1, TNF- α , IFN- γ) [65, 69]. According to several investigatory results early cellular response after imiquimod treatment includes infiltration of mostly CD4+ T-lymphocytes, activated dermal dendritic cells and macrophages [70–72]. CD8+ T-cells appear only in a later phase and they are suggested to be rather suppressor than cytotoxic cells [70, 71]. Urosevic et al. reported the predominance of CD4+ T-cells before treatment and a reversal of CD4/CD8 ratio after imiquimod therapy, proposing a substantial role of infiltrating CD8+ T-cells in tumor elimination [65]. In BCC a significant number of CD4+ CD25+Foxp3+ Treg-cells and immature dermal dendritic cells was found surrounding the tumor bed, suggesting that

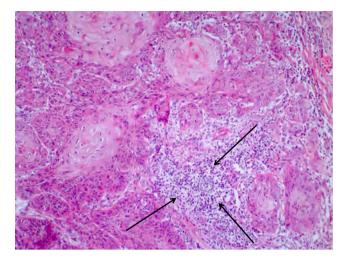


Fig. 3 Histopathological picture of squamous cell carcinoma. Marking shows the presence of tumor infiltrating lymphocytes

they contribute to insufficient anti-tumor response. The expression of CCL22, a Treg chemokine is also enhanced compared to normal skin.[73]

Skin-Homing of Malignant Lymphocytes in Primary Cutaneous T-cell Lymphomas: Mycosis Fungoides and Sézary-Syndrome

Cutaneous T-cell lymphomas (CTCL) represent a heterogenous group of non-Hodgkin lymphomas with the common presence of transformed skin-homing T-cells. Mycosis fungoides (MF) and Sézary-syndrome (SS) are epidermotrop variants of CTCL. Phenotypic features and origin of CTCLcells are of special importance as they are regarded to greatly determine functional and migratory abilities of malignant lymphocytes.

Regarding phenotypic characteristics transformed lymphocytes generally exhibit a Th2 phenotype - supported by their secreted cytokine profile (eg. IL-4, IL-5, IL-10, IL-13) and expression pattern of cell-surface chemokine receptors (CCR4). However, the observation that SS and advanced-stage MF is often associated with clinically evident immunosuppression, raised the question whether proliferating CTCL-cells can originate from lymphocytes with Treg-cell phenotype that repress healthy immune cells [74–76]. Due to current data in early-stage MF there is a significant skin infiltration of normal Treg-cells and Th1 reactive lymphocytes that control the growth of CTCLcells secreting mainly Th2 cytokines [74, 77, 78]. In fairly advanced disease there is a significant decrease in the amount of all types of cytokines which may refer to a Treg phenotype of tumor cells. As malignant Treg-cells CTCLcells don't secrete many cytokines and suppress the productive capacity of reactive lymphocytes as well. However, whether tumor T-cells are derived from skin-homing Tregcells or CD4+ Th2 memory cells that gain Treg-like functions over progression is still a matter of debate [77]. SS as a highly aggressive form of CTCL is consistently Th2 predominant, although several authors suggested a Treg phenotype of Sézary-cells [79–81]. Recently CD158k/KIR3DL2 (member of MHC class I antigen receptors) has been suggested to be a highly specific marker for Sézary-cells [82].

In connection with the *origin* of CTCL-cells novel evidence suggests that MF and SS are diverse presentations of CTCL [77, 83, 84]. Although sharing several molecular pathways important in the pathogenesis, they fundamentally show different clinical behaviour which is presumably the consequence of the fact that they arise from distinct T-cell lines with different homing abilities [20, 83].

MF is characterized by skin-localized patches and infiltrated plaques as well as a favorable clinical outcome [83, 85, 86]. Skin-infiltrating malignant T-cell clones express huge amounts of skin addressins CLA and CCR4 [83]. This cell surface pattern is combined with the lack of lymph node homing markers L-selectin and CCR7, a phenotype suggestive of effector memory lymphocytes (Tem) [20, 78, 83]. Mature Tem-cells generally migrate in a tissue-specific manner and stay permanently in peripheral non-lymphoid tissues without circulating in the blood flow and entering lymph nodes [5, 8, 83]. In the skin they seem to be restricted to fixed locations and secrete inflammatory cytokines recruiting reactive lymphocytes to the site of inflammation. This character has the consequence that in MF patients accumulation of malignant Tem-cells confined to the skin results in stable, inflamed skin lesions persisting for a long time at particular anatomic sites, without involvement of peripheral circulation and a consequent clinical progression [78, 83]. Krieg et al. suggested that in case of progression MF cells lose skin-homing properties and selectively upregulate lymph node homing marker CCR7 [44].

