REVIEW

The HNK-1 Carbohydrate Epitope and the Human Eye in Health and Disease

Marita UUSITALO and Tero KIVELÄ

Ophthalmic Pathology Laboratory, Department of Ophthalmology, Helsinki University Central Hospital, Helsinki, Finland

The HNK-1 carbohydrate epitope is part of many cell membrane and extracellular matrix molecules, several of which have been implicated in cell adhesion. It is a versatile tool in eye research. In the human eye this epitope is present in the retina, the optic and ciliary nerves, the ciliary and iris epithelia, the zonular lamella, and the sclera. It is phylogenetically conserved, but the positive cell types vary from species to species. In addition to revealing interspecies differences in the vertebrate retina, the HNK-1 epitope has been used to identify a novel cell type in the eye: the subepithelial matrix cells that reside in the inner connective tissue layer (ICTL) of the ciliary body. Although these cells resemble fibroblasts in ultrastructure, they form a distinct cell population that differs in antigenic profile from fibroblasts in other tissues. The HNK-

1 epitope is also associated with the elastic fiber system of the ICTL, which may be produced by the subepithelial matrix cells. It may help to structurally stabilize the ciliary body and the retina. The HNK-1 epitope is also involved in many important eye diseases. The subepithelial matrix cells seem to be susceptible to irrreversible atrophy as a result of glaucoma, thermal injury, and tissue compression. On the other hand, the HNK-1 epitope is found in the extracellular matrix of secondary cataracts and may contribute to its pathogenesis. Finally, this epitope has proved to be useful in identifying deposits of exfoliation material, and in tracing neuroepithelial derivatives in developmental anomalies and tumors of the eye. (Pathology Oncology Research Vol 3, No 1, 8–14, 1997)

Key words: Cell adhesion, Ciliary body, Inner connective tissue layer, Natural killer cell, Retina, Subepithelial matrix cell

HNK-1 carbohydrate epitope

In 1981 Abo and Balch¹ raised monoclonal antibodies against a membrane fraction of the HSB-2 human T-lymphoblastoid cell line and selected one, designated HNK-1 (Human Natural Killer), because it specifically labelled a subset of lymphocytes enriched in natural killer and killer cells. Although initially aimed for use as a tool in leukocyte research, the same antibody was soon found to label many neuronal, glial, neuroectodermal and neuroendocrine cells.^{2,7,30,42,52,53} The corresponding epitope common to lym-

phocytes and the nervous system is generally known as the HNK-1 epitope. It proved to be highly immunogenic,¹⁶ and a number of other antibodies raised for different purposes recognize an antigen identical or closely related to the HNK-1 epitope.^{5,28,58} In leukocyte research, the HNK-1 epitope was renamed first Leu-7 and then CD57.

The HNK-1 epitope is a 3-sulphoglucuronic acid-containing carbohydrate moiety, but its exact structure is still unknown.^{4,8,34} It is part of several extracellular matrix and cell membrane glycoproteins, glycolipids and proteoglycans, such as neural cell adhesion molecule, myelin-associated glycoprotein, 11-integrin, tenascin and chondroitin sulphate proteoglycan.^{27,28,34,37,43}

Functionally, the HNK-1 epitope has been linked to cell adhesion, and it has been proved vital in migration of fetal neural crest cells.⁶ It has been shown to be the adhesive domain in at least the neuron-extracellular matrix interaction, and it is also involved in neuron-astrocyte and astro-

Received: Dec 30, 1996; accepted: Febr, 27, 1997

Correspondence: Marita UUSITALO, M.D., Ophthalmic Pathology Laboratory, Department of Ophthalmology, Helsinki University Central Hospital, Haartmaninkatu 4C; FIN-00290 Helsinki, Finland; Tel: +358-9-471 3160, Fax: +358-9-471 5569; E-mail msuusita@cc.helsinki.fi

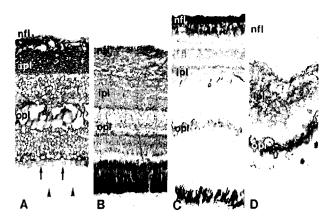


Figure 1. Phylogenetic differences in the immunostaining for the HNK-1 epitope in the retina (immunoperoxidase staining). **A.** In the human retina all retinal layers except the photoreceptor layer (arrowheads) are labelled. Note that labelling is present in the basket fibers of Müller's cells between the photoreceptors (arrows). **B.** Equal immunoreactivity is seen in the retina of the bass, a fish. **C.** In the duck, both plexiform layers, some cells adjacent to them, and the nerve fiber layer are labelled. In B and C, note pigmented epithelial cells below the retina. **D.** In the rabbit, immunoreaction is limited to both plexiform layers. Original magnifications: A x340, B x160, C x340, and D x330; nfl - nerve fiber layer, ipl - inner plexiform layer, opl - outer plexiform layer.

