

Expression of p21^{waf1/cip1}, p27^{kip1}, p63 and Androgen Receptor in Low and High Gleason Score Prostate Cancer

Imre Romics · Gergely Bánfi · Eszter Székely ·
Tibor Krenács · Béla Szende

Received: 10 March 2008 / Accepted: 25 March 2008 / Published online: 16 April 2008
© Arányi Lajos Foundation 2008

Abstract The aim of this study was to investigate the expression of p21^{waf1/cip1}, p27^{kip1}, p63 and androgen receptor proteins in relation to serum prostate specific antigen levels in low and high Gleason score prostate cancers. Biopsies of patients suffering from prostate adenocarcinoma of low (3+3 to 3+4) and high (5+4 to 5+5) Gleason scores (13 cases each group) were immunostained for positive regulators of cell cycle control (p21^{waf1/cip1} and p27^{kip1}), and essential markers of normal prostate gland ontogeny (p63) and growth (androgen receptor) to find differentially expressed markers of malignant progression. Serum prostate specific antigen levels were also monitored at the time of biopsy and following anti-androgen therapy. All cases except one in each group were androgen receptor positive. P63 and p21^{waf1/cip1} proteins detected in normal basal cell nuclei were lost in all but one studied tumors respectively. P27^{kip1} protein, however, was detected in all low Gleason score prostate cancers, but it was found in only 7/13 high score cases. Prostate specific antigen levels, either pre- or post-treatment, did not show strict correlation with the

p27^{kip1} results. The low to high grade dedifferentiation of prostate adenocarcinoma is accompanied with the down-regulation of p27^{kip1} protein, which may be an important molecular sign of the lost cell cycle control.

Keywords High Gleason score · Loss of p27^{kip1} · Prostate adenocarcinoma

Abbreviations

AR androgen receptors
H&E Haematoxylin–eosin
PSA prostate specific antigen
CAB complete androgen blockade

Introduction

Antiandrogen therapy of prostate carcinoma is based on the fact that most of these tumors are androgen dependent [1]. Androgens exert their growth promoting effect through androgen receptors (AR) [2]. Immunohistochemical demonstration of AR-s is successful in the majority of prostate carcinomas. However, AR positivity may be seen in antiandrogen-therapy resistant cases, too, which observation led to better understanding of mutations in the structure of AR-s. Androgens are also in close interaction with several tumor-suppressor and cell proliferation promoter genes [3–5]. These interactions may be manifested in up-regulation or suppression of such genes, upon the effect of androgens or deprivation of androgens by any type of antiandrogen therapy [6].

The aim of our study was to investigate the expression of growth regulating factors p21^{waf1/cip1}, p27^{kip1}, p63 and AR in tumor cells of prostate carcinomas, showing high (5+4 to 5+5) and low (3+3 to 3+4) Gleason score.

I. Romics · G. Bánfi
Department of Urology, Semmelweis University,
Budapest, Hungary

T. Krenács · B. Szende (✉)
1st Department of Pathology and Experimental Cancer Research,
Semmelweis University and Research Group of Molecular Pathology,
Hungarian Academy of Sciences and Semmelweis University,
Üllői u. 26,
1085 Budapest, Hungary
e-mail: bszende@korbl.sote.hu

E. Székely
2nd Department of Pathology, Semmelweis University,
Budapest, Hungary

Materials and Methods

Prostate biopsies of 13 patients suffering from 5+5 or 5+4 (Group 1) as well as 13 patients suffering from 3+3–3+4 (Group 2) Gleason score prostate carcinoma were investigated histologically. The primary indication of the biopsy was to confirm the diagnosis suggested by rectal digital examination and/or elevated prostate specific antigen (PSA)-level. The samples were used for immunohistochemical studies after informed consent of the patients and permission of the Local Ethical Committee.

Patients' age, PSA values at diagnosis and on average 29.96 (3–42) months after diagnosis, therapy, tumor–node–metastasis (TNM) stage and Gleason score are indicated in Tables 1 and 2. In addition to the routine H&E staining androgen receptor, p21, p27 and p63 expression was investigated using immunoperoxidase reactions.