SS is associated with a poor prognosis that seems to be explained by the origin of transformed cells. Circulating tumor cells in SS express high levels of skin-specific chemokine receptor CCR4 but variable amounts of skinhoming markers CLA, CCR6 and CCR10 [83]. They universally coexpress lymph node addressins L-selectin and CCR7, consistent with a central memory phenotype [20, 83, 87-89]. Chemokines attracting CCR7+ lymphocytes are CCL19 and CCL21, constitutively synthesized in lymph nodes [20, 87]. Central memory T-cells (Tcm) usually circulate in the blood flow and regularly visit peripheral lymph nodes. A subset of them migrate even to peripheral nonlymphoid tissues, eg. the skin in a tissue-specific manner [5, 8, 83]. That might be the reason for the detectable number of tumor cells in circulation, the lymphadenopathy and the diffuse erythrodermic skin involvement seen in SS patients [83]. Skin infiltrating malignant lymphocytes in SS were found to express skin-homing markers CLA and CCR4 in a significant percentage which seems to be necessary for the migration to cutaneous areas [83, 90]. A correlation was demonstrated between increased numbers of circulating CLA+lymphoma cells and extent of cutaneous lesions [91, 92].

Skin localization of CTCL-cells is collectively thought to be the consequence of several tissue-specific adhesion molecule pairs such as CLA and E-selectin as well as certain chemokines and chemokine receptors. Chemokine receptors expressed on skin-homing tumor cells enable a directed migration to chemokines produced by epidermal keratinocytes, endothelial cells and epidermal/dermal dendritic cells. Chemokines additionally lead to activation of integrins on leukocytes and their receptors on endothelial surfaces, thereby enabling a firm adhesion and transmigration of malignant lymphocytes to cutaneous tissues [87, 93–95] (Table 2).

Chemokine receptor *CCR4* is highly expressed on CTCL-cells [20, 74, 87, 96]. Corresponding chemokine ligands of CCR4 are *CCL17* and CCL22 [87, 97]. CCR4 was found to be highly expressed on neoplastic Th2 cells mostly in tumor stage of *MF* [98–100]. In accordance with high CCR4 expression of malignant lymphoma cells in advanced-stage MF *Kakinuma* et al. demonstrated a strong correlation between serum CCL17 levels and disease activity [99]. CCR4+ CTCL-cells form conjugates with CCL17 secreting dendritic cells in MF. These epidermal conjugates known as "Pautrier's microabscesses" are specific histopathological markers for MF [74, 87]. Several authors reported that CCR4 is present on circulating and skin-homing CLA+ CD4+ CTCL-cells in *SS* [89, 94, 95, 100, 101].

Similar to CCR4, *CCR10* is frequently expressed on CLA+skin-homing CTCL-cells [20, 74, 84, 87]. *CCL27* is the specific chemokine of CCR10 [87, 102]. Serum levels of CCL27 are increased in *MF/SS* patients and similar to CCL17 are suggested as a marker of disease activity [103, 104].

Based on their experimental data Hoeller et al. presumed that key chemokines directing CTCL-cells to skin are involved in different steps of the homing process. CCL17 seems to mediate firm arrest of tumor cells in dermal

 Table 2
 Key chemokine receptors and corresponding chemokines involved in skin- homing of CTCL-cells

Chemokine receptors	Chemokines
CCR4	CCL17, CCL22
CCR10	CCL27
CXCR3	CXCL9, CXCL10, CXCL11
CXCR4	CXCL12

microvasculature, whereas CCL27 might play a role in the transmigration [94].

Chemokine receptor *CXCR3* and its corresponding ligands *CCL9*, *CCL10*, *CCL11* are involved in skin-homing of CTCLcells. A high percentage of CXCR3+ malignant lymphocytes is present in early-stage *MF*. CXCR3+ tumor cells are mostly absent in advanced stages, consistent with the hypothesis that altered chemokine receptor expression and decreased skinhoming ability of tumor cells result in clinical progression in MF [84, 102, 105–107]. Fierro et al. reported low expression of CXCR3 on CTCL-cell in *SS* [101] while Notohamiprodjo et al. could detect a strong expression in all biopsy samples from SS patients [84].

Chemokine receptor *CXCR4* is also regarded to play a role in homing of CTCL cells [86, 87, 101, 102]. *CXCL12*, the counter-ligand of CXCR4 is usually cleaved and inactivated by the T-cell membrane-bound protease CD26. Circulating and even skin-infiltrating CTCL cells are characterized by the loss of CD26, which may contribute to their CXCR4 mediated skin recruitment and accumulation [86, 87].

Conclusion and Future Perspectives

Immune cell infiltration and function in malignant skin tumors is a complex process regulated by numerous cell types and several contributing factors in tumor microenvironment.