cyte-astrocyte adhesion.^{18,29,46} Circulating autoantibodies to the HNK-1 epitope can cross-react with myelin proteins and cause a peripheral neuropathy.^{17,41}

HNK-1 epitope and the healthy eye

1. Posterior segment

In the eye, the first tissues found to contain the HNK-1 epitope were the optic nerve and the retina.^{19,42,58,68} The oligodendrocytes and myelin sheaths of the optic nerve express the HNK-1 epitope.^{19,24} It is additionally present in the ground substance between the collagen lamellae of the sclera, but it has not been detected in the choroid.^{59,64}

The HNK-1 epitope appears in the neuroblastic layers and in the pigment epithelium of the developing retina before the 13th gestational week.^{24,64} In the adult human eye all retinal layers except the photoreceptor cell layer express the epitope (*Fig.1A*). It is probably associated with neuronal cell membranes in both plexiform layers and in the nerve fiber layer. It is present on astrocytes of the inner retina and prominent in the radial fibers of Müller. The internal limiting membrane and the basket fibers of Müller's cells between the inner segments of photoreceptors are likewise immunoreactive. Due to its adhesive properties the HNK-1 epitope may well help to stabilize the retinal architecture.

The HNK-1 cpitope in the retina is phylogenetically conserved as it is found in various fishes, birds, mammals,

and primates (Fig.1B-D).^{5,11,38,48,61,68} In fishes and primates this epitope is present throughout the retina, analogous to humans (*Fig.1A,B*). However, in other studied mammals only the plexiform layers contain the epitope, and in birds it is restricted to the plexiform and the nerve fiber layers (*Fig.1 C,D*).

2. The inner connective tissue layer of the ciliary body

An observation of major importance has been the detection of a novel cell type in the human ciliary body that is identified by its immunoreactivity for the HNK-1 epitope.

The stromal layer filling the space between the ciliary epithelia and the ciliary muscle in the ciliary body is known as the inner connective tissue layer (ICTL; *Fig.2*).¹⁵ This layer has been regarded as a nondescript tissue with no special functional role. It traditionally consists of fibroblasts, melanocytes, lymphocytes, mast cells and macrophages together with blood vessels, nerves, collagen bundles and elastin fibers.^{15,55} It merges imperceptibly with the stroma of the iris and choroid when studied by routine light microscopy. In contrast, immunoreaction for the HNK-1 epitope distinctly delineates the human ICTL from the base of the iris, from the ora serrata, and from the ciliary muscle (*Fig.3A*).^{59,64}

The HNK-1 epitope in the ICTL first appears adjacent to the pigmented epithelium of the ciliary processes of the pars plicata by the 20th week of gestation (*Fig.4*).^{24,64} This has raised the possibility that the latter induces and organises its development. The epitope gruadally spreads to the entire ICTL of the pars plicata and then to the pars plana by the first year of life (*Fig.4*). Thereafter the HNK-1 epitope remains apparently unaltered in the ICTL throughout life.

By light microscopy, immunoreaction for the HNK-1 epitope outlines in the ICTL a population of stellate cells with long processes related to coarse immunoreactive bundles and to a meshwork of fine fibers (Fig.3A-C).^{60,64}

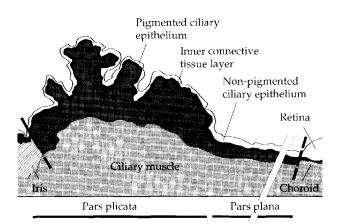


Figure 2. Diagrammatic cross-section through the ciliary body, defining the location of the inner connective tissue layer.

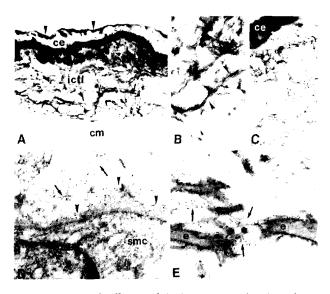


Figure 3. A novel cell type of the inner connective tissue layer (ICTL) of the human ciliary body identified by immunoreaction for the HNK-1 epitope (A-C light microscopic immunoperoxidase staining, D and E immunoelectron microscopy). A. Immunostaining distinctly delineatied the ICTL against the ciliary muscle (cm). Immunoreaction is also present at the basement membrane (arrowheads) of the non-pigmented ciliary epithelium (ce). The pigmented ciliary epithelium is dark due to its melanin content. B. With higher magnification, immunolabelling reveals in the ICTL long strands (arrowheads), and C. a meshwork of fine fibrils. D. Immunoelectron microscopy reveals that the epitope is situated on the surface of the subepithelial matrix cells (smc) at the level of their fragmented basement membrane (arrrowheads). Immunoreaction also highlights labelled microfibrils (arrows) that extend from the cell surface. No labelling is seen in the cytoplasm or the nucleus (n). E. Immunoreactive fibers (arrows) seem to connect elastic fibers to each other (e). Immunostaining is also present on the mantle of elastic fibers surrounding their amorphous core. Original magnifications: A x310, B x800, C x1000, D x37000, and E x36000.