The needle biopsy samples were fixed in 4% neutral-buffered formalin, embedded into paraffin. Five μ m thick sections were cut and stained with H&E, and proceeded for immunoperoxidase staining. Monoclonal mouse primary antibodies against human-p21, -androgen receptor (Dako, Glostrup, Denmark), -p27 (Novocastra, Newcastle upon Tyne, UK) and p63 (BioGenex, San Reno, CA, USA) antigens were used in 1:100 dilutions at 37°C overnight. Heat induced antigen/epitope retrieval before immunostaining included microwaving of dewaxed sections in TRS buffer (pH 6, Dako) at 400 W power for 30 min. Endogenous peroxidases were inactivated in methanol-H₂O₂. Ready-to-use EnVision Plus horseradish peroxidase

conjugate was used for 30 min incubation to detect primary antibodies and the reaction product was developed using diaminobenzidine-H₂O₂ chromogen-substrate system.

AR, p21, p27 and p63 indices were established by analyzing 1000 tumor cells. A tumor was considered positive for AR and all other gene products if more than 50% of the tumor cells showed obvious staining even if it was of moderate intensity.

Results

All tumors were adenocarcinomas, one of them in group1 was mucin producing. The age of the patients and the clinical data are shown in Tables 1 and 2. Thirteen prostate carcinomas were of 5+5 or 5+4 (Group 1) and 13 of 3+3–3+4 (Group 2) Gleason score.

Group 1

Elevated or highly elevated PSA values were found at the time of diagnosis, which markedly decreased upon therapy, except in one case. Five patients died on average 14.6 (3–28) months after diagnosis six patients are still alive on average 37 (31–42) months after diagnosis and no data about two patients are available. The TNM staging shown in the tables is that observed at the time of diagnosis. Six patients received complete androgen blockade (CAB). Radical prostatectomy was carried out in three cases, amended with CAB and bicalutamide therapy in one case, with CAB in another case, and with bicalutamide in the

Table 1 Group I. age, PSA value, Gleason score, TNM stage, therapy, clinical status, AR, p21^{waf1/cip1}, p63, and p27^{kip1} expression of patients with high Gleason score

No	Age (years)	PSA cc at the time of biopsy (ng/ml)	Last PSA level (ng/ml)	TNM	Therapy	AR	p21	p63	p27	Gleason-score	Status (months after biopsy)
1	77	15.6	7.6	T2cN0M0	CAB	+	–	–	+	5+5	Alive, 42
2	78	500	92.4	T3aNxM1b	CAB	+	+	–	+	4+5	Bone metastasis, died, 13
3	62	2116	504	T4NxM1b	Estramustine	+	–	–	–	4+5	Bone metastasis, died, 28
4	81	30.5	N. D.	T3aNxM0	Castration	+	–	–	+	5+5	N. D.
5	75	250	15.4	T3aN1M1b	Castration, estramustine, bisphosphonate	+	–	–	+	5+5	Died, 10
6	64	50	0.1	T2bNxM0	rad. prost., bicalutamide, CAB	+	–	–	–	4+5	Alive, 37
7	61	17	0.0	T3aN0M0	rad. prost., CAB	+	–	–	–	5+5	Alive, 39
8	74	118	76	pT2cNxM0	Castration	+	–	–	+	5+5	Alive, 37
9	64	27.1	0.3	T3bN0M0	rad. prost., bicalutamide,	+	–	–	–	4+5	Alive, 36
10	65	89	60	T2cNxM1b	Estramustine	+	–	–	+	4+5	Died, 19
11	72	133	8.9	T2cNxM0	CAB	+	–	–	–	5+5	Alive, 31
12	74	1000	N. D.	T2cNxM0	Castration	+	–	–	–	4+5	N. D.
13	77	38	38	T2cNxM0	CAB	–	–	–	+	5+5	Died, 3

Table 2 Group 2. Age, PSA value, Gleason score, TNM stage, therapy, clinical status, AR, p21^{waf1/cip1}, p63, and p27^{kip1} expression of patients with low Gleason score