TILs in non-lymphoid malignant skin tumors exert variable functions and their homing requires tissue-specific homing and adhesion molecules as well as chemokinechemokine receptor pairs. CLA expressed on skin-homing TILs and its corresponding endothelial receptors are regarded as key molecules in this migratory process. In metastatic lesions downregulation of CLA-binding selectins results in decreased amount of tumor infiltrating CLA+ reactive lymphocytes. This molecular change is of special importance in adoptive cancer therapy of metastatic melanoma, where isolated tumor reactive CD8+ T-cells are in vitro expanded and reinfused to the patient: Changes in the homing ability of TILs can greatly hamper the effectiveness of this targeted immunotherapy as reinfused lymphocytes are needed to successfully migrate to the metastatic tumor mass. As altered homing of tumor reactive immune cells is a potential target of melanoma immunotherapies, a growing number of revealed details about lymphocyte migration and its influencing circumstances is indispensable to successfully administered adoptive cancer therapy.

CTCL-cells in MF and SS alike are also guided to cutaneous tissues by the skin addressin CLA and certain chemokine receptors expressed on tumor cells. The number of circulating CLA+CTCL-cells was found to correlate with extension of skin lesions in SS, while serum levels of CTCL-attracting chemokines CCL17 and CCL27 were demonstrated to correlate with disease severity both in MF and SS. Novel evidence suggests that MF and SS arise from distinct cell lines with different homing abilities and are probably therefore characterized by diverse clinical course and prognosis. Getting further insight into the molecular background of lymphoma cell homing is necessary as inhibition of CTCL-migration is expected to bring a new perspective to targeted therapy of CTCL forms.

References

- 1. Kunkel EJ, Butcher EC (2002) Chemokines and the tissuespecific migration of lymphocytes. Immunity 16:1–4
- Hart AL, Ng SC, Mann E, Al-Hassi HO, Bernardo D, Knight SC (2010) Homing of immune cells: role in homeostasis and intestinal inflammation. Inflamm Bowel Dis 16:1969–1977
- Lalor PF, Curbishley SM, Adams DH (2010) Identifying homing interactions in T-cell traffic in human disease. Methods Mol Biol 616:231–252
- Villablanca EJ, Russo V, Mora JR (2008) Dendritic cell migration and lymphocyte homing imprinting. Histol Histopathol 23:897– 910
- Woodland DL, Kohlmeier JE (2009) Migration, maintenance and recall of memory T cells in peripheral tissues. Nat Rev Immunol 9:153–161
- Sigmundsdottir H, Butcher EC (2008) Environmental cues, dendritic cells and the programming of tissue-selective lymphocyte trafficking. Nat Immunol 9:981–987
- Nestle FO, Di Meglio P, Qin JZ, Nickoloff BJ (2009) Skin immune sentinels in health and disease. Nat Rev Immunol 9:679–691
- Clark RA (2010) Skin-resident T cells: the ups and downs of on site immunity. J Invest Dermatol 130(2):362–370
- Sigmundsdottir H (2010) Improving topical treatments for skin diseases. Trends Pharmacol Sci 31:239–245
- Fuhlbrigge RC, King SL, Sackstein R, Kupper TS (2006) CD43 is a ligand for E-selectin on CLA+human T cells. Blood 107:1421– 1426
- Magro CM, Dyrsen ME (2008) Cutaneous lymphocyte antigen expression in benign and neoplastic cutaneous B- and T-cell lymphoid infiltrates. J Cutan Pathol 35:1040–1049
- Ni Z, Walcheck B (2009) Cutaneous lymphocyte-associated antigen (CLA) T cells up-regulate P-selectin ligand expression upon their activation. Clin Immunol 133:257–264
- Yamanaka K, Dimitroff CJ, Fuhlbrigge RC, Kakeda M, Kurokawa I, Mizutani H, Kupper TS (2008) Vitamins A and D are potent inhibitors of cutaneous lymphocyte-associated antigen expression. J Allergy Clin Immunol 121:148–157.e3
- Baeke F, Takiishi T, Korf H, Gysemans C, Mathieu C (2010) Vitamin D: modulator of the immune system. Curr Opin Pharmacol 10:482–496
- Duhen T, Geiger R, Jarrossay D, Lanzavecchia A, Sallusto F (2009) Production of interleukin 22 but not interleukin 17 by a subset of human skin-homing memory T cells. Nat Immunol 10:857–863
- 16. Zlotnik A, Yoshie O (2000) Chemokines: a new classification system and their role in immunity. Immunity 12:121–127
- Sabat R, Philipp S, Höflich C, Kreutzer S, Wallace E, Asadullah K, Volk HD, Sterry W, Wolk K (2007) Immunopathogenesis of psoriasis. Exp Dermatol 16:779–798