Immunoelectron microscopy has revealed that the epitope resides on the surface of these cells (*Fig.3D*) that ultrastructurally resemble fibroblasts. It must be emphasized that these HNK-1 positive stromal cells form a distinct cell population that differs from fibroblasts of the iris, ciliary muscle and choroid. In particular, they do not react for vimentin, the intermediate filament type usually characteristic of fibroblasts.⁶⁴ We thus believe that it is justified to set this well defined population of cells aside and designate it subepithelial matrix cells based on their location in the ICTL.

In addition, the HNK-1 epitope is present along long microfibrillar bundles that consist of ca. 10 nm thick oxytalan microfibrils (*Fig.3E*).^{67.} These bundles come into close contact both with the subepithelial matrix cells and with elastic fibers (*Fig.3E*), which they seemingly connect to each other. Indeed, the HNK-1 epitope is also present in a mantle of microfibrils around the elastin core at the periphery of elastic fibers (Fig.3E). The fact that the epitope is a common denominator to the subepithelial matrix cells and to the elastic fiber system of the ciliary body suggests that it may have a role in structurally stabilizing the ICTL, for example in relation to accommodation or secretion of aqueous humor.

To date we lack a method to label these subepithelial matrix cells in species other than man, including primate eyes.⁶¹ They lack the HNK-1 epitope in spite of the fact that it is present in other structures of the eye. The corresponding carbohydrate epitope or the molecule bearing it in the ICTL, if present, may be different in these species, or the HNK-1 epitope is somehow hidden so that antibodies cannot detect it.

3. Other anterior segment structures

The HNK-1 epitope appears in the neuroectodermally derived epithelia of the iris and ciliary body by the 22nd gestational week. In the anterior segment of the adult eye, it is found at the level of the basement membrane of the nonpigmented and the pigmented ciliary epithelium, as well as in the cytoplasm of the pigmented ciliary epithelium and of the posterior pigmented epithelium of the iris.^{59,60} Schwann cells of ciliary nerves are also labelled.¹⁹

No clear immunoreaction is seen in the lens epithelial cells or the lens capsule by light microscopy, but the lamella to which the zonules attach is immunoreactive for the HNK-1 epitope.⁶⁵ Nevertheless, lesser amounts of the epitope have been detected in the outer two thirds of the lens capsule by immunoelectron microscopy.⁴⁴

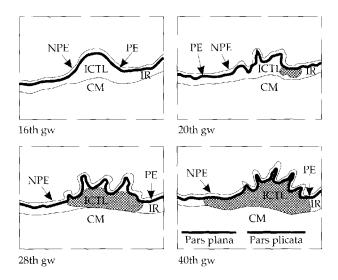


Figure 4. Appearance and development of the HNK-1 immunoreactivity (hatched area) in the human fetal ciliary body. ICTL – inner connective tissue layer, CM – ciliary muscle, NPE – non-pigmented and PE – pigmented ciliary epithelium, IR – iris, and gw – gestational week.

PATHOLOGY ONCOLOGY RESEARCH

HNK-1 epitope and the diseased eye

Analysis of changes in the HNK-1 epitope under pathological conditions is necessary to find out both its physiologic role and its significance in ocular disease.

1. Glaucoma

Glaucoma is an important cause of blindness throughout the world. In eyes with glaucoma, the immunoreaction for the HNK-1 epitope diminishes and eventually disappears from the ICTL (*Fig.5A*).⁶⁶ In contrast, the expression of the epitope is unchanged in all other structures of the eye. This might be due to the elevated intraocular pressure, which for example by decreasing blood flow may cause atrophy and degeneration of the subepithelial matrix cells that probably are responsible for the production of the HNK-1 epitope in the ICTL. Alternatively, an as yet unidentified biochemical change or long-term effect of glaucoma medication may be responsible.

Also after cyclodestructive surgery, such as contact krypton laser cyclophotocoagulation, the HNK-1 immunoreaction is all but lost from the treated area of the ICTL but not from the ciliary epithelium (*Fig.5B*).²³ This also suggests that the subepithelial matrix cells are particularly prone to undergo irreversible atrophy, in this case due to thermal injury.