No	Age (years)	PSA cc at the time of biopsy (ng/ml)	Last PSA level (ng/ml)	TNM	Therapy	AR	p21	p63	p27	Gleason-score	Status (months after biopsy)
1	78	39.4	0.0	T2bNxM0	Monoantiandrogen, castration	+	-	-	+	3+3	Alive, 37
2	65	12.7	0.0	T1cN0M0	rad. prost.	+	-	-	+	3+3	Alive, 35
3	62	16.1	0.0	T2cN0M0	rad. prost.	-	-	-	+	3+3	Alive, 41
4	71	131.4	3.7	T3aNxM1b	CAB	+	-	-	+	3+4	Bone metastasis, alive 32
5	80	173.9	0.0	T3aNxM0	CAB	+	-	-	+	3+3	Alive, 38
6	60	25.5	1.6	T2aN0M0	rad. prost.	+	-	-	+	3+3	Alive, 30 Local recurrence
7	68	102.9	11.2	T2bNxM0	CAB	+	-	-	+	3+4	Alive, 36
8	77	16.2	7.5	T2aNxM0	CAB	+	-	-	+	3+3	Alive, 40
9	68	50	2.7	T3bNxM1b	Castration	+	-	-	+	3+4	Died, 20
10	71	5.3	0.0	pT2bN0M0	rad. prost.	+	-	-	+	3+3	Alive, 31
11	83	14.5	0.1	T3aNxM0	CAB	+	-	-	+	3+3	Died, 18
12	76	148.1	0.3	T3bNxM0	Monoantiandrogen	+	-	-	+	3+3	Alive, 31
13	56	4.6	0.8	pT2cNxM0	rad. prost.	+	-	-	+	3+3	Alive, 35

CAB complete androgen blockade, *rad. prost.* radical prostatectomy

third case. In three cases castration, in another two cases estramustine treatment was the only therapy applied. One patient was treated with castration, estramustine and bisphosphonate.

The prostate carcinoma cells of all but one patient were androgen receptor positive. The patient with AR negative tumor died 3 months after the biopsy.

All tumors were p63 negative. P63 positivity was found in the cells of normal or hyperplastic glands, occasionally as a part of biopsy samples containing tumor tissue as well. p21 positivity was only found in the mucin-producing adenocarcinoma in our material.

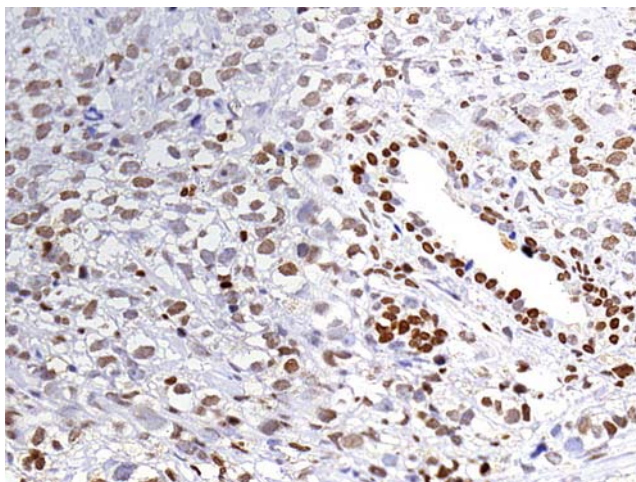


Fig. 1 Weak nuclear immunostaining for p27 antigen in Gleason score 5+5 prostate adenocarcinoma cells. Note the strong staining in the cell nuclei of a pre-existing duct on the *right* (p27 immunoperoxidase, $\times 600$)

p27 positivity was detected in seven of 13 cases (Fig. 1), four of whom died during the follow up period, two are still alive and we have no data from one patient. One of the patients in this group was AR negative (Table 1).

Group 2

Moderately or highly elevated PSA levels were found at the time of diagnosis, which were significantly decreased upon therapy. Two patient died 18 and 20 months after diagnosis. Eleven patients are still alive 35 (30–41) months after diagnosis. The TNM staging showed T1c–3bNxM0–1b at

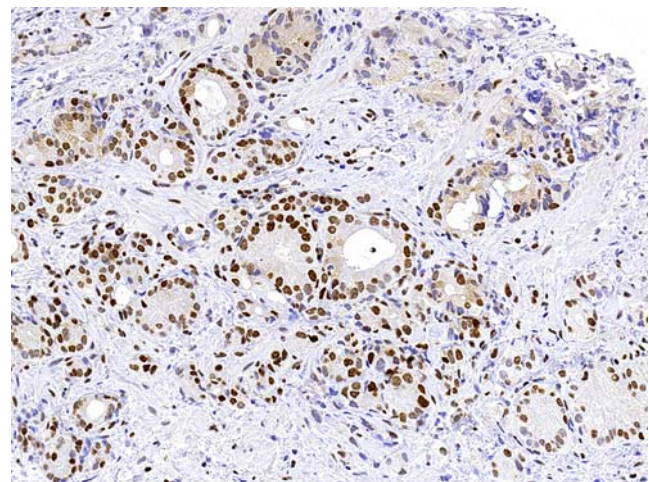


Fig. 2 Moderate to strong nuclear immunostaining for p27 antigen in Gleason score 3+4 prostate adenocarcinoma cells (p27 immunoperoxidase, $\times 300$)

the time of diagnosis. Five patients received complete androgen blockade. Radical prostatectomy was applied to five patients, monoantiandrogen to one patient alone and monoantiandrogen+castration in one and one patient was castrated.