- Clark RA, Chong B, Mirchandani N, Brinster NK, Yamanaka K, Dowgiert RK, Kupper TS (2006) The vast majority of CLA+ T cells are resident in normal skin. J Immunol 176:4431– 4439
- Santamaria Babi LF, Moser R, Perez Soler MT, Picker LJ, Blaser K, Hauser C (1995) Migration of skin-homing T cells across cytokine-activated human endothelial cell layers involves interaction of the cutaneous lymphocyte-associated antigen (CLA), the very late antigen-4 (VLA-4), and the lymphocyte functionassociated antigen-1 (LFA-1). J Immunol 154:1543–1550
- Pals ST, de Gorter DJ, Spaargaren M (2007) Lymphoma dissemination: the other face of lymphocyte homing. Blood 110:3102– 3111
- Gelb AB, Smoller BR, Warnke RA, Picker LJ (1993) Lymphocytes infiltrating primary cutaneous neoplasms selectively express the cutaneous lymphocyte-associated antigen (CLA). Am J Pathol 142:1556–1564
- Clark WH Jr, From L, Bernardino EA, Mihm MC (1969) The histogenesis and biologic behavior of primary human malignant melanomas of the skin. Cancer Res 29:705–727
- Clemente CG, Mihm MC Jr, Bufalino R, Zurrida S, Collini P, Cascinelli N (1996) Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. Cancer 77:1303–1310
- 24. Mullins IM, Slingluff CL, Lee JK, Garbee CF, Shu J, Anderson SG, Mayer ME, Knaus WA, Mullins DW (2004) CXC chemokine receptor 3 expression by activated CD8+ T cells is associated with survival in melanoma patients with stage III disease. Cancer Res 64:7697–7701
- 25. Day CL Jr, Sober AJ, Kopf AW, Lew RA, Mihm MC Jr, Hennessey P, Golomb FM, Harris MN, Gumport SL, Raker JW, Malt RA, Cosimi AB, Wood WC, Roses DF, Gorstein F, Postel A, Grier WR, Mintzis MN, Fitzpatrick TB (1981) A prognostic model for clinical stage I melanoma of the upper extremity. The importance of anatomic subsites in predicting recurrent disease. Ann Surg 193:436–440
- 26. Tuthill RJ, Unger JM, Liu PY, Flaherty LE, Sondak VK (2002) Risk assessment in localized primary cutaneous melanoma: a Southwest Oncology Group study evaluating nine factors and a test of the Clark logistic regression prediction model. Am J Clin Pathol 118:504–511
- Oble DA, Loewe R, Yu P, Mihm MC Jr (2009) Focus on TILs: prognostic significance of tumor infiltrating lymphocytes in human melanoma. Cancer Immun 9:3
- Taylor RC, Patel A, Panageas KS, Busam KJ, Brady MS (2007) Tumor-infiltrating lymphocytes predict sentinel lymph node positivity in patients with cutaneous melanoma. J Clin Oncol 25:869–875
- Prieto PA, Durflinger KH, Wunderlich JR, Rosenberg SA, Dudley ME (2010) Enrichment of CD8+ cells from melanoma tumorinfiltrating lymphocyte cultures reveals tumor reactivity for use in adoptive cell therapy. J Immunother 33:547–556
- 30. Viguier M, Lemaître F, Verola O, Cho MS, Gorochov G, Dubertret L, Bachelez H, Kourilsky P, Ferradini L (2004) Foxp3 expressing CD4+CD25(high) regulatory T cells are overrepresented in human metastatic melanoma lymph nodes and inhibit the function of infiltrating T cells. J Immunol 173:1444–1453
- Weishaupt C, Munoz KN, Buzney E, Kupper TS, Fuhlbrigge RC (2007) T-cell distribution and adhesion receptor expression in metastatic melanoma. Clin Cancer Res 13:2549–2556
- Rosenberg SA, Restifo NP, Yang JC, Morgan RA, Dudley ME (2008) Adoptive cell transfer: a clinical path to effective cancer immunotherapy. Nat Rev Cancer 8:299–308
- Labarriere N, Dreno B, Jotereau F (2008) Lymphocyte biomarkers of clinical responses to adoptive immunotherapy of malignant melanoma. Curr Cancer Ther Rev 4:125–129

