

Chromosomal Integration of the HHV-6 Genome as a Possible Cause of Persistent HHV-6 Detection in a Patient with Langerhans Cell Histiocytosis

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Dear Editor

With interest we read the article published by Csire et al. The authors present the case of a 1975 born male suffering from relapsing Langerhans cell histiocytosis (LCH, eosinophilic granuloma). Treatment consisted of irradiation, chemotherapy and repeated surgery. The human herpesvirus 6 (HHV-6) genome has been detected by PCR in several materials (blood, bone marrow, histological samples of granulomas) obtained during the clinical course of 17 years. Despite antiviral therapy the HHV-6 genome remained detectable.

The authors suggested that a persistent HHV-6 infection might be associated with development or progression of LCH [1].

In contrast it might be speculated that the chromosomal integration of the HHV-6 (CIHHV-6) genome into the genome of the patient might be the reason for the persistent detection of viral DNA.

CIHHV-6 has been described in patients with a variety of different diseases like meningitis, encephalitis, convulsions, liver dysfunction, haemolytic and aplastic anaemia, different lymphomas, and multiple sclerosis [1–12]. Furthermore, it is found in healthy individuals [4, 8, 10, 13]. It is inherited to the offspring from one or both parent(s) [3, 4, 8, 14]. Hence, the HHV-6 genome is found in every nucleated cell. The prevalence is estimated to be 0.2 to 1.3% [8, 13, 15].

A special way of acquiring CIHHV-6 is the transplantation of stem cells with CIHHV-6 by means of bone marrow or stem cell transplantation (SCT) [16, 17].

CIHHV-6 leads to persistently high levels of HHV-6 DNA in blood and tissue, which might be interpreted as persistent active HHV-6 infection [5, 10, 12].

CIHHV-6 can be proven by fluorescent in situ hybridization (FISH) on chromosome preparations with a HHV-6 specific probe [4, 8, 10, 14]. Moreover, the suspicion of CIHHV-6 can be substantiated or ruled out by means of routine diagnostic procedures.

In individuals with CIHHV-6 the HHV-6 genome is present in any somatic cell. So detection of HHV-6 DNA in hair follicle or nails indicates CIHHV-6 [5, 10, 15].

Since there is at least one copy in every nucleic cell, a high amount of DNA copies is expected in individuals with CIHHV-6. However, the copy numbers in whole blood may be below the number of white blood cells (depending on laboratory techniques).

Since CIHHV-6 is inherited, it can be expected that (at least) one of the parents and possibly some of the siblings and the offspring have the HHV-6 genome chromosomally integrated [3, 4, 8]. If HHV-6 DNA can be detected in a first degree relative CIHHV-6 seems to be most likely. However, germ line integration with negative HHV-6 PCR in both parents might be considered.

Furthermore, the persistence of positive HHV-6 DNA detection itself indicates CIHHV-6 [8, 10, 12].

After acquisition of CIHHV-6 by SCT, HHV-6 PCR becomes positive in blood (or in tissue contaminated with blood cells) at the time of engraftment and it will stay positive thereafter [16, 17]. HHV-6 PCR results from relatives and tissue specimens (if not bloody contaminated) are expected to be negative. CIHHV-6 is present in materials from the donor (including back-up harvest).

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In some individuals with CIHHV-6 HHV-6 specific mRNA was detected by reverse transcription PCR indicating the replication of (at least parts of) the HHV-6 genome [14, 18]. Furthermore, we detected viral proteins by HHV-6 antigen detection in individuals with proven CIHHV-6 (unpublished data). In contrast, viral cultures have shown negative results [8]. Thus, it has to be clarified whether replication of viral particles occurs and if there is a pathophysiological impact of CIHHV-6.

It would be of great interest to perform the above mentioned diagnostic procedures in the reported patient to confirm or rule out CIHHV-6. If the HHV-6 genome is chromosomally integrated, it can be speculated that a permanent replication of viral proteins might have an impact on the course of disease. Otherwise, CIHHV-6 might be a coincidental finding without any association with the reported disease.