Androgen receptor positivity was found in 12 cases and only one prostate carcinoma was AR negative. Like in group 1 all tumors were negative for p21 and p63, but all studied cases in this group were positive for p27 protein (Table 2; Fig. 2)

Discussion

Compared to normal prostate epithelium several genes are expressed differentially in prostate carcinoma. Genes playing a role in signal transduction (c-myc, HER-2/neu) and apoptosis (fas, bcl2) are usually up-regulated, whereas tumor-suppressor genes like p53 and p27 show down-regulation. The low expression of tumor-suppressor genes, together with decreased apoptotic and increased proliferative activity, may be considered as predictor of unfavorable outcome of the disease [7, 8]. Experimental studies showed that castration-induced atrophy was associated with an increase in p27 expression and androgen treatment decreased p27 levels [9]. Also, p27 can mediate growth controlling effects in tumor cell lines e.g. that exerted by forced connexin43 expression [10, 11]. Gorbe et al. (2005) detected evenly high levels of both p27 and p21 proteins in aligned myoblasts, which left cell cycle just preceding fusion [12].

P63 is essential for prostate development and is selectively expressed in adult prostate basal cells. However, p63 expression is lost in prostate carcinoma [13].

The action of the cell senescence inhibitor p21 is based on the inhibition of cyclin dependent kinases which mediate the expression of tumor-promoting factors [14, 15]. Upregulation of p21 has been shown to play an important role in promoting myoblast differentiation, proliferation and viability [16]. Accelerated senescence leads to decreased ability of cell proliferation. Increased p21 activity seems to counteract this process. Antiandrogen treatment appears to suppress the activity of p21 [17].

All tumors in our series were p63 negative, in accordance with previous studies [18].

Similarly, no p21 nuclear positivity could be detected in the tumor cells in either studied groups, except one, mucin-producing adenocarcinoma.

Androgen receptor positivity was found in nearly all prostate carcinomas, except of two cases. No difference was observed for androgen receptor expression between the two groups we studied.

However, difference between the high Gleason score (Group 1) and low Gleason score (Group 2) tumors occurred regarding nuclear positivity for p27. Only seven of 13 tumors showing high Gleason score showed positive reaction for p27^{kip1}, whereas all low-grade tumors were p27^{kip1} positive. Cordon-Cardo et al. [19] observed distinct altered patterns of p27^{kip1} gene expression in benign prostatic hyperplasia and prostatic carcinoma already in 1998. Correlation between loss of p27 protein in prostate cancer and tumor grade was reported by Guo et al. [20]. Kuczyk et al. [21] attributed predictive value to the decrease of p27 expression for the recurrence-free and long-term survival of prostate cancer patients. Recently, Doganavsargil et al. [22] have shown that higher p21 and lower p27 expression is correlated with adverse prognostic factors in case of prostate carcinoma.

In this study, the low to high grade progression of prostate adenocarcinoma was accompanied with the down-regulation of p27^{kip1} protein, which may be an important molecular sign of the lost cell cycle control. Regarding differential diagnosis, neither positivity nor negativity for nuclear p27^{kip1} protein exclude or prove the existence of prostate carcinoma.

References

1. Isaacs JT, Lundmo PI, Berges R, Martikainen P, Kyprianou N, English HF (1992) Androgen regulation of programmed death of normal and malignant prostatic cells. *J Androl* 13:457–464
2. Eder IE, Culig Z, Putz T, Nessler-Menardi C, Bartsch G, Klocker H (2001) Molecular biology of the androgen receptor: from molecular understanding to the clinic. *Eur Urol* 40:241–251
3. Stamey TA, Warrington JA, Caldwell MC et al (2001) Molecular genetic profiling of Gleason grade 4/5 prostate cancers compared to benign prostatic hyperplasia. *J Urol* 166:2171–2177
4. Porkka KP, Visakorpi T (2001) Detection of differentially expressed genes in prostate cancer by combining suppression subtractive hybridization and cDNA library assay. *J Pathol* 193:73–79
5. Meehan KL, Holland JW, Dawkins HJ (2002) Proteomic analysis of normal and malignant prostate tissue to identify novel proteins lost in cancer. *Prostate* 50:54–63
6. Baretton GB, Klenk U, Diebold J, Schmeller N, Lohrs U (1999) Proliferation- and apoptosis-associated factors in advanced prostatic carcinomas before and after androgen deprivation therapy: prognostic significance of p21/WAF1/CIP1 expression. *Br J Cancer* 80:546–555
7. Eder IE, Bektic J, Haag P, Bartsch G, Klocker H (2004) Genes differentially expressed in prostate cancer. *BJU Int* 93:1151–1155
8. Szende B, Romics I, Minik K, Szabó J, Torda I, Lovász S, Tóth L, Bély M, Kerényi T, Bartók K, Végh A (2001) Repeated biopsies in evaluation of therapeutic effects in prostate carcinoma. *Prostate* 49:93–100
9. Waltregny D, Leav I, Signoretti S, Soung P, Lin D, Merk F, Adams JY, Bhattecharya N, Cirenei N, Loda M (2001) Androgen-driven prostate epithelial cell proliferation and differentiation in vivo involve the regulation of p27. *Mol Endocrinol* 15:765–782