- Gattinoni L, Powell DJ Jr, Rosenberg SA, Restifo NP (2006) Adoptive immunotherapy for cancer: building on success. Nat Rev Immunol 6:383–393
- 35. Adams DH, Yannelli JR, Newman W, Lawley T, Ades E, Rosenberg SA, Shaw S (1997) Adhesion of tumour-infiltrating lymphocytes to endothelium: a phenotypic and functional analysis. Br J Cancer 75:1421–1431
- 36. Rohde D, Schlüter-Wigger W, Mielke V, von den Driesch P, von Gaudecker B, Sterry W (1992) Infiltration of both T cells and neutrophils in the skin is accompanied by the expression of endothelial leukocyte adhesion molecule-1 (ELAM-1): an immunohistochemical and ultrastructural study. J Invest Dermatol 98:794–799
- 37. Schadendorf D, Heidel J, Gawlik C, Suter L, Czarnetzki BM (1995) Association with clinical outcome of expression of VLA-4 in primary cutaneous malignant melanoma as well as Pselectin and E-selectin on intratumoral vessels. J Natl Cancer Inst 87:366–371
- Nooijen PT, Westphal JR, Eggermont AM, Schalkwijk C, Max R, de Waal RM, Ruiter DJ (1998) Endothelial P-selectin expression is reduced in advanced primary melanoma and melanoma metastasis. Am J Pathol 152:679–682
- Quezada SA, Peggs KS, Simpson TR, Shen Y, Littman DR, Allison JP (2008) Limited tumor infiltration by activated T effector cells restricts the therapeutic activity of regulatory T cell depletion against established melanoma. J Exp Med 205:2125– 2138
- 40. Walshe TE, Dole VS, Maharaj AS, Patten IS, Wagner DD, D'Amore PA (2009) Inhibition of VEGF or TGF-{beta} signaling activates endothelium and increases leukocyte rolling. Arterioscler Thromb Vasc Biol 29:1185–1192
- Walshe TE (2010) TGF-beta and microvessel homeostasis. Microvasc Res 80:166–173
- 42. Kiss J, Tímár J, Somlai B, Gilde K, Fejôs Z, Gaudi I, Ladányi A (2007) Association of microvessel density with infiltrating cells in human cutaneous malignant melanoma. Pathol Oncol Res 13:21–31
- Barton GM (2008) A calculated response: control of inflammation by the innate immune system. J Clin Invest 118:413–420
- 44. Krieg C, Boyman O (2009) The role of chemokines in cancer immune surveillance by the adaptive immune system. Semin Cancer Biol 19:76–83
- 45. Harlin H, Meng Y, Peterson AC, Zha Y, Tretiakova M, Slingluff C, McKee M, Gajewski TF (2009) Chemokine expression in melanoma metastases associated with CD8+ T-cell recruitment. Cancer Res 69:3077–3085
- 46. Navarini-Meury AA, Conrad C (2009) Melanoma and innate immunity–aActive inflammation or just erroneous attraction? Melanoma as the source of leukocyte-attracting chemokines. Semin Cancer Biol 19:84–91
- Richmond A, Yang J, Su Y (2009) The good and the bad of chemokines/chemokine receptors in melanoma. Pigment Cell Melanoma Res 22:175–186
- Somasundaram R, Herlyn D (2009) Chemokines and the microenvironment in neuroectodermal tumor-host interaction. Semin Cancer Biol 19:92–96
- Strieter RM, Belperio JA, Phillips RJ, Keane MP (2004) CXC chemokines in angiogenesis of cancer. Semin Cancer Biol 14:195–200
- Strieter RM, Burdick MD, Gomperts BN, Belperio JA, Keane MP (2005) CXC chemokines in angiogenesis. Cytokine Growth Factor Rev 16:593–609
- 51. Cole KE, Strick CA, Paradis TJ, Ogborne KT, Loetscher M, Gladue RP, Lin W, Boyd JG, Moser B, Wood DE, Sahagan BG, Neote K (1998) Interferon-inducible T cell alpha chemoattractant (I-TAC): a novel non-ELR CXC chemokine with potent activity

on activated T cells through selective high affinity binding to CXCR3. J Exp Med 187:2009-2021