2. Cataract

During accommodation the eye focuses at near distance. The contraction of the ciliary muscle is transmitted to the lens via the zonules and the ICTL. After cataract surgery the accommodative status changes. In a pseudophakic eye with an artificial intraocular lens the zonules are intact, whereas in aphakic eyes the zonules have been digested and the lens is absent. Regardless of the surgical method, the immunoreaction for the HNK-1 epitope in the ICTL is equal to that seen in normal phakic eyes, even 16 years after cataract extraction.⁶³

A common problem after cataract extraction is formation of a secondary cataract, which is thought to result from proliferation and migration of residual lens epithelial cells.³ These metaplastic cells often acquire smooth muscle actin and produce abundant extracellular matrix that forms plaques on the lens capsule.⁶² Granular immunoreaction for the HNK-1 epitope is present in this extracellular matrix of secondary cataract (*Fig.5C*).⁶³ The fact that the HNK-1 epitope is likewise present in anterior subcapsular cataracts (*Fig.5D*), in which abnormal extracellular matrix is produced under an intact lens capsule, suggests that it is produced by the metaplastic lens epithelial cells.⁶³ As the HNK-1 epitope is associated with cell adhesion and migration, it is tempting to suggest that it might contribute to the pathogenesis of secondary cataracts, for example by aiding the migration or adhesion of the metaplastic lens cpithelial cells.

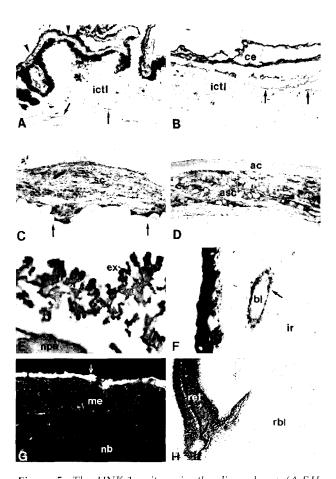


Figure 5. The HNK-1 epitope in the diseased eye (A-F,H immunoperoxidase staining, G immunofluorescence). A. The immunoreaction (arrows) is greatly diminished in the ICTL of an eye with absolute glaucoma as compared to normal amount in *Fig.3A.* Note that immunostaining persists in the non-pigmented ciliary epithelium (arrowheads). B. Likewise, in an eye after krypton laser cyclophotocoagulation, immunolabelling (arrows) is faint in the ICTL, but strong in the ciliary epithelia (ce). C. Granular immunoreaction for the HNK-1 epitope is seen in extracellular matrix of a plaque of secondary cataract (sc). The posterior lens capsule (arrows) is not labelled. D. Equal immunostaining is seen in an anterior subcapsular cataract (asc), beneath an intact anterior lens capsule (ac). E. Exfoliation material (ex) on the non-pigmented ciliary epithelium (npe) is immunoreactive for the HNK-1 epitope. F. Immunostaining is also found around some blood vessels (bl) of the iris (ir) in an eye with exfoliation syndrome. G. As in the early developing retina, the inner aspect (arrow) of medullary epithelial cords (me) and radial fibers react for the HNK-1 epitope in an intraocular medulloepithelioma. Note negative neuroblastic cells (nb). H. No labelling is seen within a retinoblastoma (rbl) that originates from the labelled retina (ret). Original magnifications: A x175, B x200, C x220, D x340, E x330, F x340, G x310, and H x70.

3. Exfoliation syndrome

Exfoliation syndrome is an intriguing disease in which whitish fibrillogranular material deposits throughout the anterior segment of the eye.⁴⁰ It is a major risk factor in cataract surgery. The origin of exfoliation material is obscure. Material resembling the ultrastructure of exfoliation fibers has been reported in various extrabulbar sites, pointing to the possibility that exfoliation syndrome may be a systemic condition.^{10,51,54,56,57} Latest evidence points toward abnormal metabolism of extracellular matrix components, in particular of basement membranes and elastic fibers.^{26,31,49,51} As mentioned, the HNK-1 epitope is associated with both elements. Indeed, intraocular deposits of exfoliation material are intensely labelled for the HNK-1 epitope (Fig.5E).⁶⁵ In exfoliation syndrome, the HNK-1 immunoreactivity in the ICTL of the ciliary body is as strong as in normal eyes, but it appears somewhat more granular.66 The reason for this was not evident by immunoelectron microscopy.67