10. Koffer L, Roshong S, Park IK, Cesen-Cummings K, Thopson DR, Dwyer-Nield LD, Rice P, Mamay C, Malkinson AM, Ruch RJ (2000) Growth inhibition in G1 and altered expression of cyclin D1 and p27kip1 after forced connexin expression in lung and liver carcinoma cells. *J Cell Biochem* 79:347–354
11. Zhang Y-W, Morite I, Ikeda M, Ma K-W, Murota S (2001) Connexin43 suppresses proliferation of osteosarcoma U2OS cells through post-transcriptional regulation of p27. *Oncogene* 20:4138–4149
12. Gorbe A, Becker DL, Dux L, Stelkovic E, Krenacs L, Bagdi E, Krenacs T (2005) Transient upregulation of connexin43 gap junctions and synchronized cell cycle control precede myoblast fusion in regenerating skeletal muscle in vivo. *Histochem Cell Biol* 123:573–583
13. Lawlor MA, Rotwein P (2000) Coordinate control of muscle cell survival by distinct insulin-like growth factor activated signaling pathways. *J Cell Biol* 151:1131–1140
14. Ostrovsky O, Bengal E (2003) The mitogen-activated protein kinase cascade promotes myoblast cell survival by stabilizing the cyclin-dependent kinase inhibitor, p21WAF1 protein. *J Biol Chem* 278:21221–21231
15. Fizazi K, Martizez LA, Sikes CR, Johnston DA, Stephens LC, McDonnell TJ, Logothetis CJ, Trapman J, Pisters LL, Ordonez NG, Troncoso P, Navone NM (2002) The association of p21^(WAF-1/CIP1) with progression to androgen-independent prostate cancer. *Clin Cancer Res* 8:775–781
16. Signoretti S, Waltregny D, Dilks J, Isaac B, Lin D, Garraway L, Yang A, Montironi R, McKeon F, Loda M (2000) P63 is a prostate basal cell marker and is required for prostate development. *Am J Pathol* 157:1769–1775
17. Ronison IB (2003) Tumor cell senescence in cancer treatment. *Cancer Res* 63:2705–2715
18. Di Como CJ, Urist MJ, Babayan I, Drobnjak M, Hedvat CV, Teruya-Feldstein J, Pohar K, Hoos A, Cordon-Cardo C (2002) P63 expression profiles in human normal and tumor tissues. *Clin Can Res* 8:494–501
19. Cordon-Cardo G, Koff A, Drobnjak M, Caodiec P, Osman I, Millard SS (1998) Distinct altered patterns of p27^{kip1} gene expression in benign prostate hyperplasia and prostatic carcinoma. *J Natl Cancer Inst* 90:1284–1291
20. Guo Y, Sklar GN, Borkowski A, Kyprianou N (1997) Loss of the cyclin dependent kinase inhibitor p27(Kip1) protein in human prostate cancers correlates with tumor grade. *Clin Cancer Res* 3:2269–2274
21. Kuczyk M, Machtens S, Hradil K et al (1999) Predictive value of p27Kip1 protein expression for the recurrence-free and long-term survival of prostate cancer patients. *Br J Cancer* 81:1052–1058
22. Doganavsargil B, Simsir A, Boyacioglu H, Cal C, Hekimgil M (2006) A comparison of p21 and p27 immunorexpression in benign glands, prostatic intraepithelial neoplasia and prostate adenocarcinoma. *BJU Int* 97:644–648