- 52. Kunz M, Toksoy A, Goebeler M, Engelhardt E, Brocker E, Gillitzer R (1999) Strong expression of the lymphoattractant C-X-C chemokine Mig is associated with heavy infiltration of T cells in human malignant melanoma. J Pathol 189:552–558
- Graves DT, Barnhill R, Galanopoulos T, Antoniades HN (1992) Expression of monocyte chemotactic protein-1 in human melanoma in vivo. Am J Pathol 140:9–14
- 54. Mellado M, de Ana AM, Moreno MC, Martinez C, Rodriguez-Frade JM (2001) A potential immune escape mechanism by melanoma cells through the activation of chemokine-induced T cell death. Curr Biol 11:691–696
- 55. Fushimi T, Kojima A, Moore MA, Crystal RG (2000) Macrophage inflammatory protein 3alpha transgene attracts dendritic cells to established murine tumors and suppresses tumor growth. J Clin Invest 105:1383–1393
- 56. Brose MS, Volpe P, Feldman M, Kumar M, Rishi I, Gerrero R, Einhorn E, Herlyn M, Minna J, Nicholson A, Roth JA, Albelda SM, Davies H, Cox C, Brignell G, Stephens P, Futreal PA, Wooster R, Stratton MR, Weber BL (2002) BRAF and RAS mutations in human lung cancer and melanoma. Cancer Res 62:6997–7000
- Robertson GP (2005) Functional and therapeutic significance of Akt deregulation in malignant melanoma. Cancer Metastasis Rev 24:273–285
- Hardy KM, Kirschmann DA, Seftor EA, Margaryan NV, Postovit LM, Strizzi L, Hendrix MJ (2010) Regulation of the embryonic morphogen Nodal by Notch4 facilitates manifestation of the aggressive melanoma phenotype. Cancer Res 70:10340–10350
- Yang CH, Fan M, Slominski AT, Yue J, Pfeffer LM (2010) The role of constitutively activated STAT3 in B16 melanoma cells. Int J Interferon Cytokine Mediator Res 2010:1–7
- 60. Clark RA, Chong BF, Mirchandani N, Yamanaka K, Murphy GF, Dowgiert RK, Kupper TS (2006) A novel method for the isolation of skin resident T cells from normal and diseased human skin. J Invest Dermatol 126:1059–1070
- 61. Clark RA, Huang SJ, Murphy GF, Mollet IG, Hijnen D, Muthukuru M, Schanbacher CF, Edwards V, Miller DM, Kim JE, Lambert J, Kupper TS (2008) Human squamous cell carcinomas evade the immune response by down-regulation of vascular E-selectin and recruitment of regulatory T cells. J Exp Med 205:2221–2234
- Hald J, Rasmussen N, Claesson MH (1995) Tumour-infiltrating lymphocytes mediate lysis of autologous squamous cell carcinomas of the head and neck. Cancer Immunol Immunother 41:243– 250
- Wei S, Kryczek I, Zou W (2006) Regulatory T-cell compartmentalization and trafficking. Blood 108:426–431
- 64. Dummer R, Urosevic M, Kempf W, Hoek K, Hafner J, Burg G (2003) Imiquimod in basal cell carcinoma: how does it work? Br J Dermatol 149(Suppl 66):57–58
- 65. Urosevic M, Maier T, Benninghoff B, Slade H, Burg G, Dummer R (2003) Mechanisms underlying imiquimod-induced regression of basal cell carcinoma in vivo. Arch Dermatol 139:1325–1332
- Verhaegh M, Beljaards R, Veraart J, Hoekzema R, Neumann M (1998) Adhesion molecule expression in basal cell carcinoma. Eur J Dermatol 8:252–255
- 67. Kooy AJ, Prens EP, Van Heukelum A, Vuzevski VD, Van Joost T, Tank B (1999) Interferon-gamma-induced ICAM-1 and CD40 expression, complete lack of HLA-DR and CD80 (B7.1), and inconsistent HLA-ABC expression in basal cell carcinoma: a possible role for interleukin-10? J Pathol 187:351–357
- 68. Kooy AJ, Tank B, Vuzevski VD, van Joost T, Prens EP (1998) Expression of interferongamma receptors and interferon-gammainduced up-regulation of intercellular adhesion molecule-1 in basal cell carcinoma; decreased expression of IFNgamma R and

shedding of ICAM-1 as a means to escape immune surveillance. J Pathol 184:169–176