Another intriguing finding is that, in all eyes with exfoliation syndrome, a granular immunoreaction for the HNK-1 epitope is present in the iris at the level of the thick laminated basement membrane of a subset of blood vessels (*Fig.5F*).⁶⁵ Exfoliation material has previously been detected in this location by electron microscopy,^{26,45} and the vasculopathy seems to have a definite association with exfoliation syndrome. In fact, the iris blood vessels become abnormal even before exfoliation deposits can be detected clinically or histopathologically in the eye.²⁰ In contrast, we have not detected immunoreactivity at the light microscopic level in the conjunctiva, an extraocular tissue in which exfoliation fibers have been claimed to be present.¹⁴

Because the HNK-1 epitope is associated with cell adhesion, it might bind together this intraocular multicomponent material and be responsible for its characteristic adhesiveness to intraocular surfaces. The HNK-1 epitope is an casy and reliable marker for exfoliation syndrome even in its early phase when the diagnosis is not immediately evident. It has been used to demonstrate that exfoliation syndrome is never truly monocular, since early changes are also present in the clinically healthy fellow eye.²⁰

4. Developmental anomalies

The glial and neuroepithelial cells that bear the HNK-1 epitope in the eye retain it under many pathologic conditions. The epitope has been successfully used to trace the derivatives of these cells in developmental ocular anomalies. It identifies gliotic retinal remnants in intraocular teratoma and optic nerve coloboma,^{22,24} a variable population of glial and neuroblastic cells in intraocular glioneuroma, medulloepithelioma and colobomatous cysts of the orbit,^{21,24} and the embryonal ciliary epithelium in medulloepithelioma (*Fig.5G*) and glioneuroma of the ciliary body,^{21,25}

5. Intraocular tumors

Even though the HNK-1 epitope is present throughout the normal adult and fetal retina, it is absent from retinoblastoma (*Fig.5H*), a tumor of the developing retina.^{19,42} Either this tumor originates from cells that do not express the epitope or it is lost during malignant transformation. Interestingly, retinoblastoma cells are loose and easily detatch to the vitreous, indicating poor adhesiveness.

The HNK-1 epitope is reportedly present in one fifth of primary uveal melanomas, mostly in large ones, and in about one fourth of their metastases.³⁹ In eyes with a melanoma of the ciliary body, the HNK-1 immunoreaction in the ICTL almost disappears near the tumor.⁶⁰ This locally decreased immunoreaction may be due to atrophy of the subepithelial matrix cells near the tumor, caused either by direct compression, decreased blood flow, or a combination of factors.

Future directions

Antibodies to the HNK-1 epitope are emerging as a very versatile tool in eye research. They have already revealed a new element in the human eye, the subepithelial matrix cells, which probably produce the epitope throughout life. They have also revealed phylogenetic changes in retinal antigenic profile not visible with other antibodies. Moreover, the HNK-1 epitope may be of pathogenetic significance in exfoliation syndrome and secondary cataract that are frequent and clinically important eye diseases.

In the future, we have to find out the physiologic role and function for the subepithelial matrix cells of the ICTL, as well as for the HNK-1 epitope in the ICTL, ciliary epithelia, and retina. In this regard, the species differences already mentioned may prove to be informative.

The most important step is to find out which molecules bear the HNK-1 epitope in various tissues, and which molecules may bind to it. In the retina, neural cell adhesion molecules may be responsible.²⁸ In the ICTL, tenascin and chondroitin sulphate proteoglycans have been ruled out.⁵⁹ Other possible candidates might be collagen types I and III, fibronectin and fibrillin that are present in this region.^{35,36,47,50} So far none of these molecules is known to bear this epitope, however. The epitope might also be part of integrins, which sometimes carry it.43 Laminin and amyloid P-component are known to have a binding site for the HNK-1 epitope,^{13,33} and the former is suggested to be one important extracellular component through which this cpitope may act.^{9,12,13} Obviously, many questions remain to be answered before the secrets of the HNK-1 epitope in the eye have been solved.