References

1. Csire M, Mikala G, Jako J et al (2007) Persistent long-term human herpesvirus 6 (HHV-6) infection in a patient with langerhans cell histiocytosis. *Pathol Oncol Res* 13:157–160
2. Boutolleau D, Agut H, Gautheret-Dejean A (2006) Human herpesvirus 6 genome integration: a possible cause of misdiagnosis of active viral infection? *J Infect Dis* 194:1019–1020 author reply 1021–1013
3. Caserta MT, Hall CB, Schnabel K, Lofthus G, McDermott MP (2007) Human herpesvirus (HHV)-6 and HHV-7 infections in pregnant women. *J Infect Dis* 196:1296–1303
4. Daibata M, Taguchi T, Nemoto Y, Taguchi H, Miyoshi I (1999) Inheritance of chromosomally integrated human herpesvirus 6 DNA. *Blood* 94:1545–1549
5. Hubacek P, Maalouf J, Zajickova M et al (2007) Failure of multiple antivirals to affect high HHV-6 DNAemia resulting from viral chromosomal integration in case of severe aplastic anaemia. *Haematologica* 92:e98–e100
6. Luppi M, Barozzi P, Morris CM, Merelli E, Torelli G (1998) Integration of human herpesvirus 6 genome in human chromosomes. *Lancet* 352:1707–1708
7. Luppi M, Marasca R, Barozzi P et al (1993) Three cases of human herpesvirus-6 latent infection: integration of viral genome in peripheral blood mononuclear cell DNA. *J Med Virol* 40:44–52
8. Tanaka-Taya K, Sashihara J, Kurahashi H et al (2004) Human herpesvirus 6 (HHV-6) is transmitted from parent to child in an integrated form and characterization of cases with chromosomally integrated HHV-6 DNA. *J Med Virol* 73:465–473
9. Torelli G, Barozzi P, Marasca R et al (1995) Targeted integration of human herpesvirus 6 in the p arm of chromosome 17 of human peripheral blood mononuclear cells in vivo. *J Med Virol* 46:178–188
10. Ward KN, Leong HN, Nacheva EP et al (2006) Human herpesvirus 6 chromosomal integration in immunocompetent patients results in high levels of viral DNA in blood, sera, and hair follicles. *J Clin Microbiol* 44:1571–1574
11. Ward KN, Leong HN, Thiruchelvam AD, Atkinson CE, Clark DA (2007) Human herpesvirus 6 DNA levels in cerebrospinal fluid due to primary infection differ from those due to chromosomal viral integration and have implications for diagnosis of encephalitis. *J Clin Microbiol* 45:1298–1304
12. Ward KN, Thiruchelvam AD, Couto-Parada X (2005) Unexpected occasional persistence of high levels of HHV-6 DNA in sera: detection of variants A and B. *J Med Virol* 76:563–570
13. Leong HN, Tuke PW, Tedder RS et al (2007) The prevalence of chromosomally integrated human herpesvirus 6 genomes in the blood of UK blood donors. *J Med Virol* 79:45–51
14. Hall CB, Caserta MT, Schnabel K et al (2008) Chromosomal integration of human herpesvirus 6 is the major mode of congenital human herpesvirus 6 infection. *Pediatrics* 122:513–520
15. Hubacek P, Muzikova K, Hrdlickova A et al (2009) Prevalence of HHV-6 integrated chromosomally among children treated for acute lymphoblastic or myeloid leukemia in the Czech Republic. *J Med Virol* 81:258–263
16. Clark DA, Nacheva EP, Leong HN et al (2006) Transmission of integrated human herpesvirus 6 through stem cell transplantation: implications for laboratory diagnosis. *J Infect Dis* 193:912–916
17. Kamble RT, Clark DA, Leong HN, Heslop HE, Brenner MK, Carrum G (2007) Transmission of integrated human herpesvirus-6 in allogeneic hematopoietic stem cell transplantation. *Bone Marrow Transplant* 40:563–566
18. Clark DA, Tsao EH, Leong HN, Ward KN, Nacheva EP, Griffiths PD (2006) Reply to Boutolleau et al. and Luppi et al. *J Infect Dis* 194:1019–1020 author reply 1021–1013