- 69. Kovach BT, Stasko T (2005) Use of topical immunomodulators in organ transplant recipients. Dermatol Ther 18:19–27
- Barnetson RS, Satchell A, Zhuang L, Slade HB, Halliday GM (2004) Imiquimod induced regression of clinically diagnosed superficial basal cell carcinoma is associated with early infiltration by CD4 T cells and dendritic cells. Clin Exp Dermatol 29:639–643
- 71. De Giorgi V, Salvini C, Chiarugi A, Paglierani M, Maio V, Nicoletti P, Santucci M, Carli P, Massi D (2009) In vivo characterization of the inflammatory infiltrate and apoptotic status in imiquimod-treated basal cell carcinoma. Int J Dermatol 48:312–321
- Salasche S (2002) Imiquimod 5 % cream: a new treatment option for basal cell carcinoma. Int J Dermatol 41(Suppl 1):16–20
- 73. Kaporis HG, Guttman-Yassky E, Lowes MA, Haider AS, Fuentes-Duculan J, Darabi K, Whynot-Ertelt J, Khatcherian A, Cardinale I, Novitskaya I, Krueger JG, Carucci JA (2007) Human basal cell carcinoma is associated with Foxp3+ T cells in a Th2 dominant microenvironment. J Invest Dermatol 127:2391–2398
- 74. Kim EJ, Hess S, Richardson SK, Newton S, Showe LC, Benoit BM, Ubriani R, Vittorio CC, Junkins-Hopkins JM, Wysocka M, Rook AH (2005) Immunopathogenesis and therapy of cutaneous T cell lymphoma. J Clin Invest 115:798–812
- Berger CL, Tigelaar R, Cohen J, Mariwalla K, Trinh J, Wang N, Edelson RL (2005) Cutaneous T-cell lymphoma: malignant proliferation of T-regulatory cells. Blood 105:1640–1647
- Clark RA (2009) Regulation gone wrong: a subset of Sézary patients have malignant regulatory T cells. J Invest Dermatol 129:2747–2750
- 77. Chong BF, Wilson AJ, Gibson HM, Hafner MS, Luo Y, Hedgcock CJ, Wong HK (2008) Immune function abnormalities in peripheral blood mononuclear cell cytokine expression differentiates stages of cutaneous T-cell lymphoma/mycosis fungoides. Clin Cancer Res 14:646–653
- Beyer M, Möbs M, Humme D, Sterry W (2011) Pathogenesis of mycosis fungoides. J Dtsch Dermatol Ges 9:594–598
- Heid JB, Schmidt A, Oberle N, Goerdt S, Krammer PH, Suri-Payer E, Klemke CD (2009) FOXP3+CD25- tumor cells with regulatory function in Sézary syndrome. J Invest Dermatol 129:2875–2885
- Capriotti E, Vonderheid EC, Thoburn CJ, Wasik MA, Bahler DW, Hess AD (2008) Expression of T-plastin, FoxP3 and other tumorassociated markers by leukemic T-cells of cutaneous T-cell lymphoma. Leuk Lymphoma 49:1190–1201
- 81. Krejsgaard T, Gjerdrum LM, Ralfkiaer E, Lauenborg B, Eriksen KW, Mathiesen AM, Bovin LF, Gniadecki R, Geisler C, Ryder LP, Zhang Q, Wasik MA, Odum N, Woetmann (2008) A Malignant Tregs express low molecular splice forms of FOXP3 in Sézary syndrome. Leukemia 22:2230–2239
- 82. Bouaziz JD, Remtoula N, Bensussan A, Marie-Cardine A, Bagot M (2010) Absolute CD3+ CD158k+lymphocyte count is reliable and more sensitive than cytomorphology to evaluate blood tumor burden in Sézary syndrome. Br J Dermatol 162:123–128
- Campbell JJ, Clark RA, Watanabe R, Kupper TS (2010) Sezary syndrome and mycosis fungoides arise from distinct T-cell subsets: a biologic rationale for their distinct clinical behaviors. Blood 116:767–771
- 84. Notohamiprodjo M, Segerer S, Huss R, Hildebrandt B, Soler D, Djafarzadeh R, Buck W, Nelson PJ, von Luettichau I (2005) CCR10 is expressed in cutaneous T-cell lymphoma. Int J Cancer 115:641–647
- Hwang ST, Janik JE, Jaffe ES, Wilson WH (2008) Mycosis fungoides and Sézary syndrome. Lancet 371:945–957
- Narducci MG, Scala E, Bresin A, Caprini E, Picchio MC, Remotti D, Ragone G, Nasorri F, Frontani M, Arcelli D, Volinia S, Lombardo GA, Baliva G, Napolitano M, Russo G (2006) Skin

homing of Sézary cells involves SDF-1-CXCR4 signaling and down-regulation of CD26/dipeptidylpeptidase IV. Blood 107:1108–1115