References

- Abo T and Balch CM: A differentiation antigen of human NK and K cells identified by a monoclonal antibody (HNK-1). J Immunol 127:1024-1029, 1981.
- Ando I and Tamaki K: HNK-1 antibody reacts with peripheral nerves and sweat glands in the skin. Br J Dermatol 113:175-178, 1985.
- 3. Apple DJ, Manalis N, Olson RJ and Kincaid MC: Intraocular lenses. Evolution, designs, complications, and pathology. Williams & Wilkins, USA, 327-334, 1989.
- Ariga T. Kohriyama T. Freddo L, et al: Characterization of sulfated glucuronic acid containing glycolipids reacting with IgM M-proteins in patients with neuropathy. J Biol Chem 262:848-853, 1987.
- 5. Arimatsu Y, Naegele JR and Barnstable CJ: Molecular markers of neuronal subpopulations in layers 4, 5, and 6 of cat primary visual cortex. J Neurosci 7:1250-1263, 1987.
- Bronner-Fraser M: Perturbation of cranial neural crest migration by the HNK-1 antibody. Dev Biol 123:321-331, 1987.
- Bunn PA Jr, Linnoila I, Minna JD, et al: Small cell lung cancer, endocrine cells of the fetal bronchus, and other neuroendocrine cells express the Leu-7 antigenic determinant present on natural killer cells. Blood 65:764-768, 1985.
- Chou DKH, Ilyas AA, Evans JE, et al. Structure of sulfated glucuronyl glycolipids in the nervous system reacting with HNK-t antibody and some IgM paraproteins in neuropathy. J Biol Chem 261:11717-11725, 1986.
- Dow KE, Mirski SEL, Roder JC and Riopelle RJ: Neuronal proteoglycans: biosynthesis and functional interaction with neurons in vitro. J Neurosci 8:3278-3289, 1988.
- Eagle RC, Font RL and Fine BS: The basement membrane exfoliation syndrome. Arch Ophthalmol 97:510-515, 1979.
- Effrench-Constant C, Miller RH, Kruse J, et al: Molecular specialization of astrocyte processes at nodes of Ranvier in rat optic nerve. J Cell Biol 102:844-852, 1986.
- Hall H, Liu L, Schahner M and Schmitz B: The L2/HNK-1 carbohydrate mediates adhesion of neural cells to laminin. Eur J Neurosci 5:34-42, 1993.
- Hall H, Vorherr T and Schachner M: Characterization of a 21 amino acid peptide sequence of the laminin G2 domain that is involved in HINK-1 carbohydrate binding and cell adhesion. Glycobiology 5:435-441, 1995.
- 14. Hietanen J, Uusitalo M, Tarkkanen A and Kivelä T: Lectin and immunohistochemical comparison of glycoconjugates in the conjunctiva of patients with and without exfoliation syndrome. Br J Ophthalmol 79:467-472, 1995.
- Hogan MJ, Alvarado JA and Weddel JE: Histology of the human cyc. An atlas and textbook. WB Saunders, Philadelphia, PA, 260-319, 1971.
- Ilyas AA, Dobersen MJ, Willison HJ and Quarles RH: Mouse monoclonal and rabbit polyclonal antibodies prepared to human myclin-associated glycoproteins also react with glycosphingolipids of peripheral nerve. J Neuroimmunol 12:99-106, 1986.
- Inuzuka T, Quarles RH, Noronha AB, Dobersen MJ and Brady RO: A human lymphocyte antigen is shared with a group of glycoproteins in peripheral nerve. Neurosci Lett 51:105-111, 1984.
- Keilhauer G, Faissner A and Schachner M: Differential inhibition of neurone-neurone, neurone-astrocyte and astrocyte-astrocyte adhesion by L1, L2 and N-CAM antibodies. Nature 316:728-730, 1985.

- Kivelä T: Expression of the HNK-1 carbohydrate epitope in human retina and retinoblastoma. An immunohistochemical study with the anti-Leu-7 monoclonal antibody. Virchows Arch A Pathol Anat Histopathol 410:139-146, 1986.
- Kivelä T. Hietanen J and Uusitalo M: Exfoliation syndrome in one eye only - is it truly unilateral? Klin Monatsbl Augenheilkd 206:Suppl 2:5, 1995.
- 21. Kivelä T, Kauniskangas L, Miettinen P and Tarkkanen A: Glioneuroma associated with colobomatous dysplasia of the anterior uvea and retina. A case simulating medulloepithelioma. Ophthalmol 96:1799-1808, 1989.
- 22. Kivelä T. Merenmies L. Ilveskoski I and Tarkkanen A: Congenital intraocular teratoma. Ophthalmol 100:782-791, 1993.
- 23. Kivelä T, Puska P. Raitta C, Immonen I and Tarkkanen A: Clinically successful contact transcleral krypton laser cyclophotocoagulation. Arch Ophthalmol 113:1447-1453, 1995.
- 24. Kivelä T, Salonen R and Paetau A: Hydrolethalus: a midline malformation syndrome with optic nerve coloboma and hypoplasia. Acta Neuropathol 91:511-518, 1996.
- Kivelä T and Tarkkanen A: Recurrent medulloepithelioma of the ciliary body. Immunohistochemical characteristics. Ophthalmol 95:1565-1575, 1988.
- Konstas AG, Marshall GE and Lee WR: Immunogold localisation of laminin in normal and exfoliative iris. Br J Ophthalmol 74:450-457, 1990.
- Kruse J, Keilhauer G, Faissner A, et al.: The J1 glycoprotein a novel nervous system cell adhesion molecule of the L2/HNK-1 family. Nature 316:146-148, 1985.
- Kruse J, Mailhammer R, Wernecke II, et al: Neural cell adhesion molecules and myelin-associated glycoprotein share a common carbohydrate moiety recognized by monoclonal antibody L2 and HNK-1. Nature 311:153-155, 1984.
- 29. Künemund V, Jungalwala FB, Fischer G, et al: The L2/HNK-1 carbohydrate of neural cell adhesion molecules is involved in cell interactions. J Cell Biol 106:213-223, 1988.
- 30. Lauweryns JM and van Ranst L: Leu-7 immunoreactivity in human, monkey, and pig bronchopulmonary neuroepithelial bodies and neuroendocrine cells. J Histochem Cytochem 35:687-691, 1987.
- 31. Li ZY, Streeten BW and Wallace RN: Association of elastin with pseudoexfoliative material: an immunoelectron microscopic study. Curr Eye Res 7:1163-1172, 1988.
- Lipinski M, Braham K, Caillaud JM, et al: HNK-1 antibody detects an antigen expressed on neuroectodermal cells. J Exp Med 158:1775-1780, 1983.
- 33. Loveless RW, Floyd-O'Sullivan G, Raynes JG, et al: Human serum amyloid P is a multispecific adhesive protein whose ligands include 6-phosphorylated mannose and the 3-sulphated saccharides galactose, N-acetylgalactosamine and glucuronic acid. EMBO J 11:813-819, 1992.
- 34. *Margolis RK*, *Ripellino JA*, *Goossen B*, *et al*: Occurrence of the HNK-1 epitope (3-sulfoglucuronic acid) in PC12 pheocromocytoma cells, chromaffin granule membranes, and chondroitin sulfate proteoglycans. Biochem Biophys Res Commun 145:1142-1148, 1987.
- 35. Marshall GE, Konstas AGP, Abraham S and Lee WR: Extracellular matrix in aged human ciliary body: an immunoelectron microscope study. Invest Ophthalmol Vis Sci 33:2546-2560, 1992.
- Marshall GE, Konstas AGP and Lee WR: Collagens in ocular tissues. Br J Ophthalmol 77:515-524, 1993.
- McGarry RC, Helfand SL, Quarles RH and Roder JC: Recognition of myclin-associated glycoprotein by the monoclonal antibody HNK-1. Nature 306:376-378, 1983.

- 38. Merkouri E and Matsas R: Monoclonal antibody BM89 recognizes a novel cell surface glycoprotein of the L2/IINK-1 family in the developing mammalian nervous system. Neuroscience 50:53-68, 1992.
- Mooy CM, Luyten GPM, De Jong PTVM, et al: Neural cell adhesion molecule distribution in primary and metastatic uveal melanoma. Human Pathol 11:1185-1190, 1995.
- Morrison JC and Green WR: Light microscopy of the exfoliation syndrome. Acta Ophthalmol 66:5-27, 1988.
- Nobile-Orazio E, Hays AP, Latov N, et al: Specificity of mouse and human monoclonal antibodies to myelin-associated glycoprotein. Neurology 34:1336-1342, 1984.
- Perentes E. Herbort CP, Rubinstein LJ, et al: Immunohistochemical characterization of human retinoblastomas in situ with multiple markers. Am J Ophthalmol 103:647-658, 1987.
- 43. Pesheva P, Horwitz AF and Schachner M: Integrin, the cell surface receptor for fibronectin and laminin, expresses the L2/HNK-1 and L3 carbohydrate structures shared by adhesion molecules. Neurosci Lett 83:303-306, 1987.
- 44. *Qi Y, Wallace RN, Hoepner JA and Streeten BW:* HNK-1 epitopes in normal and pseudoexfoliation (PSX) lens capsules by TEM. Invest Ophthalmol Vis Sci 36:S329, 1995.
- 45. *Ringvold A:* Light and electron microscopy of the wall of iris vessels in eyes with and without exfoliation syndrome (pseudoexfoliation of the lens capsule). Virchows Arch A Pathol Anat Histopathol 349:1-9, 1970.
- 46. *Riopelle RJ, McGarry RC and Roder JC:* Adhesion properties of a neuronal epitope recognized by the monoclonal antibody HNK-1. Brain Res 367:20-25, 1986.
- Rittig M, Lütjen-Drecoll F, Rauterberg J. et al: Type-VI collagen in the human iris and ciliary body. Cell Tissue Res 259:305-312, 1990.
- Sadaghiani B and Vielkind JR: Distribution and migration pathways of HNK-1-immunoreactive neural crest cells in teleost fish embryos. Development 110:197-209, 1990.
- 49. Schlötzer-Schrehardt U, Dörfler S and Naumann GOH: Immunohistochemical localization of basement membrane components in pseudoexfoliation material of the lens capsule. Curr Eye Res 11:343-355, 1992.
- Schlötzer-Schrehardt U, Endress K and Naumann GOH: Immunohistochemical localization of fibrillin in the anterior segment of pseudocxfoliation cycs. Vis Res 36Suppl:S60, 1996.
- Schlötzer-Schrehardt U, Kühle M and Naumann GOH: Electron-microscopic identification of pseudoexfoliation material in extrabulbar tissue. Arch Ophthalmol 109:565-570, 1991.
- Schuller-Petrovic S, Gebhart W, Lassmann H, et al: A shared antigenic determinant between natural killer cells and nervous tissue. Nature 306:179-181, 1983.
- 53. Shioda Y. Nagura II, Tsutsumi Y, et al: Distribution of Leu 7 (IINK-1) antigen in human digestive organs: an immunohistochemical study with monoclonal antibody. Histochem J 16:843-854, 1984.