- Wu XS, Lonsdorf AS, Hwang ST (2009) Cutaneous T-cell lymphoma: roles for chemokines and chemokine receptors. J Invest Dermatol 129:1115–1119
- Capriotti E, Vonderheid EC, Thoburn CJ, Bright EC, Hess AD (2007) Chemokine receptor expression by leukemic T cells of cutaneous T-cell lymphoma: clinical and histopathological correlations. J Invest Dermatol 127:2882–2892
- 89. Sokolowska-Wojdylo M, Wenzel J, Gaffal E, Lenz J, Speuser P, Erdmann S, Abuzahra F, Bowman E, Roszkiewicz J, Bieber T, Tüting T (2005) Circulating clonal CLA(+) and CD4(+) T cells in Sezary syndrome express the skin-homing chemokine receptors CCR4 and CCR10 as well as the lymph node-homing chemokine receptor CCR7. Br J Dermatol 152:258–264
- Drillenburg P, Pals ST (2000) Cell adhesion receptors in lymphoma dissemination. Blood 95:1900–1910
- Borowitz MJ, Weidner A, Olsen EA, Picker LJ (1993) Abnormalities of circulating T-cell subpopulations in patients with cutaneous T-cell lymphoma: cutaneous lymphocyte-associated antigen expression on T cells correlates with extent of disease. Leukemia 7:859–863
- Heald PW, Yan SL, Edelson RL, Tigelaar R, Picker LJ (1993) Skin-selective lymphocyte homing mechanisms in the pathogenesis of leukemic cutaneous T-cell lymphoma. J Invest Dermatol 101:222–226
- 93. Chong BF, Murphy JE, Kupper TS, Fuhlbrigge RC (2004) Eselectin, thymus- and activation-regulated chemokine/CCL17, and intercellular adhesion molecule-1 are constitutively coexpressed in dermal microvessels: a foundation for a cutaneous immunosurveillance system. J Immunol 172:1575–1581
- 94. Hoeller C, Richardson SK, Ng LG, Valero T, Wysocka M, Rook AH, Weninger W (2009) In vivo imaging of cutaneous T-cell lymphoma migration to the skin. Cancer Res 69:2704–2708
- Wu CS, Wang ST, Liao CY, Wu MT (2008) Differential CCR4 expression and function in cutaneous T-cell lymphoma cell lines. Kaohsiung J Med Sci 24:577–590
- 96. Santamaria-Babí LF (2004) CLA(+) T cells in cutaneous diseases. Eur J Dermatol 14:13–18
- Saeki H, Tamaki K (2006) Thymus and activation regulated chemokine (TARC)/CCL17 and skin diseases. J Dermatol Sci 43:75–84
- 98. Yagi H, Seo N, Ohshima A, Itoh T, Itoh N, Horibe T, Yoshinari Y, Takigawa M, Hashizume H (2006) Chemokine receptor expression in cutaneous T cell and NK/T-cell lymphomas: immunohistochemical staining and in vitro chemotactic assay. Am J Surg Pathol 30:1111–1119
- 99. Kakinuma T, Sugaya M, Nakamura K, Kaneko F, Wakugawa M, Matsushima K, Tamaki K (2003) Thymus and activationregulated chemokine (TARC/CCL17) in mycosis fungoides: serum TARC levels reflect the disease activity of mycosis fungoides. J Am Acad Dermatol 48:23–30
- 100. Ferenczi K, Fuhlbrigge RC, Pinkus J, Pinkus GS, Kupper TS (2002) Increased CCR4 expression in cutaneous T cell lymphoma. J Invest Dermatol 119:1405–1410
- 101. Fierro MT, Comessatti A, Quaglino P, Ortoncelli M, Osella Abate S, Ponti R, Novelli M, Bernengo MG (2006) Expression pattern of chemokine receptors and chemokine release in inflammatory erythroderma and Sézary syndrome. Dermatology 213:284–292
- 102. Sugaya M (2010) Chemokines and cutaneous lymphoma. J Dermatol Sci 59:81–85
- 103. Kagami S, Sugaya M, Minatani Y, Ohmatsu H, Kakinuma T, Fujita H, Tamaki K (2006) Elevated serum CTACK/CCL27 levels in CTCL. J Invest Dermatol 126:1189–1191

- 104. Fujita Y, Abe R, Sasaki M, Honda A, Furuichi M, Asano Y, Norisugi O, Shimizu T, Shimizu H (2006) Presence of circulating CCR10+ T cells and elevated serum CTACK/CCL27 in the early stage of mycosis fungoides. Clin Cancer Res 12:2670–2675
- 105. Lu D, Duvic M, Medeiros LJ, Luthra R, Dorfman DM, Jones D (2001) The T-cell chemokine receptor CXCR3 is expressed highly in low-grade mycosis fungoides. Am J Clin Pathol 115:413–421
- 106. Kallinich T, Muche JM, Qin S, Sterry W, Audring H, Kroczek RA (2003) Chemokine receptor expression on neoplastic and

reactive T cells in the skin at different stages of mycosis fungoides. J Invest Dermatol 121:1045–1052

107. Yamaguchi T, Ohshima K, Tsuchiya T, Suehuji H, Karube K, Nakayama J, Suzumiya J, Yoshino T, Kikuchi M (2003) The comparison of expression of cutaneous lymphocyte-associated antigen (CLA), and Th1- and Th2- associated antigens in mycosis fungoides and cutaneous lesions of adult T-cell leukemia/lymphoma. Eur J Dermatol 13:553–559