- 54. Speakman JS and Ghosh M: The conjunctiva in senile lens exfoliation. Arch Ophthalmol 94:1757-1759, 1976.
- 55. Streeten BW: Ciliary body. In: Duane TD, Jaeger EA (eds.), Biomedical Foundations of Ophthalmology. Vol 1, Philadelphia. PA: Harper & Row, ch 13:1-28, 1985.
- Streeten BW, Dark AJ, Wallace RN, et al: Pseudoexfoliative fibrillopathy in the skin of patients with ocular pseudoexfoliation. Am J Ophthalmol 110:490-499, 1990.
- 57. *Streeten BW, Li ZY, Wallace RN, et al*: Pseudocxfoliative fibrillopathy in visceral organs of a patient with pseudoexfoliation syndrome. Arch Ophthalmol 110:1757-1762, 1992.
- 58. Ticker GC, Aoyama H, Lipinski M, et al: Identical reactivity of monoclonal antibodies HNK-1 and NC-1: conservation in vertebrates on cells derived from the neural primordium and on some leucocytes. Cell Differ 14:223-230, 1984.
- 59. Uusitalo M: Immunohistochemical localization of chondroitin sulfate proteoglycan and tenascin in the human eye compared with the HNK-1 epitope. Graefes Arch Clin Exp Ophthalmol 232:657-665, 1994.
- 60. Uusitalo M: The HNK-1 carbohydrate epitope in the anterior segment of the eye. The inner connective tissue layer of the human ciliary body as a distinct element. Medical Doctoral Thesis. University of Helsinki, 1995. Summary published in Acta Ophthalmol Scand 73:363, 1995.
- 61. *Uusitalo M and Kivelä T:* Differential distribution of the HNK-1 carbohydrate epitope in the vertebrate retina. Curr Eye Res 13:697-704, 1994.
- 62. Uusitalo M and Kivelä T: Cell types of secondary cataract an immunohistochemical analysis with antibodies to cytoskeletal elements and macrophages. Graefes Arch Clin Exp Ophthalmol, in press.
- Uusitalo M and Kivelä T: The HNK-1 epitope in the pseudophakic and aphakic eye and secondary cataract. Acta Ophthalmol, in press.
- 64. Uusitato M, Kivelä T and Tarkkanen A: Identification of a novel element in the human eye: The inner connective tissue layer of the eiliary body characterized with antibodies to the HNK-1 epitope. Invest Ophthalmol Vis Sci 34:2372-2381, 1993.
- Uusitalo M, Kivelä T and Tarkkanen A: Immunoreactivity of exfoliation material for the cell adhesion-related HNK-1 carbohydrate epitope. Arch Ophthalmol 111:1419-1423, 1993.
- 66. Uusitalo M, Kivelä T and Tarkkanen A: The HNK-1 epitope in the inner connective tissue layer of the human ciliary body in exfoliation syndrome and various types of glaucoma. Graefes Arch Clin Exp Ophthalmol 232:8-15, 1994.
- 67. Uusitalo M. Schlötzer-Schrehardt U, et al: Immunoelectron microscopic localization of the HNK-1 epitope in the human ciliary body. Vis Res 36, Suppl:S73, 1996.
- Vincent M and Thiery J-P: A cell surface marker for neural crest and placodal cells: Further evolution in peripheral and central nervous system. Dev Biol 103:468-481, 1984.